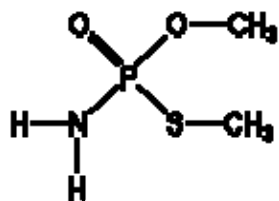


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ACUTE EXPOSURE GUIDELINE LEVELS (AEGLs)
FOR
METHAMIDOPHOS
(CAS Reg. No. 10265-92-6)



PROPOSED

PREFACE

Under the authority of the Federal Advisory Committee Act (FACA) P. L. 92-463 of 1972, the National Advisory Committee for Acute Exposure Guideline Levels for Hazardous Substances (NAC/AEGL Committee) has been established to identify, review and interpret relevant toxicologic and other scientific data and develop AEGLs for high priority, acutely toxic chemicals.

AEGLs represent threshold exposure limits for the general public and are applicable to emergency exposure periods ranging from 10 minutes to 8 hours. Three levels – AEGL-1, AEGL-2 and AEGL-3 – are developed for each of five exposure periods (10 and 30 minutes, 1 hour, 4 hours, and 8 hours) and are distinguished by varying degrees of severity of toxic effects. The three AEGLs are defined as follows:

AEGL-1 is the airborne concentration (expressed as parts per million or milligrams per cubic meter [ppm or mg/m³]) of a substance above which it is predicted that the general population, including susceptible individuals, could experience notable discomfort, irritation, or certain asymptomatic, non-sensory effects. However, the effects are not disabling and are transient and reversible upon cessation of exposure.

AEGL-2 is the airborne concentration (expressed as ppm or mg/m³) of a substance above which it is predicted that the general population, including susceptible individuals, could experience irreversible or other serious, long-lasting adverse health effects or an impaired ability to escape.

AEGL-3 is the airborne concentration (expressed as ppm or mg/m³) of a substance above which it is predicted that the general population, including susceptible individuals, could experience life-threatening health effects or death.

Airborne concentrations below the AEGL-1 represent exposure levels that could produce mild and progressively increasing but transient and nondisabling odor, taste, and sensory irritation or certain asymptomatic, non-sensory effects. With increasing airborne concentrations above each AEGL, there is a progressive increase in the likelihood of occurrence and the severity of effects described for each corresponding AEGL. Although the AEGL values represent threshold levels for the general public, including susceptible subpopulations, such as infants, children, the elderly, persons with asthma, and those with other illnesses, it is recognized that individuals, subject to unique or idiosyncratic responses, could experience the effects described at concentrations below the corresponding AEGL.

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SUMMARY

Technical methamidophos (CAS No. 10265-92-6) is a colorless to white crystalline solid organophosphate pesticide with a strong mercaptan-like odor. The solid material has a low vapor pressure and is readily soluble in water. Commercially, methamidophos is available as an aqueous liquid or emulsifiable concentrate. It is applied as a spray or by sprinkler irrigation to a variety of food crops.

Organophosphate pesticides such as methamidophos are neurotoxic in that they are inhibitors of cholinesterase enzymes. Inhibition of acetylcholinesterase, responsible for termination of the biological activity of the neurotransmitter acetylcholine at various nerve endings, results in sustained stimulation of electrical activity. Depending on concentrations administered, cholinergic signs following acute exposure of rats to methamidophos may include salivation, lacrimation, decreased activity, muscle fasciculation, ataxia, gasping, and tremors. In humans, inhibition of erythrocyte acetylcholinesterase activity, the form found in human erythrocytes, is used as a biomarker of exposure and effects of organophosphate pesticides. No inhalation studies involving human subjects were located.

No human inhalation data relevant to derivation of AEGL values were found. Acute inhalation toxicity studies with rats were available from two different laboratories. Both studies administered the test compound as liquid aerosols. Results of 4-hour inhalation LC₅₀ values differed by a factor of approximately 3 between the two laboratories. In both studies, nominal concentrations were a poor indicator of analytical concentrations, indicating the difficulty in maintaining aerosols at these concentrations. A 6-hour inhalation study addressed non-lethal effects.

The study of Pauluhn (1986) was used for derivation of AEGL values. This study showed an adequate concentration-response curve, and cholinesterase activity was measured in plasma and erythrocytes. No clinical signs were observed following exposure of rats to 11.4 or 24.3 mg/m³ for 4 hours or following exposure to 1.4 or 5.4 mg/m³ for 6 hours. Clinical signs were observed at the next higher concentrations, 45.0 mg/m³ for 4 hours and 33.1 mg/m³ for 6 hours. At 11.4 mg/m³, plasma cholinesterase activity was inhibited by approximately 50%, but erythrocyte acetylcholinesterase activity was unaffected. At 24.3 and 45.0 mg/m³, plasma cholinesterase activity was 36 and 13% of the control value, respectively, and erythrocyte cholinesterase activity was 92 and 70% of the control value, respectively.

The exposure of rats to 24.3 mg/m³ for 4-hours (Pauluhn 1986) was chosen as the point of departure for the AEGL-1. At this concentration, plasma and erythrocyte cholinesterase activity were depressed by 64 and 8%, respectively. Although there were no clinical signs at this concentration and the lower concentration of 11.4 mg/m³, exposure to the next higher concentration of 45.0 mg/m³ resulted in clinical signs consistent with the definition of the AEGL-2. Because of the apparent difficulty in maintaining liquid aerosols at these concentrations and the disparate data between the two available studies, the 24.3 mg/m³ value was divided by a data base modifying factor of 2. Methamidophos is rapidly metabolized and excreted in rats and humans as indicated by oral dosing studies (Moser 1999; Garofalo et al. 1973). Therefore, an interspecies uncertainty factor of 3 was applied. Infants and juveniles may

1 be more sensitive to organophosphate pesticides than adults, but an acute oral dosing study with
2 adult and juvenile rats failed to show differences in sensitivity to methamidophos (Moser 1999).
3 Based on repeat-dose oral studies with adult and juvenile rats, the U.S. EPA (2006a) identified
4 juveniles as being twice as sensitive as adults. Because there were no differences in sensitivity
5 between adult and juvenile rats in the acute oral dosing study, and in keeping with intraspecies
6 uncertainty factors derived for other organophosphate pesticides that did not show differences in
7 sensitivity between adult and juvenile rats, an intraspecies uncertainty factor of 3 was considered
8 adequate. The total uncertainty factor is 10. The 4-hour 24.3 mg/m³ value was divided by a
9 total modifying/uncertainty factor of 20 (2x10). In the absence of reliable time-scaling
10 information, the resulting 4-hour value of 1.2 mg/m³ was time-scaled ($C^n \times t = k$) using n values
11 of 3 and 1 for shorter and longer exposure durations, respectively (NRC 2001). Because of
12 uncertainty in scaling from 4 hours to 10 minutes, the 10-minute value was set equal to the 30-
13 minute value.

14
15 The 4-hour exposure of rats to 45.0 mg/m³ in the study by Pauluhn (1986) was chosen as
16 the point of departure for the AEGL-2. Clinical signs consisted of tremor, staggering, and
17 reduced motility. Plasma and erythrocyte cholinesterase activity were 13 and 70% of control.
18 Mortality of 30% occurred at the next higher exposure of 195.5 mg/m³. Because of the apparent
19 difficulty in maintaining liquid aerosols at these concentrations and the disparity in data between
20 the two available studies, the 45.0 mg/m³ value was divided by a data base modifying factor of 2.
21 Methamidophos is rapidly metabolized and excreted in rats and humans as indicated by oral
22 dosing studies (Moser 1999; Garofalo et al. 1973). Therefore, an interspecies uncertainty factor
23 of 3 was applied. Infants and juveniles may be more sensitive to organophosphate pesticides
24 than adults. An acute oral dosing study with adult and juvenile rats failed to show age-related
25 differences in sensitivity to methamidophos (Moser 1999). Based on repeat-dose oral studies
26 with adult and juvenile rats, the U.S. EPA (2006a) identified an uncertainty factor of 2 to protect
27 the sensitive population of children. Because there were no differences in sensitivity between
28 adult and juvenile rats in the acute oral dosing study, and in keeping with intraspecies
29 uncertainty factors derived for other organophosphate pesticides that did not show differences in
30 sensitivity between adult and juvenile rats, an intraspecies uncertainty factor of 3 was considered
31 adequate. The total uncertainty factor is 10. The 4-hour 45.0 mg/m³ value was divided by a
32 total modifying/uncertainty factor of 20 (2x10). In the absence of reliable time-scaling
33 information, the resulting 4-hour value of 2.3 mg/m³ was time-scaled ($C^n \times t = k$) using n values
34 of 3 and 1 for shorter and longer exposure durations, respectively (NRC 2001). Because of
35 uncertainty in scaling from 4 hours to 10 minutes, the 10-minute value was set equal to the 30-
36 minute value.

37
38 The 4-hour exposures of rats to methamidophos delivered as a liquid aerosol at
39 concentrations of 11.4 to 350.3 mg/m³ in the study of Pauluhn (1986) were used to develop
40 AEGL-3 values. The threshold for lethality was calculated using U.S. EPA's Benchmark
41 Concentration (BMC) program (V2.8). The BMCL₀₅ was 56.27 mg/m³, and the BMC₀₁ was
42 101.54 mg/m³ (see Appendix C for program output). Although the lower value, in this case the
43 BMCL₀₅ of 56.27 mg/m³, is generally chosen as the threshold for mortality in developing
44 AEGL-3 values, this value was considered an artifact of the large gap between tested
45 concentrations of 45.0 and 195.5 mg/m³. The 56.27 mg/m³ value is also close to the 45.0 mg/m³
46 value that resulted in effects considered consistent with the definition of AEGL-2. The 4-hour
47 BMC₀₁ of 101.54 mg/m³ for methamidophos delivered as a liquid aerosol was considered the

1 threshold for mortality in rats. Because of the apparent difficulty in maintaining liquid aerosols
 2 at these concentrations and the disparity in data between the two available studies, the 101.54
 3 mg/m³ value was divided by a data base modifying factor of 2. Methamidophos is rapidly
 4 metabolized and excreted in rats and humans as indicated by oral dosing studies (Moser 1999;
 5 Garofalo et al. 1973). Therefore, an interspecies uncertainty factor of 3 was applied. Infants and
 6 juveniles may be more sensitive to organophosphate pesticides than adults. An acute oral dosing
 7 study with adult and juvenile rats failed to show age-related differences in sensitivity to
 8 methamidophos (Moser 1999). Based on repeat-dose oral studies with adult and juvenile rats,
 9 the U.S. EPA (2006a) identified an uncertainty factor of 2 to protect children. Because there
 10 were no differences in sensitivity between adult and juvenile rats in the acute oral dosing study,
 11 and in keeping with intraspecies uncertainty factors derived for other organophosphate pesticides
 12 that did not show differences in sensitivity between adult and juvenile rats, an intraspecies
 13 uncertainty factor of 3 was considered adequate. The total uncertainty factor is 10. The 4-hour
 14 101.54 mg/m³ value was divided by a total modifying/uncertainty factor of 20 (2x10). In the
 15 absence of reliable time-scaling information, the resulting 4-hour value of 5.01 mg/m³ was time-
 16 scaled ($C^n \times t = k$) using n values of 3 and 1 for shorter and longer exposure durations,
 17 respectively (NRC 2001). Because of uncertainty in scaling from 4 hours to 10 minutes, the 10-
 18 minute value was set equal to the 30-minute value.

19
 20 The calculated values are supported by tested concentrations in a subchronic study with
 21 rats (Pauluhn and Cole 1988). No treatment related effects were observed in rats inhaling 1.1
 22 mg/m³ for 13 weeks. At 5.4 mg/m³, erythrocyte and brain cholinesterase activities were
 23 inhibited by <30% throughout the treatment period. Rats inhaling 23.1 mg/m³ showed clinical
 24 signs consistent with cholinesterase activity inhibition, but no deaths were reported. At study
 25 termination, brain acetylcholinesterase activity was inhibited by 45-47%.

26
 27 The calculated values are listed in the table below.

28

| TABLE ES 1. Summary of AEGL Values for Methamidophos | | | | | | |
|--|-----------------------|-----------------------|-----------------------|-----------------------|------------------------|---|
| Classification | 10-min | 30-min | 1-h | 4-h | 8-h | Endpoint (Reference) |
| AEGL-1 (Nondisabling) | 2.4 mg/m ³ | 2.4 mg/m ³ | 1.9 mg/m ³ | 1.2 mg/m ³ | 0.61 mg/m ³ | No clinical signs – rat (Pauluhn 1986)* |
| AEGL-2 (Disabling) | 4.5 mg/m ³ | 4.5 mg/m ³ | 3.6 mg/m ³ | 2.3 mg/m ³ | 1.1 mg/m ³ | Clinical signs of tremor, reduced motility – rat (Pauluhn 1986)* |
| AEGL-3 (Lethal) | 10 mg/m ³ | 10 mg/m ³ | 8.1 mg/m ³ | 5.1 mg/m ³ | 2.5 mg/m ³ | 4-hour BMC ₀₁ – rat (Pauluhn 1986) |

29 *At the AEGL-1 point of departure, 24.3 mg/m³ for 4 hours, plasma cholinesterase activity was 36% of the control
 30 value and erythrocyte acetylcholinesterase activity was 92% of the control value. At the AEGL-2 point of
 31 departure, 45.0 mg/m³ for 4 hours, plasma cholinesterase activity was 13% of the control value and erythrocyte
 32 acetylcholinesterase activity was 70% of the control value.

33 34 1. INTRODUCTION

35
 36 Technical methamidophos (CAS No. 10265-92-6) is a colorless to white crystalline solid
 37 organophosphate pesticide with a strong mercaptan-like odor (U.S. EPA 2006b). The presence
 38 of both an amine group and a sulfur atom single-bonded to phosphorus place it in the class

1 phosphoramidothioates. The solid material has a low vapor pressure and is readily soluble in
 2 water. Additional chemical and physical properties are listed in Table 1.

3
 4 In 1972, methamidophos was registered in the United States under the trade name
 5 Monitor[®]. The application of methamidophos as an insecticide/acaricide is restricted to the food
 6 crops cotton, potatoes, and tomatoes and on the non-food crop alfalfa, grown for seed.
 7 Registered formulations include the technical grade liquid containing 60-72% a.i. (active
 8 ingredient) and an emulsifiable concentrate containing 40% a.i. Methamidophos is applied as a
 9 spray or by sprinkler irrigation (U.S. EPA 2006b).

10
 11 Methamidophos is manufactured commercially by the reaction of sodium methyl
 12 phosphoroamidothioate and dimethyl sulfate or by the isomerization of O,O-
 13 dimethylthiophosphamidate (HSDB 2004). According to U.S. EPA (2006b), approximately
 14 640,000 pounds of a.i. are used annually in the United States. Most usage is on potatoes. World
 15 production figures were not available.

16
 17 Methamidophos is an environmental degradate of the organophosphate pesticide
 18 acephate (C₄H₁₀NO₃PS) and may be a metabolite of acephate following oral dosing of rats
 19 (Singh 1985; IPCS 2002). Acephate is the N-acetyl derivative of methamidophos.

20

| TABLE 1. Chemical and Physical Properties | | |
|---|--|---------------------------------------|
| Parameter | Value | Reference |
| Synonyms | O,S-Dimethyl phosphoramidothioate, Bayer 71628, Ortho 9006, Monitor, Tamaron | O'Neil et al. 2001a |
| Chemical formula | C ₂ H ₈ NO ₂ PS | O'Neil et al. 2001a |
| Molecular weight | 141.1 | U.S. EPA 2006b |
| CAS Reg. No. | 10265-92-6 | O'Neil et al. 2001a |
| Physical state | white crystalline solid thick clear liquid (75% technical solution) | U.S. EPA 2006b Sangha 1983 |
| Solubility in water | 2 kg/L | IPCS 1993 |
| Vapor pressure | 1.7 x 10 ⁻⁵ mm Hg 3 x 10 ⁻⁴ mm Hg at 30°C | U.S. EPA 2006b O'Neil et al. 2001a |
| Vapor density, saturated (air =1) | 1.7 mg/m ³ at 30°C | Pauluhn 1986 |
| Liquid density (water =1) | 1.3 at 44.5°C | O'Neil et al. 2001a |
| Melting point | 44.5°C (pure); 37-39 °C (technical) | IPCS 1993 |
| Boiling point | thermally unstable | IPCS 1993 |
| Flammability limits in air | Not available | |
| Conversion factors | 1 ppm = 5.77 mg/m ³ 1 mg/m ³ = 0.17 ppm | Calculated |

21
 22
 23 **2. HUMAN TOXICITY DATA**

24
 25 No inhalation studies other than reports of accidental exposures were located. These
 26 reports lacked information on concentration and exposure duration. Symptoms of cholinesterase
 27 activity inhibition have been observed following ingestion of food containing methamidophos
 28 residue (HSDB 2004). Methamidophos has been implicated in causing organophosphate-

1 induced delayed neurotoxicity in humans; however, these incidents involved accidental or
2 suicidal exposure to excessively high levels (Costa 2008).

3
4 In a clinical study, seven male and seven female volunteers, ages 21-48 years, were
5 administered combinations of methamidophos (Monitor) and acephate (Orthene) at doses of 0.1,
6 0.2, 0.3, or 0.4 mg/kg/day in a gelatin capsule containing corn oil (Garofalo et al. 1973;
7 reviewed in U.S. EPA 1988; 2000a).¹ A group of three males and three females received
8 methamidophos/acephate in a 1:9 ratio (equivalent to 0.01, 0.02, 0.03, or 0.04 mg/kg/day
9 methamidophos). A second group of two males and two females was given 0.1 or 0.2 mg/kg/day
10 of a 1:4 mixture (equivalent to 0.02 or 0.04 mg/kg/day methamidophos). The controls (two
11 males and two females) received gelatin capsules containing corn oil. Volunteers were blind to
12 the dose administered. The daily dose was administered in three equally divided doses. Dosing
13 was continued over a 37-73 day period (maximum administration of 21 days) until plasma
14 cholinesterase activity inhibition reached two standard deviations below mean pretest activity for
15 two successive cholinesterase assays. These mixtures had no effect on erythrocyte
16 cholinesterase activity, hematology, blood chemistry, blood pressure, pulse rate, pupil size, light
17 reflex, eye accommodation, chest sound, muscle tone, knee jerk, tongue tremor, or finger tremor.
18 After 16 days, plasma cholinesterase activity was significantly inhibited in all subjects in the 1:4
19 ratio group that received 0.2 mg/kg/day of the mixture. Significant plasma cholinesterase
20 activity inhibition occurred after 21 days of dosing only in males in the group receiving the 1:9
21 ratio at 0.3 mg/kg/day. In the group that received 0.4 mg/kg/day (1:9 ratio), two of three females
22 showed significant plasma cholinesterase activity inhibition after 10 days of dosing. Plasma
23 cholinesterase activity returned to pretest values during a 7-day recovery period. Pre-test
24 erythrocyte acetylcholinesterase activity values were similar in male and female volunteers. Pre-
25 test plasma cholinesterase activity in females was approximately one-half of that in males.
26

27 The U.S. EPA relies in part on data from studies in which adult human subjects were
28 intentionally exposed to a pesticide. The human oral dosing studies, contained in the Pesticide
29 Handlers Exposure Database, “have been reviewed by the Agency and found on the basis of
30 available evidence to have been neither fundamentally unethical nor significantly deficient
31 relative to standards of ethical research conduct prevailing when they were conducted” (U.S.
32 EPA 2008).

33 34 **3. ANIMAL TOXICITY DATA**

35
36 Using standard protocols, methamidophos was tested for acute oral and dermal toxicity
37 and skin irritation and sensitization (U.S. EPA 2006b). The acute oral LD₅₀ was 15.6 mg/kg in
38 male rats and 13.0 mg/kg in female rats. The acute dermal toxicity in rabbits was 118 mg/kg.
39 Instillation into the eye resulted in corneal opacity and death of one of six rabbits (dose not
40 reported). Methamidophos was moderately irritating to the eyes and mildly irritating to the skin
41 of rabbits. Methamidophos was not a skin sensitizer in guinea pigs.
42

43 **3.1. Acute Toxicity**

44
45 All acute studies were conducted with rats. These studies are summarized in Table 2.

¹ The oral LC₅₀ values in male and female rats for acephate and methamidophos are 700 mg/kg (O’Neil et al. 2001b) and 14 mg/kg (U.S. EPA 2006b), respectively.

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Groups of ten male and ten female young-adult Sprague-Dawley rats inhaled an aerosol of technical methamidophos (purity 75.1%), head-only, for 1 hour (Sangha 1983). Liquid aerosol atmospheres were generated by pumping the test material through a fine nozzle and then mixing the spray with compressed, filtered and dried air. The compressed air finely atomized the test material. A constant air flow was maintained through the exposure chamber. Particles, collected on a cascade impactor averaged 1.1-2.1 μ ; 90% of particles were $<5 \mu$. Atmospheres were measured by collecting samples on filters and analyzing by gas chromatography. Cascade impactor sample values correlated better with nominal concentrations than filter samples and therefore were the basis for concentration measurements. Male rats were exposed to analytical concentrations of 160, 163, 253, or 319 mg/m^3 and female rats were exposed to analytical concentrations of 60, 160, 168, 196, 259, or 319 mg/m^3 . Nominal concentrations which ranged from 550-1390 mg/m^3 did not correlate with analytical concentrations. Control groups were run with most exposures and were exposed to room air. During exposure and for up to 5 days post-exposure, all methamidophos-exposed rats showed cholinergic signs including salivation, lacrimation, decreased activity, muscle fasciculation, ataxia, gasping, tremors, tearing, and rhinorrhea. Deaths occurred either during exposure or within 5 days post-exposure. For male rats that inhaled 160, 163, 253, or 319 mg/m^3 , mortality was 3/10, 1/10, 5/10, and 8/10, respectively. For female rats that inhaled 60, 160, 168, 196, 259, or 319 mg/m^3 , mortality was 1/10, 7/10, 5/10, 5/10, and 9/10, respectively. The author calculated 1-hour LC_{50} values of 377 mg/m^3 (95% confidence limits of 301 to 502 mg/m^3) for male rats and 241 mg/m^3 (95% confidence limits of 205 to 280 mg/m^3) for female rats. From post-exposure days 2-7, mean body weights for both sexes were reduced compared to controls. Congested nasal passages, congested lungs, and congested cervical lymph nodes were observed in many of the rats that died.

TABLE 2. Acute Toxicity of Methamidophos Liquid Aerosol to Rats

| Concentration (mg/m^3) | Exposure Duration | Effect/ LC_{50} (mg/m^3) | Reference |
|---|-------------------|--|-------------|
| 60 160 163 168 196 253 259 319 | 1 hour | Clinical signs, all exposures; Mortality: 1/10 females ^a Mortality: 3/10 males; 7/10 females Mortality: 1/10 males Mortality: 0/10 females Mortality: 5/10 females Mortality: 8/10 males Mortality: 5/10 males; 5/10 females Mortality: 9/10 females Calculated LC_{50} (males): 377 mg/m^3 Calculated LC_{50} (females): 241 mg/m^3 | Sangha 1983 |
| 19 33 56 57 63 83 173 | 4 hours | Cholinergic signs at all concentrations Mortality: no mortality (0/20) Mortality: 1/10 males; 0/10 females Mortality: 5/10 males; 4/10 females Mortality: 3/10 males; 4/10 females Mortality: 6/10 females Mortality: 8/10 males; 5/10 females Mortality: 10/10 females Calculated LC_{50} (males): 63.2 mg/m^3 Calculated LC_{50} (females): 76.5 mg/m^3 | Sangha 1984 |

| | | | |
|---|------------------------------|--|--------------|
| 11.4 24.3 45.0 195.5 241.7 350.3 | 4 hours | No clinical signs, no mortality No clinical signs, no mortality Tremor, reduced motility, no mortality Tremor, additional clinical signs; mortality: 1/5 males, 2/5 females Tremor, additional clinical signs; mortality: 3/5 males, 5/5 females Tremor, additional clinical signs; mortality: 5/5 males, 4/5 females Calculated LC ₅₀ : 213 mg/m ³ | Pauluhn 1986 |
| 1.4 5.4 33.1 | 6 hours, 5 days ^b | No clinical signs No clinical signs Slight tremor following exposure | Pauluhn 1986 |

^a At some concentrations, only one sex was tested.

^b Clinical signs were recorded after one 6-hour exposure.

1
2 Groups of ten male and ten female young adult Sprague Dawley rats inhaled a liquid
3 aerosol of technical methamidophos (a.i. 70.5%), head only for four hours (Sangha 1984). The
4 protocol and generation and measurement of the test atmospheres followed the same methods as
5 in the 1-hour study described above. Particles, collected on a cascade impactor ranged from
6 0.13-1.0 μ and averaged 0.53 μ . Male and female rats inhaled analytical concentrations of 19,
7 33, 56, 57, or 83 mg/m³. Additional groups of females were exposed to 63 or 173 mg/m³.
8 Mortalities occurred within five days postexposure. All rats exhibited clinical signs including
9 salivation, lacrimation, muscle fasciculations, tremors, decreased activity, pilo erection, and
10 hypothermia. Ocular and nasal irritation were observed on occasion. Body weight was
11 decreased in some groups up to day 14. Mortality in male rats inhaling 19, 33, 56, 57, or 83
12 mg/m³ was 0/10, 1/10, 5/10, 3/10, and 8/10, respectively. Mortality in female rats inhaling 19,
13 33, 56, 57, 63, 83, or 173 mg/m³ was 0/10, 0/10, 4/10, 4/10, 6/10, 5/10, and 10/10, respectively.
14 The 4-hour LC₅₀ for male rats was 63.2 mg/m³ (52-79 mg/m³), and the 4-hour LC₅₀ for female
15 rats was 76.5 mg/m³ (62-128 mg/m³).

16
17 In a range-finding study, groups of 10 male and 10 female young-adult Wistar rats
18 inhaled a liquid aerosol of technical methamidophos (75.7%), head-only, for 6 hours/day, for 5
19 days (Pauluhn 1986). The test material was vaporized in a polyethylene glycol E 400-ethanol
20 mixture as vehicle. Measured concentrations were 0 (vehicle control), 1.4, 5.4, and 33.1 mg/m³
21 Measured concentrations, determined by capillary gas chromatography/flame ionization, were a
22 factor of 5 less than nominal concentrations (measured concentrations refer to the 75.7% a.i.).
23 Measured concentrations were taken in the area of the rats' inhalation air. The low analytical
24 concentrations were attributed to precipitation of larger particles in the baffle chamber. Particle
25 size averaged 1.5 μ ; 98% were inhalable. Air exposed and water aerosol control groups were
26 non-concurrent. Clinical signs were recorded after each exposure. No clinical signs were
27 observed in the groups at 1.4 or 5.4 mg/m³ after any exposure. Both sexes of rats in the 33.1
28 mg/m³ group showed slight tremors after the first and second exposures; with subsequent
29 exposures, the rats appeared un-groomed and showed reduced motility and weakness of the rear
30 extremities. These signs were reversible during a 14-day post-exposure observation period. No
31 deaths were recorded. Blood samples taken from male rats for cholinesterase activity inhibition
32 were taken within 10-20 minutes post-exposure and analyzed within one hour post-exposure;
33 measurement of activity was by a modified Ellman method. Erythrocyte, plasma, and brain
34 cholinesterase activity were unaffected at the lower two concentrations, after both one and five

1 exposures (Table 3). Relative to control values, erythrocyte and plasma cholinesterase activity
 2 were inhibited by 21 and 82% after one exposure to 33.1 mg/m³ and by 19 and 84% after five
 3 exposures. These values indicate no progressive increase in cholinesterase activity inhibition
 4 with repeat exposure. In the group that inhaled 33.1 mg/m³, brain cholinesterase activity,
 5 measured only after the fifth exposure, was reduced by 67% relative to the control value.
 6

7 Following the range-finding study described above, Pauluhn (1986) exposed groups of
 8 five male and five female young-adult Wistar rats, head-only, to measured liquid aerosol at
 9 methamidophos concentrations of 11.4, 24.3, 45.0, 195.5, 241.7, or 350.3 mg/m³ for 4 hours.
 10 The protocol was the same as described for the range-finding study above. There were no
 11 clinical signs observed in the vehicle control or groups exposed to 11.4 or 24.3 mg/m³.
 12 Moderate tremor, staggering and high gait, reduced motility, bristling, and un-groomed coats
 13 were observed in rats exposed to 45.0 mg/m³. These clinical signs were more marked in the
 14 groups that inhaled 195.5, 241.7, or 350.3 mg/m³; additional signs in the groups exposed to
 15 350.3 mg/m³ included exophthalmos, reddened and bloody eyelids, corneal opacity, and
 16 dyspnea. No rats died in the lower three exposure groups. Mortality was 30% (1 male, two
 17 females), 80% (3 males and five females), and 90% in the groups exposed to 195.5, 241.7, and
 18 350.3 mg/m³, respectively (Table 2). The calculated LC₅₀ for the sexes combined was 213
 19 (174.6-260.7) mg/m³. Rats that died during exposure showed reddened noses, lung edema, pale
 20 organs, and hemorrhagic areas in the gastrointestinal tract. Rats that were sacrificed following
 21 the 14-day observation period showed no apparent organ effects. Pulmonary function tests
 22 showed lung resistance increased in a concentration-related manner, but only following
 23 acetylcholine challenge. The effect of methamidophos on pulmonary function of rats exposed to
 24 24 mg/m³ is more fully discussed in Pauluhn et al. (1987). Blood was sampled for cholinesterase
 25 activity approximately one hour after exposure. Plasma cholinesterase was inhibited by
 26 approximately 50% at 11.4 mg/m³, but erythrocyte acetylcholinesterase activity was unaffected
 27 in this group (102% of pre-exposure value) (Table 3). Erythrocyte acetylcholinesterase activity
 28 was inhibited by 9% and 30% relative to pre-exposure values in the 24.3 and 45.0 mg/m³ groups,
 29 respectively. Groups exposed to higher concentrations were not tested.
 30

TABLE 3. Plasma, Erythrocyte, and Brain Cholinesterase Activity in Rats Following Acute and Repeat Exposure (percent of control value)

| Concentration (mg/m ³) | Plasma | Erythrocyte | Brain ^a |
|--|-----------------------|-------------|--------------------|
| Six hour, 5-day repeat exposure | | | |
| 0 | 86 ^b | 96 | — |
| 1.4 | 114, 113 ^c | 109, 109 | 118 |
| 5.4 | 95, 87 | 99, 101 | 103 |
| 33.1 | 18, 16 | 79, 81 | 33 |
| Four-hour exposure | | | |
| 0 | 84 | 119 | — |
| 6.4 | 85 | 116 | — |
| 11.4 | 53 | 102 | — |
| 24.3 | 36 | 92 | — |
| 45.0 | 13 | 70 | — |

Source: Pauluhn 1986 (data taken from pp. 45 and 79-80).

^a Brain cholinesterase values are activity inhibition relative to the control group on day 5.

^b Control values for plasma and erythrocyte cholinesterase activity are before and after sham exposure.

^c Values for plasma and erythrocyte cholinesterase activity are after one and five exposures.

n = 5-10 rats.

1
2 In an unpublished study, rats inhaled the vapors of Monitor (95% technical) which was
3 heated to 140°C to enhance vaporization (U.S. EPA 1976). The exposure duration was four
4 hours. No deaths occurred although plasma and erythrocyte cholinesterase activity was
5 depressed by 20-30%. No further details were reported.
6

7 **3.2. Repeat-Exposure Studies**

8
9 The five-day repeat inhalation toxicity study conducted by Pauluhn (1986) is discussed in
10 the previous section. In a subchronic inhalation toxicity study, groups of 10 Wistar
11 rats/sex/concentration inhaled an aerosol of methamidophos, 73.4%, for three months (head/nose
12 only) (Pauluhn and Cole 1988). Exposures were for 6 hours/day, 5 days/week. Methamidophos
13 was aerosolized in polyethylene glycol E 400:ethanol (1:1). The mean analytical concentrations
14 in the exposure chambers were 0, 1.1, 5.4, and 23.1 mg/m³. These concentrations are similar to
15 those used in the five-day repeat-exposure study. The mean mass aerodynamic diameters of the
16 methamidophos particles in the groups were 1.52±0.13, 1.26±0.04, and 1.53±0.09 μ,
17 respectively. Treatment-related effects were not observed in the low-concentration group.
18 Relative to the vehicle control values, cholinesterase activities in erythrocytes and plasma were
19 inhibited by 7-28% and 38-63%, respectively, throughout the treatment period in the mid-
20 concentration group. At the end of the study, brain cholinesterase activity was inhibited by 25-
21 29% in the mid-concentration group. There was no substantive difference in the magnitude of
22 the response on plasma or erythrocyte cholinesterase inhibition from week 1-13. The following
23 effects were observed in males and females in the 23.1 mg/m³ concentration group: slight to
24 moderate muscle tremors, aggressive behavior, decreased food consumption (5-28%)
25 accompanied by decreased body weight gain (53%), increased plasma lactate dehydrogenase and
26 glutamate oxaloacetate transaminase activities (males only), small decreases in some clinical
27 chemistry values, decreased absolute and relative spleen weight, and inhibition of cholinesterase
28 activities in erythrocytes (15-44%) plasma (53-93%) throughout the treatment period, and brain
29 (45-47%) at study termination. There was no substantive difference in the magnitude of the
30 response on plasma or erythrocyte cholinesterase activity inhibition from weeks 1-13. When
31 treatment was discontinued, cholinesterase activities in erythrocyte and plasma returned to
32 pretreatment values (brain was not determined).
33

34 **3.3. Neurotoxicity**

35
36 Acute toxicity studies showed that methamidophos is neurotoxic. Signs of
37 acetylcholinesterase activity inhibition were observed in rats inhaling methamidophos for 1 to 6
38 hours (Sangha 1983; 1984; Pauluhn 1986). Cholinergic signs were also observed in oral studies
39 of developmental and reproductive toxicity. See Section 4.2 for mechanism of toxicity of
40 organophosphate pesticides. Doses below the LD₅₀ failed to instill delayed polyneuropathy in
41 hens (IPCS 2002).
42

43 **3.4. Developmental/Reproductive Toxicity**

44
45 No inhalation studies were conducted that addressed the developmental or reproductive
46 toxicity of methamidophos. Reproductive and developmental toxicity studies that used the oral
47 route of administration were reviewed by IPCS (2002) and HSDB (2004). These studies are

1 briefly reviewed here to show that methamidophos is not a teratogen. In a two-generation
2 reproductive toxicity study, male and female CD rats received diets containing technical
3 methamidophos (70.5% purity) at a concentration of 0, 3, 10, or 33 ppm (equivalent to 0, 0.15,
4 0.5, or 1.6 mg/kg body weight per day) for at least 100 days before mating. Females were
5 treated throughout gestation and lactation. Methamidophos at 33 ppm increased the incidence of
6 clinical signs in both parental rats and pups, generally reduced body weight of parental rats,
7 reduced litter and pup weights, decreased the viability of pups, and reduced the proportion of
8 fertilized females giving birth to second generation pups. The NOAEL was 0.5 mg/kg/day.
9 Similar effects were found in a second two-generation study of reproductive toxicity with CD
10 rats. The NOAEL for parental and developmental toxicity was 1 ppm in the diet (0.1 mg/kg/day)
11 on the basis of >20% inhibition of plasma, erythrocyte and brain cholinesterase activity and an
12 8% reduction in body weight. The NOAEL for reproductive toxicity was 30 ppm in the diet,
13 equivalent to 2.4 mg/kg/day.

14
15 Developmental studies with mice, rats and rabbits showed developmental toxicity at high
16 oral concentrations but no teratogenicity. Developmental delay occurred in mice at all doses
17 administered to the dams from day 16 of gestation through day 21 of lactation (0.4 to 4
18 mg/kg/day). Several FOB parameters were affected in weaned pups and neurons in the brain
19 showed structural changes. Rat dams that received methamidophos (70.5%) orally at doses of 0.
20 0.3, 1, or 3 mg/kg body weight on days 6-17 of gestation showed typical signs of cholinergic
21 toxicity at 3 mg/kg/day. The maternal and developmental NOAEL was 1 mg/kg/day. Fetal and
22 litter weight were adversely affected at the 3 mg/kg/day dose. There were no fetal anomalies. In
23 another study with rat dams treated with 0, 1, or 2 mg/kg/day on days 6-15 of gestation,
24 fetotoxicity and increased anomalies were observed at both test concentrations. Rabbit dams
25 administered 0, 0.1, 0.5, or 2.5 mg/kg/day showed reduced weight gain only at the highest dose.
26 There was no developmental toxicity and no external, visceral, or skeletal anomalies occurred.

27 28 **3.5. Genotoxicity**

29
30 The genetic toxicology of methamidophos was reviewed by IPCS (2002) and HSDB
31 (2004). An extensive range of studies has been performed with methamidophos in bacteria and
32 mammalian cells *in vitro* and in mammals *in vivo*. Unless otherwise stated, technical
33 methamidophos was tested. Assay results were negative in two studies of reverse mutation in
34 *Salmonella typhimurium* (TA98, TA100, TA1535, and TA1537), one study of DNA repair in
35 *Escherichia coli*, one study of unscheduled DNA synthesis in rat primary hepatocytes, two
36 studies of point mutation in Chinese hamster ovary cells, one study of chromosomal aberration in
37 Chinese hamster ovary cells (results were equivocal without metabolic activation), and one study
38 of sister chromatid exchange in Chinese hamster ovary cells. An assay for chromosomal
39 aberrations in mouse spleen cells was positive for the pure compound.

40
41 In *in vivo* tests for micronucleus formation in bone marrow cells of male and female
42 mice, two tests with the technical material, delivered orally or intraperitoneally, were negative,
43 and one test with the pure chemical was positive following intraperitoneal injection. Tests for
44 chromosomal aberrations in bone marrow cells of mice and rats following oral or subcutaneous
45 delivery were negative. Two assays for dominant lethal mutation in male mice following oral
46 administration of the technical material were negative. A single assay for sister chromatid

1 exchange in the same system (intraperitoneal delivery) was positive when the pure compound
2 was delivered intraperitoneally.

3 4 **3.6. Chronic Toxicity/Carcinogenicity**

5
6 Long-term studies of toxicity and carcinogenicity were conducted with dogs, rats, and
7 mice. These unpublished oral studies were reviewed by the IPCS (2002) and HSDB (2004).
8 Four groups of six male and six female beagle dogs consumed diets containing methamidophos
9 (70% pure) at concentrations of 0, 2, 8, or 32 ppm, equivalent to 0, 0.06, 0.24, or 0.96 mg/kg/day
10 for one year. At the end of one year, there were no deaths and no effects on body weight, food
11 consumption, blood chemistry, hematology and urine parameters, or gross or microscopic
12 changes of tissues or organs. At concentrations of 8 and 32 ppm, erythrocyte
13 acetylcholinesterase activity was inhibited in both sexes by >20% and 84-87%, respectively.
14 The NOAEL was 2 ppm in the diet and the LOAEL was 8 ppm on the basis of erythrocyte and
15 brain cholinesterase activity inhibition.

16
17 Technical methamidophos was tested for chronic toxicity and carcinogenicity in a two-
18 year dietary study with male and female F344 rats. Groups of 50 rats of each sex were
19 administered methamidophos in the diet at concentrations of 0, 2, 6, 18, or 54 ppm, equivalent to
20 0, 0.01, 0.29, 0.85, or 2.9 mg/kg/day for two years. Interim sacrifices of 10 rats of each sex took
21 place at 6 and 12 months and at one year. Clinical signs, food consumption, body weight,
22 hematology, and clinical chemistry were monitored throughout the study. At sacrifice, major
23 organs were weighed and organs and tissues were examined grossly and microscopically.
24 Plasma, erythrocyte, and brain cholinesterase were assayed. After about 20 weeks, clinical signs
25 of rough coat, urine staining, loose stools, and skin lesions increased in male and female rats that
26 received 18 and 54 ppm in the diet. Body weight was depressed in males at 18 and 54 ppm and
27 in females at 54 ppm. The relative weight of the testes was decreased in male rats that received
28 18 or 54 ppm, and the relative brain weight was increased in both sexes at 54 ppm. No
29 neoplasms related to treatment were observed. Cholinesterase activity was inhibited by
30 treatment in a dose-related manner (time of sampling not clearly stated). At 6, 18, and 54 ppm,
31 plasma cholinesterase activity was inhibited by 26-47%, 70-71%, and 91%, respectively. At the
32 respective concentrations, brain cholinesterase activity was inhibited by 31-39%, 64%, and 75-
33 79%, and erythrocyte cholinesterase activity was inhibited by 32-36%, 65-68%, and 75-81%.
34 The NOAEL was 2 ppm in the diet, equivalent to 0.1 mg/kg/day.

35
36 Technical methamidophos was tested for chronic toxicity and carcinogenicity in a two-
37 year dietary study with male and female CD1 mice. Groups of 50 mice of each sex were
38 administered methamidophos in the diet at concentrations of 0, 1, 5, or 25 ppm, equivalent to 0,
39 0.14, 0.67, or 3.5 mg/kg/day for males and 0, 0.18, 0.78, and 4 mg/kg/day for females. The
40 protocol was the same as in the study with rats described above, except that cholinesterase
41 activity was not measured. Mortality rate, clinical signs, hematology, and gross pathological
42 appearance were not affected by treatment. At 25 ppm in the diet, food consumption and body
43 weight gain were significantly decreased in both sexes. At 25 ppm, relative organ weights were
44 affected in one or both sexes, and males showed diffuse interstitial pneumonia. There were no
45 neoplasms that could be attributed to treatment. The NOAEL was 5 ppm in the diet; decreased
46 weight gain was seen at the higher concentration.

47

3.7. Summary

Acute inhalation lethality studies were conducted with the rat. Studies conducted in two different laboratories, both with liquid aerosols and conducted over the same exposure duration, produced different results. The 4-hour LC₅₀ values were 63.2-76.5 mg/m³ (Sangha 1984) and 213 mg/m³ (Pauluhn 1986). In both studies, nominal concentrations were up to 8-10 times greater than analytical concentrations, indicating difficulty in sampling and measuring methamidophos. The 1-hour LC₅₀ was 377-241 mg/m³ (Sangha 1983). In both studies, cholinergic signs including salivation, lacrimation, decreased activity, muscle fasciculation, ataxia, gasping and dyspnea, tremors, tearing, and exophthalmos, cornea opacity, and rhinorrhea were observed. Concentrations of 1.4, 5.4, and 33.1 mg/m³, administered for 6 hours were non-lethal; no clinical signs were observed at the lower two concentrations (Pauluhn 1986).

The majority of evidence indicates that methamidophos is not genotoxic, carcinogenic, or a reproductive or developmental toxicant. Delays in development were associated with maternal toxicity.

4. SPECIAL CONSIDERATIONS

4.1. Metabolism and Disposition

Inhalation studies with methamidophos that addressed metabolism were not located. Dermal absorption in humans is estimated at 4.8% (IPCS 2002). Following oral administration to rats, radiolabeled methamidophos (S-methyl-¹⁴C) was rapidly absorbed, distributed evenly throughout the body (due to its high aqueous solubility), metabolized, and excreted (Crossley and Tutass 1969, reviewed in IPCS 2002). Excretion was mainly via the urine in the form of acid metabolites and through expired air as carbon dioxide. Greater than 50% of the administered dose was excreted within 3 days. Biotransformation involved hydrolysis at the P-N bond and at the O- and S-ester bonds. Several intermediate metabolites were found in the urine, with the ultimate metabolite being phosphoric acid. A postulated metabolite, methyl mercaptan, was metabolized to carbon dioxide. Methamidophos does not contain aryl or carboxyl groups and so is not metabolized by A-esterases or carboxylesterases, enzymes that show age-related dependency in detoxification (Moser 1999; Padilla et al. 2000).

The metabolism of methamidophos was followed in lactating goats (studies reviewed by WHO 2003). ¹⁴C-Methamidophos was administered orally for 1-7 days. Methamidophos was not detected in tissues following sacrifice at 18 hours post-dose. Most of the radioactive residues were associated with the metabolic pool including proteins and amino acids, specifically methionine. Only trace amounts were secreted in the milk. The proposed metabolic pathway included hydrolysis to form desamino methamidophos and desmethyl methamidophos and methyl transfer from the S-methyl moiety to form methionine with subsequent transformation to form choline and phospholipids. Oxidation of small carbon fragments formed by the ester/thioester hydrolysis of methamidophos and /or methionine may lead to production of CO₂ and incorporation of ¹⁴C into the metabolic pool.

Following intravenous administration of 8 mg/kg (considered a near-toxic dose) ¹⁴CH₃S-labeled methamidophos to female Sprague-Dawley rats, cholinergic signs of tremors, salivation, and lacrimation were most severe at 20-60 minutes which correlated with peak inhibition of

1 acetylcholinesterase activity in the brain of 15-20% of control (Gray et al. 1982). Peak levels of
2 radioactivity were achieved in the tissues within 1-10 minutes. Within 24 hours, 47% of the
3 radioactivity was recovered in the urine and 34% as $^{14}\text{CO}_2$.

4
5 In rats, the maximally tolerated dose following either intravenous or oral dosing was the
6 same, 8 mg/kg (Gray et al. 1982; Moser 1999). The intravenous route of administration can be
7 used as a surrogate for the inhalation route. Because the same dose results in similar toxicity
8 following intravenous and oral administration, oral toxicity should reflect inhalation toxicity.

9 10 **4.2. Mechanism of Toxicity**

11
12 Methamidophos is an organophosphate ester pesticide containing both an amide group
13 and a sulfur group single bonded to a pentavalent phosphorus. The presence of an oxygen
14 double-bonded to the phosphorus (oxon group) indicates that methamidophos does not need to
15 be bioactivated *in vivo* to its oxygen analogue to exert its toxic action. The mode of action of
16 organophosphate pesticides involves inhibition of the B-esterase, acetylcholinesterase (Costa
17 2008). Organophosphate esters attach to the serine hydroxyl group of the active site of
18 acetylcholinesterase, the enzyme responsible for the destruction and termination of the biological
19 activity of the neurotransmitter acetylcholine. When unbound acetylcholine accumulates at the
20 cholinergic nerve endings, there is continual stimulation of electrical activity. The resulting
21 signs of toxicity from stimulation of the muscarinic receptors of the parasympathetic autonomic
22 nervous system are manifest as increased secretions, bronchoconstriction, miosis, gastrointestinal
23 cramps, diarrhea, urination, and bradycardia. Stimulation of the parasympathetic junctions of the
24 autonomic nervous system as well as the junctions between nerves and muscles cause
25 tachycardia, hypertension, muscle fasciculation, tremors, muscle weakness, and flaccid paralysis.
26 Symptoms resulting from effects on the central nervous system include restlessness, emotional
27 lability, ataxia, lethargy, mental confusion, loss of memory, generalized weakness, convulsion,
28 cyanosis, and coma. Acute toxicity of the organophosphate pesticides does not correspond with
29 anticholinesterase potency (Chambers et al. 1990), indicating that metabolism is an important
30 factor in determining overall toxicity.

31
32 Inhibition of acetylcholinesterase activity and other cholinesterases by organophosphate
33 esters is generally long lasting, hours to days (Costa 2008). In the case of methamidophos; the
34 blood and brain cholinesterase activity of rats administered an oral maximum tolerated dose
35 recovered partially within 24 hours and nearly completely within 72 hours (Moser 1999).
36 Preweanling rats (post-natal day 17) recovered faster than adult rats; at 24 hours post-exposure,
37 both blood and brain cholinesterase activity were approximately 30-50% of control in adult rats
38 and approximately 65-70% of control in preweanling rats. *In vitro*, methamidophos was found
39 to be a weak to moderate inhibitor of cholinesterase in tissues of several species including
40 humans (IPCS 2002). In these assays, enzyme activity was rapidly and spontaneously
41 reactivated.

42
43 Organophosphate pesticides also inhibit butylcholinesterase, the primary form of
44 cholinesterase found in blood plasma. The toxicological significance of butylcholinesterase
45 activity inhibition is unknown. Acetylcholinesterase is the primary form of cholinesterase found
46 in erythrocytes and is present at neuromuscular and nerve-nerve junctions. Due to human
47 variability, it is difficult to measure cholinesterase activity inhibition of <20% (U.S. EPA

1 2000b). At greater than 30% erythrocyte acetylcholinesterase activity inhibition or 50% plasma
2 activity inhibition, workers are withdrawn from pesticide application areas (U.S. EPA 2000b;
3 ACGIH 2008).

4.3. Structure-Activity Relationships

7 Organophosphate and carbamate pesticides have a common mode of action (Costa 2008).
8 Compared to carbamic acid esters which are poor substrates for cholinesterase-type enzymes,
9 the organophosphate ester pesticides form a stable bond with acetylcholinesterase.

11 Information is available on the relative oral toxicity of multiple organophosphate
12 pesticides (U.S. EPA 2006a; 2007). The relative toxicity of methamidophos and the related
13 pesticide acephate as indicated by calculated oral benchmark dose values, BMD₁₀ and BMDL₁₀,
14 is summarized in Table 4. The benchmark doses were based on rat brain cholinesterase activity
15 inhibition following repeated oral administration. Chronic toxicity as determined by benchmark
16 doses does not correlate with acute oral toxicity values (Section 2), indicating the role of
17 metabolism in toxicity. If methamidophos is assigned a potency of 1.00 for all routes of
18 administration, then the relative potency for acephate is 0.08 (oral), 0.0025 (dermal), and 0.208
19 (inhalation) (U.S. EPA 2007).

4.4. Other Relevant Information

4.4.1. Species Variability

24 Inhalation studies with usable data were conducted only with rats. The route and rate of
25 biotransformation of organophosphate pesticides is highly species-specific and dependent on the
26 substituent chemical groups attached to the parent ester (Costa 2008). Baseline erythrocyte
27 acetylcholinesterase activity is higher in humans than in other species (Ellin 1981). Oral dosing
28 studies with methamidophos were available for rats, but the human oral dosing study used both
29 acephate and methamidophos. An acute oral dose of 1 mg/kg methamidophos to male and
30 female Long-Evans rats decreased whole blood cholinesterase activity to 40-50% of the control
31 value (Moser 1999). In the human study, a 0.2 mg/kg/day oral dose of methamidophos and
32 acephate in a 1:4 ratio (0.04 mg/kg/day methamidophos or 2.8 mg/kg/day for a 70-kg person)
33 had no effect on erythrocyte cholinesterase activity but significantly inhibited plasma
34 cholinesterase activity (raw data not provided) after 16 days of dosing (Garofalo et al. 1973).

36 In an *in vitro* study, the concentration of methamidophos required to inhibit 50% of the
37 activity of cholinesterase from rat and mouse brain was the same, 2.0×10^{-5} M (Hussain et al.
38 1985). The concentration required to inhibit acetylcholinesterase activity in human erythrocytes
39 was 2.3×10^{-5} M. No useful data on species variability could be gleaned from this study.

4.4.2. Susceptible Populations

43 Humans vary by gender, age, and genetic make-up in their sensitivity to cholinesterase
44 inhibitors. The erythrocyte acetylcholinesterase activity of adults (153 ± 24 activity units;
45 acetylthiocholine substrate) is greater than that of healthy newborn infants (97 ± 15 activity units)
46 by a factor of 1.6 (Herz et al. 1975). Following *in utero* exposure to organophosphate pesticides,
47 dams exhibit greater cholinesterase inhibition than fetuses (U.S. EPA 2006a). Developmental

1 neurotoxicity studies with methamidophos showed that protection of the rat dam against
 2 cholinesterase activity inhibition is protective against pup acetylcholinesterase activity inhibition
 3 *in utero*.

4
 5 The U.S. EPA (U.S. EPA 2006a) identified infants and juveniles as populations
 6 susceptible to the toxicity of organophosphate pesticides. In the absence of human data on age-
 7 related sensitivity, the sensitivity to cholinesterase activity inhibition in adult and juvenile rats to
 8 methamidophos can be used as a surrogate for humans. The U.S. EPA used a dose-response
 9 modeling approach for evaluating quantitatively the relative sensitivity between juvenile and
 10 adult rats. For organophosphate pesticides, only repeated dosing exposure studies were
 11 considered. In this approach, benchmark dose values (BMD) were calculated for inhibition of
 12 juvenile and adult brain cholinesterase data. The ratio of juvenile and adult BMDs for each
 13 organophosphate pesticide was calculated and used as a chemical-specific uncertainty factor to
 14 protect the young. For methamidophos, the BMDs for adult and juvenile female rats were 0.18
 15 and 0.09 mg/kg/day, respectively (Table 4). For adult and juvenile male rats, the BMDs were
 16 0.10 and 0.08 mg/kg/day, respectively. The U.S. EPA assigned an factor of 2 to methamidophos.
 17 For AEGLS, this factor corresponds to an intraspecies uncertainty factor. Data on the related
 18 chemical, acephate, are also summarized in Table 4.
 19

| TABLE 4. Adult and Juvenile Rat BMD₁₀ and BMDL₁₀ Values for Brain Acetylcholinesterase Activity Following Repeated Oral Dosing with Methamidophos or Acephate | | |
|--|-------------------------------------|--------------------------------------|
| Age and Sex | BMD₁₀ (mg/kg/day) | BMDL₁₀ (mg/kg/day) |
| Methamidophos | | |
| Adult male | 0.10 | 0.08 |
| Adult female | 0.18 | 0.11 |
| Juvenile male | 0.08 | 0.06 |
| Juvenile female | 0.09 | 0.08 |
| Acephate | | |
| Adult male | 0.27 | 0.22 |
| Adult female | 1.25 | 0.30 |
| Juvenile male | 0.42 | 0.47 |
| Juvenile female | 1.13 | 0.60 |

20 The BMD values were based on a 10% response relative to the control.

21 Source: Table I.B-6, U.S. EPA 2006a.

22
 23 A study of the sensitivity of pre-weanling and adult rats to methamidophos neurotoxicity
 24 following an acute oral exposure showed little age-related difference (Moser 1999). Using the
 25 oral route of exposure, the authors evaluated the acute age-related differences in sensitivity to
 26 methamidophos among preweanling (post-natal day 17; PND 17) and adult (PND 70) Long
 27 Evans rats of both sexes. Control data for cholinesterase activity (nmol ³H-labeled acetylcholine
 28 iodide hydrolyzed/min/mg/tissue) were (a) male brain: PND 17, 4.8, adult 9.0; male blood: PND
 29 17, 0.69, adult, 0.56; (b) female brain: PND 17, 5.2, adult 9.7; female blood: PND 17, 0.75,
 30 adult, 0.66. Range-finding studies determined the maximally tolerated dose of 8 mg/kg in both
 31 PND 17 and adult rats. Tested doses were 1.0, 2.0, 5.0, 10, and 15 mg/kg. The tested dose of 10
 32 mg/kg resulted in severe cholinergic signs and weight loss. Doses for the definitive study were
 33 1, 4, and 8 mg/kg. The time of peak effect of blood cholinesterase activity inhibition (measured
 34 in whole blood) was 1.5 hours in both groups. Recovery was faster in PND 17 rats than in
 35 adults. PND 17 and adult rats were similarly sensitive to methamidophos as measured by blood

1 and brain cholinesterase activity inhibition. Blood cholinesterase activity inhibition, reported as
2 percent of control activity, was greater than that of brain. The lowest dose of 1 mg/kg produced
3 brain and blood cholinesterase activity inhibition of 30-40%, but there were no behavioral effects
4 observed in that dose group. A U.S. EPA standard functional observational battery (FOB) and
5 motor activity observations were carried out. ED₅₀ values for tremors were approximately 5
6 mg/kg in PND 17 rats and 6 mg/kg in adult rats. The respective values for gait changes/ataxia
7 were approximately 4.3 and 6.0 mg/kg. Based on the Moser data, methamidophos toxicity failed
8 to show age-related sensitivity.
9

10 Although age-related sensitivity is not apparent based on the acute and repeat-exposure
11 oral studies with rats, humans are known to differ in sensitivity to the toxic effects of
12 organophosphate pesticides. Although there is no data on differences among humans regarding
13 metabolism of methamidophos, the oral dosing study of Garofalo et al. (1973) did not reveal
14 differences within the range of tested doses. Therefore, an intraspecies uncertainty factor of 3 is
15 considered appropriate.
16

17 **4.4.3. Concentration-Exposure Duration Relationship**

18

19 Toxicity studies were performed with exposure durations of 1, 4, and 6 hours, but the
20 disparate data made time-scaling calculations inappropriate. The concentration-time relationship
21 for a single endpoint for many irritant and systemically acting vapors and gases may be
22 described by $C^n \times t = k$ (ten Berge et al. 1986). In the absence of empirical data, the time scaling
23 factors of $n = 3$ and $n = 1$ are used to scale to shorter and longer exposure durations, respectively
24 (NRC 2001)
25

26 **4.4.4. Concurrent Exposure Issues**

27

28 Dermal absorption may occur, but percutaneous toxicity is low compared to inhalation
29 exposure as indicated by 4.8% skin absorption in humans (IPCS 2002).
30

31 The pesticide acephate is partially metabolized to methamidophos (Singh 1985). In *in*
32 *vitro* studies with tissues from dogs, methamidophos was 75 to 100 times more potent than
33 acephate in inhibiting acetylcholinesterase activity in brain and erythrocytes and cholinesterase
34 activity in plasma. As indicated by a 4-hour LC₅₀ in rats of 2130 mg/m³ (MSDS 2003), acephate
35 is a factor of 10- to 33-fold less toxic than methamidophos. However, benchmark dose
36 calculations following repeat oral dosing (Table 4) indicate that methamidophos is a factor of 3 -
37 to 12 times more toxic, indicating the role of metabolism following inhalation.
38

39 **5. DATA ANALYSIS FOR AEGL-1**

40 **5.1. Summary of Human Data Relevant to AEGL-1**

41

42 No human inhalation studies were located in the available literature. Occupational
43 monitoring data involved dermal contact and consumer exposure involved oral intake of crop
44 residues. In a 21-day repeat-oral dosing study of methamidophos combined with acephate, a
45 dose of 0.04 mg/kg/day methamidophos (2.8 mg for a 70-kg person) did not inhibit erythrocyte
46 acetylcholinesterase activity of male or female volunteers (Garofalo et al. 1973, reviewed in U.S.

1 EPA 1988; 2000a). Plasma cholinesterase activity was significantly inhibited, but the data were
2 not provided in the review.

3 4 **5.2. Summary of Animal Data Relevant to AEGL-1**

5
6 Pauluhn (1986) exposed rats to several concentrations of methamidophos delivered as a
7 liquid aerosol for 4 and 6 hours. No clinical signs were observed following 4-hour exposures to
8 11.4 or 24.3 mg/m³, and no clinical signs were observed following 6-hour exposures to 1.4 or 5.4
9 mg/m³. In each case the next higher concentration, 45.0 mg/m³ for 4 hours and 33.1 mg/m³ for 6
10 hours produced tremors, but no mortality. At the 11.4 mg/m³ concentration, plasma
11 cholinesterase activity was 53% of the control value and erythrocyte acetylcholinesterase activity
12 was 102% of the control value. At the 24.3 mg/m³ concentration, plasma cholinesterase activity
13 was 36% of the control value and erythrocyte acetylcholinesterase activity was 92% of the
14 control value.

15
16 In a second study, cholinergic signs were observed during the 4-hour exposure of rats to
17 19 mg/m³ (Sangha 1984). The data of Sangha (1984) neither followed an adequate
18 concentration-response curve nor correlated with nominal values, probable indications of the
19 difficulty in sampling and measuring the liquid aerosol. Cholinesterase activity was not
20 measured.

21 22 **5.3. Derivation of AEGL-1**

23
24 The study of Pauluhn (1986) was chosen for AEGL derivations. The data showed a good
25 concentration-response curve, and measured cholinesterase activity correlated with clinical
26 signs. The exposure of rats to 24.3 mg/m³ for 4-hours was chosen as the point of departure for
27 the AEGL-1. There were no clinical signs at this exposure. Clinical signs of tremor and reduced
28 motility were observed during the 4-hour exposure to the next higher concentration of 45.0
29 mg/m³. Plasma cholinesterase activity was 36 percent of the control value and erythrocyte
30 cholinesterase was slightly inhibited (92% of the control value). Because of the apparent
31 difficulty in maintaining liquid aerosols at these concentrations and the disparate data between
32 the two available studies, the 24.3 mg/m³ value was divided by a data base modifying factor of 2.
33 Methamidophos is rapidly metabolized and excreted in rats and humans as indicated by oral
34 dosing studies in both rats and humans (Moser 1999; Garofalo et al. 1973). Therefore, an
35 interspecies uncertainty factor of 3 was applied. Infants and juveniles may be more sensitive to
36 organophosphate pesticides than adults. An acute oral dosing study with adult and juvenile rats
37 failed to show differences in sensitivity to methamidophos (Moser 1999). Based on repeat-dose
38 oral studies with adult and juvenile rats, the U.S. EPA (2006a) identified a toxicity ratio between
39 adults and juveniles of 2. Because there were no differences in sensitivity between adult and
40 juvenile rats in the acute oral dosing study, an intraspecies uncertainty factor of 3 is adequate.
41 The total uncertainty factor is 10. The 4-hour 11.4 mg/m³ value was divided by a total
42 modifying/uncertainty factor of 20 (2x10). In the absence of reliable time-scaling information,
43 the resulting 4-hour value of 1.2 mg/m³ was time-scaled ($C^n \times t = k$) using n values of 3 and 1 for
44 shorter and longer exposure durations, respectively (NRC 2001). Because of uncertainty in
45 scaling from 4 hours to 10 minutes, the 10-minute value was set equal to the 30-minute value.
46 Values are listed in Table 5. Calculations are in Appendix A and a graph of the AEGL values in
47 relation to toxicity data is in Appendix B.

1

| TABLE 5. AEGL-1 Values for Methamidophos | | | | |
|--|-----------------------|-----------------------|-----------------------|------------------------|
| 10-min | 30-min | 1-h | 4-h | 8-hour |
| 2.4 mg/m ³ | 2.4 mg/m ³ | 1.9 mg/m ³ | 1.2 mg/m ³ | 0.61 mg/m ³ |

2

3 The AEGL-1 values are supported by the subchronic study of Pauluhn and Cole (1988).
 4 In that study no treatment related effects were observed in rats inhaling 1.1 mg/m³ for 13 weeks.
 5 At 5.4 mg/m³, erythrocyte and brain cholinesterase activities were inhibited by <30%
 6 throughout the treatment period.

7

8 **6. DATA ANALYSIS FOR AEGL-2**

9 **6.1. Summary of Human Data Relevant to AEGL-2**

10

11 No human inhalation studies relevant to development of AEGL-2 values were located in
 12 the available literature.

13

14 **6.2. Summary of Animal Data Relevant to AEGL-2**

15

16 Pauluhn (1986) exposed rats to several concentrations of methamidophos delivered as a
 17 liquid aerosol for 4 and 6 hours. No clinical signs were observed following 4-hour exposures to
 18 11.4 or 24.3 mg/m³, and no clinical signs were observed following 6-hour exposures to 1.4 or 5.4
 19 mg/m³. In each case the next higher concentration, 45.0 mg/m³ for 4 hours and 33.1 mg/m³ for 6
 20 hours produced tremors, but no mortality. At 45.0 mg/m³ plasma cholinesterase activity was
 21 13% of the control value and erythrocyte cholinesterase activity was 70% of the control value.
 22 In a second study, cholinergic signs were observed during the 4-hour exposure of rats to 19
 23 mg/m³ (Sangha 1984). The data of Sangha (1984) did not follow a good concentration-response
 24 curve nor correlate with nominal concentrations, probable indications of the difficulty in
 25 sampling and measuring the liquid aerosol. Cholinesterase activity was not measured.

26

27 **6.3. Derivation of AEGL-2**

28

29 The 4-hour exposure of rats to 45.0 mg/m³ was chosen as the point of departure for the
 30 AEGL-2. Clinical signs consisted of tremor and reduced motility. Cholinesterase activity was
 31 inhibited to 13 percent of control in plasma and 70% of control in erythrocytes. Because of the
 32 apparent difficulty in maintaining liquid aerosols at these concentrations and the disparate data
 33 between the two available studies, the 45.0 mg/m³ value was divided by a data base modifying
 34 factor of 2. Methamidophos is rapidly metabolized and excreted in rats and humans as indicated
 35 by oral dosing studies (Moser 1999; Garofalo et al. 1973). Therefore, an interspecies uncertainty
 36 factor of 3 was applied. Infants and juveniles may be more sensitive to organophosphate
 37 pesticides than adults. An acute oral dosing study with adult and juvenile rats failed to show
 38 age-related differences in sensitivity to methamidophos (Moser 1999). Based on repeat-dose
 39 oral studies with adult and juvenile rats, the U.S. EPA (2006a) identified an uncertainty factor of
 40 2 to protect children. Because there were no differences in sensitivity between adult and juvenile
 41 rats in the acute oral dosing study, an intraspecies uncertainty factor of 3 is adequate. The total
 42 uncertainty factor is 10. The 4-hour 45.0 mg/m³ value was divided by a total
 43 modifying/uncertainty factor of 20 (2x10). In the absence of reliable time-scaling information,
 44 the resulting 4-hour value of 2.3 mg/m³ was time-scaled ($C^n \times t = k$) using n values of 3 and 1 for

shorter and longer exposure durations, respectively (NRC 2001). Because of uncertainty in scaling from 4 hours to 10 minutes, the 10-minute value was set equal to the 30-minute value. Values are summarized in Table 6. Calculations are in Appendix A and a category graph of the toxicity data in relation to AEGL values is in Appendix B.

| 10-min | 30-min | 1-h | 4-h | 8-h |
|-----------------------|-----------------------|-----------------------|-----------------------|-----------------------|
| 4.5 mg/m ³ | 4.5 mg/m ³ | 3.6 mg/m ³ | 2.3 mg/m ³ | 1.1 mg/m ³ |

The AEGL-2 values are supported by the subchronic study of Pauluhn and Cole (1988). In that study no treatment related effects were observed in rats inhaling 1.1 mg/m³ for 13 weeks. At 5.4 mg/m³, erythrocyte and brain cholinesterase activities were inhibited by <30% throughout the treatment period. At study termination following treatment with 23.1 mg/m³, brain acetylcholinesterase was inhibited by 45-47%.

7. DATA ANALYSIS FOR AEGL-3

7.1. Summary of Human Data Relevant to AEGL-3

No human inhalation studies relevant to derivation of AEGL-3 values were located in the available literature.

7.2. Summary of Animal Data Relevant to AEGL-3

The 4-hour exposure of rats to methamidophos delivered as a liquid aerosol at concentrations of 11.4 to 350.3 mg/m³ in the study of Pauluhn (1986) followed an adequate concentration-response curve. Mortalities for rats inhaling 11.4, 24.3, 45.0, 195.5, 241.7, or 350.3 mg/m³ for 4 hours were 0/10, 0/10, 0/10, 3/10, 8/10, and 9/10, respectively. Using U.S. EPA's Benchmark Concentration (BMC) program (V2.8), Benchmark concentration were calculated. The 4-hour BMCL₀₅ was 56.27 mg/m³, and the 4-hour BMC₀₁ was 101.54 mg/m³ (see Appendix C for program output).

7.3. Derivation of AEGL-3

Both the 4-hour BMCL₀₅ of 56.27 mg/m³ and the 4-hour BMC₀₁ of 101.54 mg/m³ were considered as points of departure for developing AEGL-3 values for methamidophos. Although the lower value, in this case the BMCL₀₅ of 56.27 mg/m³, is generally chosen as the threshold for mortality in developing AEGL-3 values, this value was considered an artifact of the large gap between tested concentrations of 45.0 and 195.5 mg/m³. The BMCL₀₅ of 56.27 mg/m³ appeared unrealistically low based on 30% mortality when the concentration was increased more than 4-fold to 195.5 mg/m³. This value is also close to the 45.0 mg/m³ value that resulted in effects considered consistent with the AEGL-2. The 4-hour BMC₀₁ for methamidophos delivered as a liquid aerosol was considered the threshold for mortality of rats. Because of the apparent difficulty in maintaining liquid aerosols at these concentrations and the disparate data between the two available studies, the 101.54 mg/m³ value was divided by a data base modifying factor of 2. Methamidophos is rapidly metabolized and excreted in rats and humans as indicated by oral dosing studies (Moser 1999; Garofalo et al. 1973). Therefore, an interspecies uncertainty factor

1 of 3 was applied. Infants and juveniles may be more sensitive to organophosphate pesticides
 2 than adults. An acute oral dosing study with adult and juvenile rats failed to show age-related
 3 differences in sensitivity to methamidophos (Moser 1999). Based on repeat-dose oral studies
 4 with adult and juvenile rats, the U.S. EPA (2006a) identified an uncertainty factor of 2 to protect
 5 the sensitive population of children. Because there were no differences in sensitivity between
 6 adult and juvenile rats in the acute oral dosing study, an intraspecies uncertainty factor of 3 is
 7 adequate. The total uncertainty factor is 10. The 4-hour 101.54 mg/m³ value was divided by a
 8 total modifying/uncertainty factor of 20 (2x10). In the absence of reliable time-scaling
 9 information, the resulting 4-hour value of 5.1 mg/m³ was time-scaled ($C^n \times t = k$) using n values
 10 of 3 and 1 for shorter and longer exposure durations, respectively (NRC 2001). Because of
 11 uncertainty in scaling from 4 hours to 10 minutes, the 10-minute value was set equal to the 30-
 12 minute value. Values are summarized in Table 7, calculations are in Appendix A, and a category
 13 graph of the toxicity data in relation to AEGL values is in Appendix B.
 14

TABLE 7. AEGL-3 Values for Methamidophos

| 10-min | 30-min | 1-h | 4-h | 8-h |
|----------------------|----------------------|-----------------------|-----------------------|-----------------------|
| 10 mg/m ³ | 10 mg/m ³ | 8.1 mg/m ³ | 5.1 mg/m ³ | 2.5 mg/m ³ |

15
16

17 The AEGL-3 values are supported by the subchronic study of Pauluhn and Cole (1988).
 18 In that study, treatment of rats with 23.1 mg/m³ for 90 days produced clinical signs of tremor;
 19 plasma, erythrocyte, and brain cholinesterase activity were substantially inhibited, but no deaths
 20 were reported.

21

22 8. SUMMARY OF AEGLs

23 8.1. AEGL Values and Toxicity Endpoints

24

25 AEGL values are summarized in Table 8. Derivation summaries are in Appendix D.

26

TABLE 8. Summary of AEGL Values for Methamidophos

| Classification | Exposure Duration | | | | |
|--------------------------|-----------------------|-----------------------|-----------------------|-----------------------|------------------------|
| | 10-min | 30-min | 1-h | 4-h | 8-h |
| AEGL-1 (Nondisabling) | 2.4 mg/m ³ | 2.4 mg/m ³ | 1.9 mg/m ³ | 1.2 mg/m ³ | 0.61 mg/m ³ |
| AEGL-2 (Disabling) | 4.5 mg/m ³ | 4.5 mg/m ³ | 3.6 mg/m ³ | 2.3 mg/m ³ | 1.1 mg/m ³ |
| AEGL-3 (Lethal) | 10 mg/m ³ | 10 mg/m ³ | 8.1 mg/m ³ | 5.1 mg/m ³ | 2.5 mg/m ³ |

27

28

29 8.2. Comparison with Other Standards and Guidelines

30

31 No standards or guidelines for methamidophos in air were found (Table 9). The American
 32 Conference of Government Industrial Hygienists (ACGIH) has not derived a Threshold Limit
 33 Value-Time Weighted Average for methamidophos. The ACGIH has calculated a Biological
 34 Exposure Index for acetylcholinesterase inhibiting chemicals (ACGIH 2008). The value, based
 35 on erythrocyte cholinesterase activity inhibition, is 70% of an individual's baseline.
 36

| TABLE 9. Standards and Guidelines for Methamidophos | | | | | |
|--|--------------------------|-----------------------|-----------------------|-----------------------|------------------------|
| Guideline | Exposure Duration | | | | |
| | 10 min | 30 min | 1 h | 4 h | 8 h |
| AEGL-1 | 2.4 mg/m ³ | 2.4 mg/m ³ | 1.9 mg/m ³ | 1.2 mg/m ³ | 0.61 mg/m ³ |
| AEGL-2 | 4.5 mg/m ³ | 4.5 mg/m ³ | 3.6 mg/m ³ | 2.3 mg/m ³ | 1.1 mg/m ³ |
| AEGL-3 | 10 mg/m ³ | 10 mg/m ³ | 8.1 mg/m ³ | 5.1 mg/m ³ | 2.5 mg/m ³ |
| ERPG-1 (AIHA) ^a | | | — | | |
| ERPG-2 (AIHA) | | | — | | |
| ERPG-3 (AIHA) | | | — | | |
| IDLH (NIOSH) ^b | | — | | | |
| REL-TWA (NIOSH) ^c | | | | | — |
| OSHA PEL (NIOSH) ^d | | | | | — |
| TLV-TWA (ACGIH) ^e | | | | | — |
| WEEL (AIHA) ^f | | | | | — |
| MAK (Germany) ^g | | | | | — |
| MAC (The Netherlands) ^h | | | | | — |
| LLV (Sweden) ⁱ | | | | | — |

^aERPG (Emergency Response Planning Guidelines, American Industrial Hygiene Association

The ERPG-1 is the maximum airborne concentration below which it is believed nearly all individuals could be exposed for up to one hour without experiencing other than mild, transient adverse health effects or without perceiving a clearly defined objectionable odor.

The ERPG-2 is the maximum airborne concentration below which it is believed nearly all individuals could be exposed for up to one hour without experiencing or developing irreversible or other serious health effects or symptoms that could impair an individual's ability to take protective action.

The ERPG-3 is the maximum airborne concentration below which it is believed nearly all individuals could be exposed for up to one hour without experiencing or developing life-threatening health effects.

^bIDLH (Immediately Dangerous to Life and Health, National Institute of Occupational Safety and Health)

represents the maximum concentration from which one could escape within 30 minutes without any escape-impairing symptoms, or any irreversible health effects.

^cNIOSH REL-TWA (National Institute of Occupational Safety and Health, Recommended Exposure Limits - Time Weighted Average) is defined analogous to the ACGIH-TLV-TWA.

^dOSHA PEL-TWA (Occupational Safety and Health Administration, Permissible Exposure Limits - Time Weighted Average) is defined analogous to the ACGIH-TLV-TWA, but is for exposures of no more than 10 hours/day, 40 hours/week.

^eACGIH TLV-TWA (American Conference of Governmental Industrial Hygienists, Threshold Limit Value - Time Weighted Average) is the time-weighted average concentration for a normal 8-hour workday and a 40-hour workweek, to which nearly all workers may be repeatedly exposed, day after day, without adverse effect.

^fWEEL (Workplace Environmental Exposure Level Guide) (AIHA 2009) is the 8-hour time-weighted average that is expected to be without adverse health effects during a normal 8-hour day and 40-hour workweek.

^gMAK (Maximale Arbeitsplatzkonzentration [Maximum Workplace Concentration]) (Deutsche Forschungsgemeinschaft [German Research Association] is defined analogous to the ACGIH-TLV-TWA.

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^h**MAC (Maximaal Aanvaarde Concentratie [Maximal Accepted Concentration])** (SDU Uitgevers [under the auspices of the Ministry of Social Affairs and Employment], The Hague, The Netherlands is defined similar to the ACGIH TLV.

ⁱ**LLV (Level Limit Value)** Occupational Exposure Limit of the Swedish National Board of Occupational Safety and Health, Solna, Sweden is defined similar to the ACGIH.

8.3. Data Adequacy and Research Needs

Methamidophos has a low vapor pressure and no usable studies involving inhalation exposure of humans were located in the available literature. An oral dosing study with human volunteers addressed effects consistent with cholinesterase activity inhibition. Inhalation studies with rats as the test species involving several exposure durations produced disparate results. Atmospheres were difficult to maintain and sample or measure as indicated by large differences in nominal and measured concentrations. AEGL values were based on an adequately conducted study with application of a data base modifying factor to account for the disparate results in the available studies. Acute and repeat-dose oral studies involving comparisons of cholinesterase activity inhibition between juvenile and adult rats addressed derivation of a chemical-specific intraspecies uncertainty factor. Compared to some organophosphate pesticides, metabolism in mammalian species appears to be fairly rapid. Metabolism pathways and mode of action are well understood.

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- 3 of Pesticide Programs to Susan Bartow, Office of Pesticide Programs, dated 11/25/08.
- 4
- 5 WHO (World Health Organization). 2003. Pesticide Residues in Food: 2003 Evaluations, Residues.
- 6 Available online:
- 7 [http://www.fao.org/ag/AGP/AGPP/Pesticid/JMPR/Download/2003_eva/methamidophos%202003.pd](http://www.fao.org/ag/AGP/AGPP/Pesticid/JMPR/Download/2003_eva/methamidophos%202003.pdf)
- 8 [f.](http://www.fao.org/ag/AGP/AGPP/Pesticid/JMPR/Download/2003_eva/methamidophos%202003.pdf)

APPENDIX A: Derivation of Methamidophos AEGLs

Derivation of AEGL-1 Values

| | | |
|----|----------------------|---|
| 1 | | |
| 2 | | |
| 3 | | |
| 4 | | |
| 5 | | |
| 6 | Key Study: | Pauluhn, J. 1986. Study for Acute Inhalation Toxicity to the Rat to OECD |
| 7 | | Guideline No 403. Report No. 15661, Bayer AG Institute of Toxicology, |
| 8 | | Wuppertal, Germany. |
| 9 | | |
| 10 | Toxicity endpoint: | No clinical signs; inhibition of plasma (64%) and erythrocyte (8%) |
| 11 | | cholinesterase activity in rats inhaling 24.3 mg/m ³ for 4 hours. The next |
| 12 | | higher concentration of 45.0 mg/m ³ resulted in tremors and greater inhibition |
| 13 | | of cholinesterase activity. |
| 14 | | |
| 15 | Time scaling | C ⁿ x t = k, where n = 3 and 1 for shorter and longer exposure durations, |
| 16 | | respectively (ten Berge et al. 1986) |
| 17 | | |
| 18 | Modifying factor: | 2, based on disparate data |
| 19 | | |
| 20 | Uncertainty factors: | Total uncertainty factor: 10 |
| 21 | | Interspecies: 3, based on rapid metabolism in both rats and humans (Moser |
| 22 | | 1999; Garofalo et al. 1973). |
| 23 | | Intraspecies: 3, based on no differences in sensitivity to acetylcholinesterase |
| 24 | | activity inhibition between juvenile and adult rats in an acute oral study |
| 25 | | (Moser 1999). |
| 26 | | |
| 27 | Calculations: | 24.3 mg/m ³ /20 = 1.22 mg/m ³ |
| 28 | | k = (1.22 mg/m ³) ³ x 240 min = 430.47 mg/m ³ •min |
| 29 | | |
| 30 | 10-min AEGL-1: | Set equal to the 30-minute value (2.4 mg/m ³) based on the 4-hour exposure |
| 31 | | duration |
| 32 | | |
| 33 | 30-min AEGL-1: | C = $\sqrt[3]{(430.47 \text{ mg/m}^3 \cdot \text{min}/30)} = 2.4 \text{ mg/m}^3$ |
| 34 | | |
| 35 | 1-h AEGL-1: | C = $\sqrt[3]{(430.47 \text{ mg/m}^3 \cdot \text{min} /60)} = 1.9 \text{ mg/m}^3$ |
| 36 | | |
| 37 | 4-h AEGL-1: | C = 1.2 mg/m ³ |
| 38 | | |
| 39 | 8-h AEGL-1: | C = (1.2 mg/m ³ x 240)/480 min = 0.61 mg/m ³ |
| 40 | | |

Derivation of AEGL-2 Values

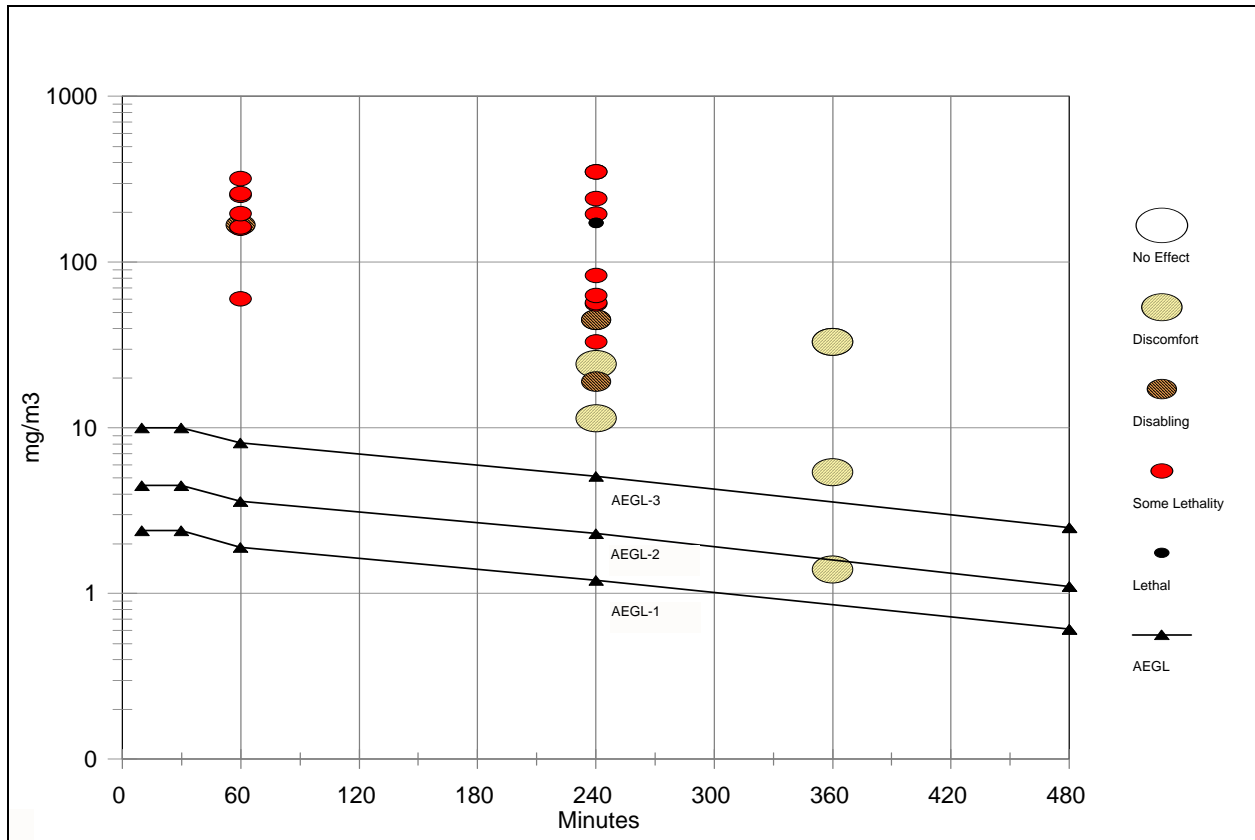
| | | |
|----|----------------------|---|
| 1 | | |
| 2 | | |
| 3 | | |
| 4 | Key Study: | Pauluhn, J. 1986. Study for Acute Inhalation Toxicity to the Rat to OECD |
| 5 | | Guideline No 403. Report No. 15661, Bayer AG Institute of Toxicology, |
| 6 | | Wuppertal, Germany. |
| 7 | | |
| 8 | Toxicity endpoint: | Tremors and inhibition of plasma (87%) and erythrocyte (30%) |
| 9 | | cholinesterase activity inhibition in rats inhaling 45.0 mg/m ³ for 4 hours |
| 10 | | |
| 11 | Time scaling | $C^n \times t = k$, where n = 3 and 1 for shorter and longer exposure durations, |
| 12 | | respectively (ten Berge et al. 1986) |
| 13 | | |
| 14 | Modifying factor: | 2, based on disparate data |
| 15 | | |
| 16 | Uncertainty factors: | Total uncertainty factor: 10 |
| 17 | | Interspecies: 3, based on rapid metabolism in both rats and humans (Moser |
| 18 | | 1999; Garofalo et al. 1973). |
| 19 | | Intraspecies: 3, based on no differences in sensitivity to acetylcholinesterase |
| 20 | | activity inhibition between juvenile and adult rats in an acute oral study |
| 21 | | (Moser 1999). |
| 22 | | |
| 23 | Calculations: | $45.0 \text{ mg/m}^3 / 20 = 2.25 \text{ mg/m}^3$ (rounded to 2.3 mg/m ³ in summary tables) |
| 24 | | $k = (2.25 \text{ mg/m}^3)^3 \times 240 \text{ min} = 2733.75 \text{ mg/m}^3 \cdot \text{min}$ |
| 25 | | |
| 26 | 10-min AEGL-2: | Set equal to the 30-minute value (4.5 mg/m ³) based on the 4-hour exposure |
| 27 | | duration |
| 28 | | |
| 29 | 30-min AEGL-2: | $C = \sqrt[3]{(2733.75 \text{ mg/m}^3 \cdot \text{min} / 30)} = 4.5 \text{ mg/m}^3$ |
| 30 | | |
| 31 | 1-h AEGL-2: | $C = \sqrt[3]{(2733.75 \text{ mg/m}^3 \cdot \text{min} / 60)} = 3.6 \text{ mg/m}^3$ |
| 32 | | |
| 33 | 4-h AEGL-2: | $C = 2.3 \text{ mg/m}^3$ |
| 34 | | |
| 35 | 8-h AEGL-2: | $C = (2.25 \text{ mg/m}^3 \times 240) / 480 \text{ min} = 1.1 \text{ mg/m}^3$ |
| 36 | | |
| 37 | | |

Derivation of AEGL-3 Values

| | | |
|----|----------------------|---|
| 1 | | |
| 2 | | |
| 3 | | |
| 4 | Key Study: | Pauluhn, J. 1986. Study for Acute Inhalation Toxicity to the Rat to OECD |
| 5 | | Guideline No 403. Report No. 15661, Bayer AG Institute of Toxicology, |
| 6 | | Wuppertal, Germany. |
| 7 | | |
| 8 | Toxicity endpoint: | Threshold for lethality in rats at the BMC_{01} of 101.54 mg/m^3 calculated from |
| 9 | | the rat lethality data of Pauluhn (1986). The $BMCL_{05}$ of 56.27 mg/m^3 |
| 10 | | appeared unrealistically low based on 30% mortality when the concentration |
| 11 | | was increased more than 4-fold to 195.5 mg/m^3 . |
| 12 | | |
| 13 | Time scaling | $C^n \times t = k$ where $n = 3$ and 1 for shorter and longer exposure durations, |
| 14 | | respectively (ten Berge et al. 1986). |
| 15 | | |
| 16 | Modifying Factor: | 2, based on disparate data |
| 17 | | |
| 18 | Uncertainty factors: | Total uncertainty factor: 10 |
| 19 | | Interspecies: 3, based on rapid metabolism in both rats and humans (Moser |
| 20 | | 1999; Garofalo et al. 1973). |
| 21 | | Intraspecies: 3, based on no differences in sensitivity to acetylcholinesterase |
| 22 | | activity inhibition between juvenile and adult rats in an acute oral study |
| 23 | | (Moser 1999). |
| 24 | | |
| 25 | Calculations: | $101.54/20 = 5.08 \text{ mg/m}^3$ (rounded to 5.1 in the summary tables) |
| 26 | | $(5.08 \text{ mg/m}^3)^3 \times 240 \text{ minutes} = 31407.45 \text{ mg/m}^3 \cdot \text{min}$ |
| 27 | | |
| 28 | 10-min AEGL-3: | Set equal to the 30-minute value (10 mg/m^3) based on the 4-hour exposure |
| 29 | | duration |
| 30 | | |
| 31 | 30-min AEGL-3: | $C = \sqrt[3]{(5345.05 \text{ mg/m}^3 \cdot \text{min} / 30)} = 10 \text{ mg/m}^3$ |
| 32 | | |
| 33 | 1-h AEGL-3: | $C = \sqrt[3]{(5345.05 \text{ mg/m}^3 \cdot \text{min} / 60)} = 8.1 \text{ mg/m}^3$ |
| 34 | | |
| 35 | 4-h AEGL-3: | $C = 56.27/20 = 5.1 \text{ mg/m}^3$ |
| 36 | | |
| 37 | 8-h AEGL-3: | $C = (5.08 \times 240 \text{ min})/480 \text{ min} = 2.5 \text{ mg/m}^3$ |
| 38 | | |

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APPENDIX B: Category Graph of AEGL Values and Toxicity Data



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Data:

| For Category 0 = No effect, 1 = Discomfort, 2 = Disabling, SL = Some Lethality, 3 = Lethal | | | | |
|--|---------|-------------------|---------|----------|
| Source | Species | mg/m ³ | Minutes | Category |
| NAC/AEGL-1 | | 2.4 | 10 | AEGL |
| NAC/AEGL-1 | | 2.4 | 30 | AEGL |
| NAC/AEGL-1 | | 1.9 | 60 | AEGL |
| NAC/AEGL-1 | | 1.2 | 240 | AEGL |
| NAC/AEGL-1 | | 0.61 | 480 | AEGL |
| NAC/AEGL-2 | | 4.5 | 10 | AEGL |
| NAC/AEGL-2 | | 4.5 | 30 | AEGL |
| NAC/AEGL-2 | | 3.6 | 60 | AEGL |
| NAC/AEGL-2 | | 2.3 | 240 | AEGL |
| NAC/AEGL-2 | | 1.1 | 480 | AEGL |
| NAC/AEGL-3 | | 10 | 10 | AEGL |
| NAC/AEGL-3 | | 10 | 30 | AEGL |
| NAC/AEGL-3 | | 8.1 | 60 | AEGL |
| NAC/AEGL-3 | | 5.1 | 240 | AEGL |

| | | | | |
|--------------|-----|-------|------|------------------------------|
| NAC/AEGL-3 | | 2.5 | 480 | AEGL |
| | | | | |
| Sangha 1983 | rat | 60 | 60 | SL, 1/10 females |
| | rat | 160 | 60 | SL, 3/10 males, 7/10 females |
| | rat | 163 | 60 | SL, 1/10 males |
| | rat | 168 | 60 | SL, 0/10 females |
| | rat | 196 | 60 | SL, 5/10 females |
| | rat | 253 | 60 | SL, 8/10 males |
| | rat | 259 | 60 | SL, 5/10 males, 5/10 females |
| | rat | 319 | 60 | SL, 5/10 males, 9/10 females |
| | | | | |
| Sangha 1984 | rat | 19 | 240 | 2, cholinergic signs |
| | rat | 33 | 240 | SL, 1/10 males, 0/10 females |
| | rat | 56 | 240 | SL, 5/10 males, 4/10 females |
| | rat | 57 | 240 | SL, 3/10 males, 4/10 females |
| | rat | 63 | 240 | SL, 6/10 females |
| | rat | 83 | 240 | SL, 8/10 males, 5/10 females |
| | rat | 173 | 240 | 3, 10/10 females |
| | | | | |
| Pauluhn 1986 | rat | 11.4 | 240 | 1, no clinical signs |
| | rat | 24.3 | 240 | 1, no clinical signs |
| | rat | 45.0 | 240 | 2, tremor |
| | rat | 195.5 | 240 | SL, 1/5 males, 2/5 females |
| | rat | 241.7 | 240 | SL3/5 males, 5/5 females |
| | rat | 350.3 | 240 | SL, 5/5 males, 4/5 females |
| | rat | 1.4 | 360* | 1, no clinical signs |
| | rat | 5.4 | 360* | 1, no clinical signs |
| | rat | 33.1 | 360* | 1, slight tremor |

1 * The 6-hour exposures were repeated for 5 days.

1 **APPENDIX C: Benchmark Concentration Calculation for Methamidophos**

2
3 **Pauluhn 1986, Methamidophos; rats BMCL₀₅**

4
5 =====

6 Probit Model. (Version: 2.8; Date: 02/20/2007)

7 Input Data File: C:\BMDS\UNSAVED1.d

8 Gnuplot Plotting File: C:\BMDS\UNSAVED1.plt

9 Mon Jul 13 14:03:22 2009

10 =====

11 BMDS MODEL RUN

12 ~~~~~

13 The form of the probability function is:

14 $P[\text{response}] = \text{Background} + (1 - \text{Background}) * \text{CumNorm}(\text{Intercept} + \text{Slope} * \text{Log}(\text{Dose}))$, where CumNorm(.) is the cumulative normal

15 distribution function

16 Dependent variable = COLUMN3

17 Independent variable = COLUMN1

18 Slope parameter is not restricted

19 Total number of observations = 7

20 Total number of records with missing values = 0

21 Maximum number of iterations = 250

22 Relative Function Convergence has been set to: 1e-008

23 Parameter Convergence has been set to: 1e-008

24 User has chosen the log transformed model

25 Default Initial (and Specified) Parameter Values

26 Background = 0

27 Intercept = -4.36719

28 Slope = 0.872461

29 Asymptotic Correlation Matrix of Parameter Estimates

30 (*** The model parameter(s) -background have been estimated at a boundary point, or have been

31 specified by the user, and do not appear in the correlation matrix)

32 intercept slope

33 intercept 1 -1

34 slope -1 1

35 Parameter Estimates

36 95.0% Wald Confidence Interval

| Variable | Estimate | Std. Err. | Lower Conf. Limit | Upper Conf. Limit |
|------------|----------|-----------|-------------------|-------------------|
| background | 0 | NA | | |
| intercept | -16.834 | 6.7935 | -30.149 | -3.51897 |
| slope | 3.13988 | 1.2412 | 0.707169 | 5.57258 |

37 NA - Indicates that this parameter has hit a bound implied by some inequality constraint and thus has no

38 standard error.

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Analysis of Deviance Table

| Model | Log(likelihood) | # Param's | Deviance | Test d.f. | P-value |
|---------------|-----------------|-----------|----------|-----------|---------|
| Full model | -14.3635 | 7 | | | |
| Fitted model | -15.1945 | 2 | 1.66197 | 5 | 0.8937 |
| Reduced model | -41.8789 | 1 | 55.0307 | 6 | <.0001 |

AIC: 34.389

Goodness of Fit

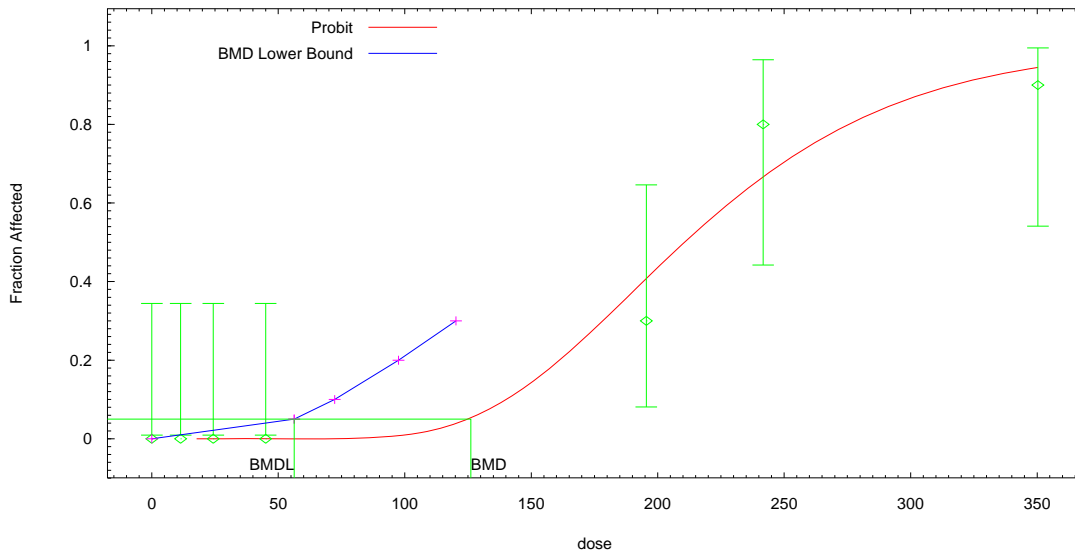
| Dose | Est._Prob. | Expected | Observed | Scaled Size | Residual |
|----------|------------|----------|----------|----------------|----------|
| 0.0000 | 0.0000 | 0.000 | 0 | 10 | 0.000 |
| 11.4000 | 0.0000 | 0.000 | 0 | 10 | -0.000 |
| 24.3000 | 0.0000 | 0.000 | 0 | 10 | -0.000 |
| 45.0000 | 0.0000 | 0.000 | 0 | 10 | -0.002 |
| 195.5000 | 0.3938 | 3.938 | 3 | 10 | -0.607 |
| 241.7000 | 0.6542 | 6.542 | 8 | 10 | 0.969 |
| 350.3000 | 0.9408 | 9.408 | 9 | 10 | -0.547 |

Chi^2 = 1.61 d.f. = 5 P-value = 0.9003

Benchmark Dose Computation

Specified effect = 0.05
 Risk Type = Extra risk
 Confidence level = 0.95
 BMC = 126.153
BMCL₀₅ = 56.2749

Probit Model with 0.95 Confidence Level



14:03 07/13 2009

Pauluhn 1986, Methamidophos; rats BMC₀₁

Probit Model. (Version: 2.8; Date: 02/20/2007)

Input Data File: C:\BMDS\UNSAVED1.d

Gnuplot Plotting File: C:\BMDS\UNSAVED1.plt

Mon Jul 13 14:11:17 2009

BMDS MODEL RUN

The form of the probability function is:

$P[\text{response}] = \text{Background} + (1 - \text{Background}) * \text{CumNorm}(\text{Intercept} + \text{Slope} * \text{Log}(\text{Dose}))$, where
CumNorm(.) is the cumulative normal distribution function

Dependent variable = COLUMN3

Independent variable = COLUMN1

Slope parameter is not restricted

Total number of observations = 7

Total number of records with missing values = 0

Maximum number of iterations = 250

Relative Function Convergence has been set to: 1e-008

Parameter Convergence has been set to: 1e-008

User has chosen the log transformed model

Default Initial (and Specified) Parameter Values

Background = 0
Intercept = -4.36719
Slope = 0.872461

Asymptotic Correlation Matrix of Parameter Estimates

(*** The model parameter(s) -background have been estimated at a boundary point, or have been specified by the user, and do not appear in the correlation matrix)

| | | |
|-----------|-----------|-------|
| | intercept | slope |
| intercept | 1 | -1 |
| slope | -1 | 1 |

Parameter Estimates

| Variable | Estimate | Std. Err. | 95.0% Wald Confidence Interval | |
|------------|----------|-----------|--------------------------------|-------------------|
| | | | Lower Conf. Limit | Upper Conf. Limit |
| background | 0 | NA | | |
| intercept | -16.834 | 6.7935 | -30.149 | -3.51897 |
| slope | 3.13988 | 1.2412 | 0.707169 | 5.57258 |

NA - Indicates that this parameter has hit a bound implied by some inequality constraint and thus has no standard error.

Analysis of Deviance Table

| Model | Log(likelihood) | # Param's | Deviance | Test d.f. | P-value |
|---------------|-----------------|-----------|----------|-----------|---------|
| Full model | -14.3635 | 7 | | | |
| Fitted model | -15.1945 | 2 | 1.66197 | 5 | 0.8937 |
| Reduced model | -41.8789 | 1 | 55.0307 | 6 | <.0001 |
| AIC: | 34.389 | | | | |

Goodness of Fit

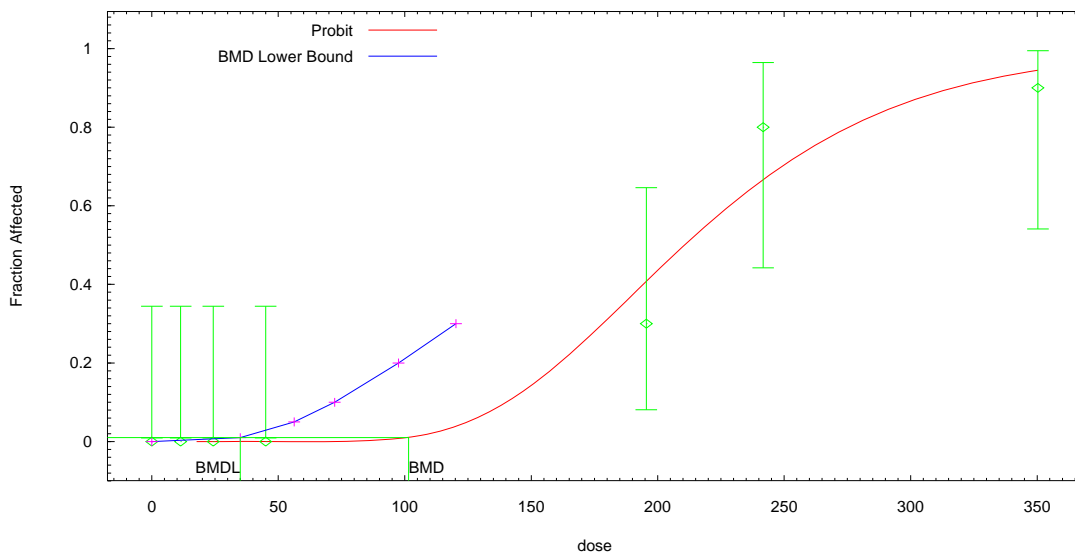
| Dose | Est._Prob. | Expected | Observed | Scaled Size | Residual |
|----------|------------|----------|----------|-------------|----------|
| 0.0000 | 0.0000 | 0.000 | 0 | 10 | 0.000 |
| 11.4000 | 0.0000 | 0.000 | 0 | 10 | -0.000 |
| 24.3000 | 0.0000 | 0.000 | 0 | 10 | -0.000 |
| 45.0000 | 0.0000 | 0.000 | 0 | 10 | -0.002 |
| 195.5000 | 0.3938 | 3.938 | 3 | 10 | -0.607 |
| 241.7000 | 0.6542 | 6.542 | 8 | 10 | 0.969 |
| 350.3000 | 0.9408 | 9.408 | 9 | 10 | -0.547 |

Chi^2 = 1.61 d.f. = 5 P-value = 0.9003

Benchmark Dose Computation

Specified effect = 0.01
 Risk Type = Extra risk
 Confidence level = 0.95
BMC₀₁ = **101.54**
 BMCL = 34.977

Probit Model with 0.95 Confidence Level



14:11 07/13 2009

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APPENDIX D: Derivation Summary for Methamidophos AEGIs
Acute Exposure Guideline Levels For Methamidophos
(CAS Reg. No. 10265-92-6)

| AEGl-1 VALUES | | | | |
|---|-----------------------|-----------------------|-----------------------|------------------------|
| 10-min | 30-min | 1-h | 4-h | 8-hour |
| 2.4 mg/m ³ | 2.4 mg/m ³ | 1.9 mg/m ³ | 1.2 mg/m ³ | 0.61 mg/m ³ |
| Key Reference: Pauluhn, J. 1986. Study for Acute Inhalation Toxicity to the Rat to OECD Guideline No 403. Report No. 15661, Bayer AG Institute of Toxicology, Wuppertal, Germany. | | | | |
| Test Species/Strain/Sex/Number: Rat/Wistar/groups of 5 per sex | | | | |
| Exposure Route/Concentration/Duration: Inhalation/11.4, 24.3, 45.0, 195.5, 241.7, 350.3 mg/m ³ /4 hours | | | | |
| Effects: 11.4 mg/m ³ – no clinical signs; approximately 50% inhibition of plasma cholinesterase activity. 24.3 mg/m ³ - no clinical signs; plasma cholinesterase activity was 36 percent of the control value and erythrocyte cholinesterase was slightly inhibited (92% of the control value). 45.0 mg/m ³ – clinical signs; inhibition of plasma and erythrocyte cholinesterase activity of 67 and 30%, respectively. | | | | |
| Endpoint/Concentration/Rationale: 24.3 mg/m ³ for 4 hours; no clinical signs, plasma cholinesterase activity was 36 percent of the control value and erythrocyte cholinesterase was slightly inhibited (92% of the control value). | | | | |
| Uncertainty Factors/Rationale: Total uncertainty factor: 10 Interspecies: 3, based on rapid metabolism in both rats and humans (Moser 1999; Garofalo et al. 1973). Intraspecies: 3, based on no differences in sensitivity to acetylcholinesterase activity inhibition between juvenile and adult rats in an oral study (Moser 1999). | | | | |
| Modifying Factor: 2, conflicting data sets | | | | |
| Animal to Human Dosimetric Adjustment: Not applicable | | | | |
| Time Scaling: C ⁿ x t = k, where n = 3 and 1 for shorter and longer exposure durations, respectively | | | | |
| Data Adequacy: The two inhalation studies, although well-conducted, provided conflicting data, indicating the difficulty in generating, sampling, and measuring liquid aerosols. For that reason a modifying factor was applied to the data set. The toxicity and mechanism of organophosphate pesticides is well understood. An oral human dosing study provides an estimate of human toxicity. | | | | |

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| AEGL-2 VALUES | | | | |
|---|-----------------------|-----------------------|-----------------------|-----------------------|
| 10-min | 30-min | 1-h | 4-h | 8-h |
| 4.5 mg/m ³ | 4.5 mg/m ³ | 3.6 mg/m ³ | 2.3 mg/m ³ | 1.1 mg/m ³ |
| Key Reference: Pauluhn, J. 1986. Study for Acute Inhalation Toxicity to the Rat to OECD Guideline No 403. Report No. 15661, Bayer AG Institute of Toxicology, Wuppertal, Germany. | | | | |
| Test Species/Strain/Number: Rat/Wistar/groups of 5 per sex | | | | |
| Exposure Route/Concentration/Duration: Inhalation/11.4, 24.3, 45.0, 195.5, 241.7, 350.3 mg/m ³ /4 hours | | | | |
| Effects: 11.4 mg/m ³ – no clinical signs; approximately 50% inhibition of plasma cholinesterase activity. 24.3 mg/m ³ - no clinical signs; plasma cholinesterase activity was 36 percent of the control value and erythrocyte cholinesterase was slightly inhibited (92% of the control value). 45.0 mg/m ³ – clinical signs; inhibition of plasma and erythrocyte cholinesterase activity of 67 and 30%, respectively. | | | | |
| Endpoint/Concentration/Rationale: 45.0 mg/m ³ for 4 hours – clinical signs of tremor and reduced motility; inhibition of plasma and erythrocyte cholinesterase activity of 67 and 30%, respectively. | | | | |
| Uncertainty Factors/Rationale: Total uncertainty factor: 10 Interspecies: 3, based on rapid metabolism in both rats and humans (Moser 1999; Garofalo et al. 1973). Intraspecies: 3, based on no differences in sensitivity to acetylcholinesterase activity inhibition between juvenile and adult rats in an oral study (Moser 1999). | | | | |
| Modifying Factor: 2, conflicting data sets | | | | |
| Animal to Human Dosimetric Adjustment: Not applicable | | | | |
| Time Scaling: C ⁿ x t = k, where n = 3 and 1 for shorter and longer exposure durations, respectively | | | | |
| Data Adequacy: The two inhalation studies, although well-conducted, provided conflicting data, indicating the difficulty in generating, sampling, and measuring liquid aerosols. For that reason a modifying factor was applied to the data set. The toxicity and mechanism of organophosphate pesticides is well understood. An oral human dosing study provides an estimate of human toxicity. | | | | |

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| AEGL-3 VALUES | | | | |
|---|----------------------|-----------------------|-----------------------|-----------------------|
| 10-min | 30-min | 1-h | 4-h | 8-h |
| 10 mg/m ³ | 10 mg/m ³ | 8.1 mg/m ³ | 5.1 mg/m ³ | 2.5 mg/m ³ |
| Key Reference: Pauluhn, J. 1986. Study for Acute Inhalation Toxicity to the Rat to OECD Guideline No 403. Report No. 15661, Bayer AG Institute of Toxicology, Wuppertal, Germany. | | | | |
| Test Species/Strain/Number: Rat/Wistar/groups of 5 per sex | | | | |
| Exposure Route/Concentration/Duration: Inhalation/11.4, 24.3, 45.0, 195.5, 241.7, 350.3 mg/m ³ /4 hours | | | | |
| Effect: Mortalities: 0/10, 0/10, 0/10, 3/10, 8/10, 9/10, respectively | | | | |
| Endpoint/Concentration/Rationale: the 4-hour BMC ₀₁ , 101.54 mg/m ³ , estimated as the threshold for lethality | | | | |
| Uncertainty Factors/Rationale: | | | | |
| Total uncertainty factor: 10 | | | | |
| Interspecies: 3, based on rapid metabolism in both rats and humans (Moser 1999; Garofalo et al. 1973). | | | | |
| Intraspecies: 3, based on no differences in sensitivity to acetylcholinesterase activity inhibition between juvenile and adult rats in an oral study (Moser 1999). | | | | |
| Modifying Factor: 2, conflicting data | | | | |
| Animal to Human Dosimetric Adjustment: Not applicable | | | | |
| Time Scaling: C ⁿ x t = k, where n = 3 and 1 for shorter and longer exposure durations, respectively. | | | | |
| Data Adequacy: The two inhalation studies, although well-conducted, provided conflicting data, indicating the difficulty in generating, sampling, and measuring liquid aerosols. For that reason a modifying factor was applied to the data set. The toxicity and mechanism of organophosphate pesticides is well understood. An oral human dosing study provides an estimate of human toxicity. | | | | |

2