

## 2 Objective

The objective of this study was to demonstrate that method EL-001-W08-01 ("An Analytical Method for the Determination of Residues of BYF 14182 And Its Metabolites BYF 14182-3-hydroxy-butyl and BYF 14182-pyrazolyl-AAP in Water Using LC/MS/MS") can be performed with acceptable recoveries for determination of the compounds BYF 14182, BYF 14182-3-hydroxy-butyl and BYF 14182-pyrazolyl-AAP at an independent laboratory having no prior experience with the method. The method was developed by Bayer CropScience Residue and Environmental Chemistry, Stilwell, USA, and reported as Method EL-001-W08-01, by Jami M. Wade, dated May 27, 2009. River Rhine water (taken at Leverkusen-Hitdorf, Germany) and tap water Monheim, Germany were chosen as representative matrices for validation within the present study.

This study was performed in accordance with US EPA Ecological Effects Test Guidelines, OPPTS 850.7100 Data Reporting for Environmental Chemistry Methods, EPA 712-C-96-348, April 1996.

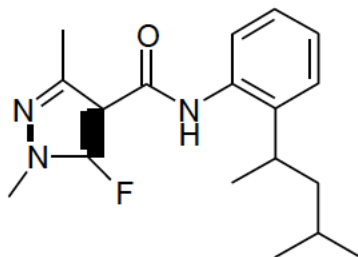
## 3 Materials

### 3.1 Test and Reference Items

**Code Name:** BYF 14182 (Active Ingredient, Parent Molecule)

**CAS Name:** N-[2-(1,3-dimethylbutyl)phenyl]-5-fluoro-1,3-dimethyl-1H-pyrazole-4-carboxamide

**Structural formula:**



**Molecular Formula:** C<sub>18</sub>H<sub>24</sub>F N<sub>3</sub> O

**Molecular Weight:** 317 g/mol

**Certificate of analysis:** AZ 15302

**Origin Batch No.:** NLL7306-17

**Purity:** 99.5%

**Expiry date:** 2011-09-17

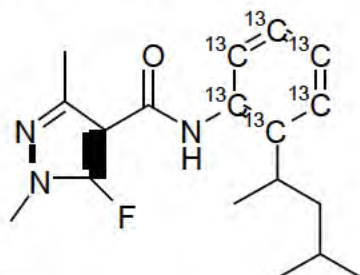
**Appearance:** light beige powder

**Code Name:** **BYF 14182 [phenyl-<sup>13</sup>C<sub>6</sub>]**

(Parent Molecule, Labeled Internal Standard)

**IUPAC Name:** N-[2-(1,3-dimethylbutyl)phenyl]-5-fluoro-1,3-dimethyl-1H-pyrazole-4-carboxamide

**Structural formula:**



**Molecular Formula:** <sup>13</sup>C<sub>6</sub>C<sub>12</sub>H<sub>24</sub>F N<sub>3</sub>O

**Molecular Weight:** 323 g/mol

**Standard No.:** KML3450-1-6

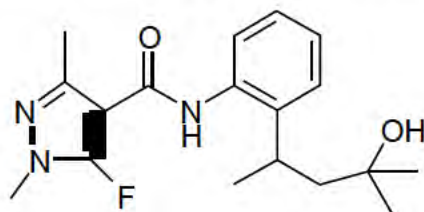
**Purity:** >98%

**Certification date:** 2004-10-21

**Code Name:** **BYF14182-3-hydroxy-butyl** (Metabolite)

**CAS Name** 5-fluoro-N-[2-(3-hydroxy-1,3-dimethylbutyl)phenyl]-1,3-dimethyl-1H-pyrazole-4-carboxamide

**Structural formula:**



**Molecular Formula:** C<sub>18</sub>H<sub>24</sub>F N<sub>3</sub>O<sub>2</sub>

**Molecular Weight:** 333 g/mol

**Certificate of analysis:** AZ 15083

**Origin Batch No.:** SES 10139-3-3

**Purity:** 99.2%

**Expiry date:** 2010-06-20

**Appearance:** light beige powder

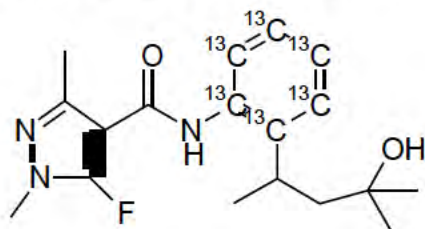
**Code Name:** **BYF14182-3-hydroxy-butyl-<sup>13</sup>C<sub>6</sub>**

BCS-AA10006-[phenyl-<sup>13</sup>C<sub>6</sub>]

(Metabolite, Isotopic Internal Standard)

**CAS Name** 5-fluoro-N-[2-(3-hydroxy-1,3-dimethylbutyl)phenyl]-1,3-dimethyl-1H-pyrazole-4-carboxamide

**Structural formula:**



**Molecular Formula:** <sup>13</sup>C<sub>6</sub>C<sub>12</sub> H<sub>24</sub> F N<sub>3</sub> O<sub>2</sub>

**Molecular Weight:** 339 g/mol

**Batch No.:** KML 3694-2-30

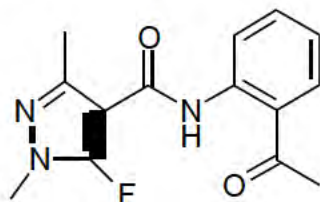
**Purity:** >99 %

**Certification date:** 2006-09-19

**Code Name:** **BYF14182-pyrazoly-AAP (Metabolite)**

**CAS Name:** N-(2-acetylphenyl)-5-fluoro-1,3-dimethyl-1H-pyrazole-4-carboxamide

**Structural formula:**



**Molecular Formula:** C<sub>14</sub> H<sub>14</sub> F N<sub>3</sub> O<sub>2</sub>

**Certificate of analysis:** AZ 14665

**Molecular Weight:** 275 g/mol

**Origin Batch No.:** KTS10231-2-3

**Purity:** 98.6%

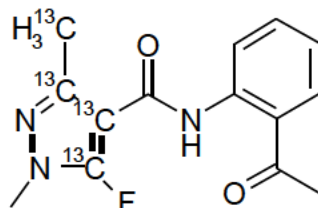
**Expiry date:** 2010-12-08

**Appearance:** beige powder

**Code Name:** **BYF14182-pyrazolyl-AAP-<sup>13</sup>C<sub>4</sub>**  
 [3-methyl-<sup>13</sup>C,pyrazolyl-<sup>13</sup>C<sub>3</sub>] BCS-AF73126  
 (Metabolite, Isotopic Internal Standard)

**CAS Name:** N-(2-acetylphenyl)-5-fluoro-1,3-dimethyl-1H-pyrazole-4-carboxamide

**Structural formula:**



**Molecular Formula:** <sup>13</sup>C<sub>4</sub>C<sub>10</sub> H<sub>14</sub> F N<sub>3</sub> O<sub>2</sub>

**Molecular Weight:** 279 g/mol

**Batch No.:** KML 3937-1-15

**Purity:** > 99%

**Expiry date:** 2010-05-05

More details about the reference items are documented in the raw data. For handling of the reference items and for preparation of standard solutions see SOP 1001 and SOP 1014.

### 3.2 Test System

The method was validated using river Rhine and tap water Monheim. Two different water types were used in order to assess a possible influence of different water characteristics. The water samples were analysed for TOC, DOC, conductivity, water hardness, dry residue after filtration and pH by Currenta, 51368 Leverkusen, Germany. Water types are summarised in Table 1. Complete water parameters are reported in Table 13 and Table 14.

Table 1: Water Types

| Water Type    | Source of Water   |
|---------------|-------------------|
| Surface Water | River Rhine Water |
| Tap Water     | Tap Water Monheim |

## 4 Experimental

### 4.1 Analytical Method

The recovery data for the study were generated using the following method, which gives full details of preparing the analytical sample extracts and the conditions for high performance liquid chromatography (HPLC):

Number of the method: EL-001-W08-01  
 Title of the method: An Analytical Method for the Determination of Residues of BYF 14182 And Its Metabolites BYF 14182-3-hydroxy-butyl and BYF 14182-pyrazolyl-AAP in Water Using LC/MS/MS  
 Author of the method: Jami M. Wade  
 Bayer CropScience  
 Residue and Environmental Chemistry  
 17745 South Metcalf Avenue  
 Stilwell, Kansas 66085  
 Reference: Method EL-001-W08-01  
 Limit of quantitation: 0.1 µg/L

The following sample sets were analysed:

**Table 2: Level and Number of Recoveries per Fortification Level**

| Water             | Control sample | Level 0.1 µg/L | Level 1.0 µg/L |
|-------------------|----------------|----------------|----------------|
| River Rhine       | 2              | 5              | 5              |
| Tap Water Monheim | 2              | 5              | 5              |

#### 4.1.1 Outline of the Method

Appropriate volumes of fortification and internal standard solutions were added to the water samples. After mixing the analysis was performed by LC-MS/MS.

#### 4.1.2 Instruments

Liquid Chromatograph: 1200 Column Compartment G1316B  
 1200 Binary Pump G1312B Bin Pump SL  
 1200 Isocratic Pump G1310A  
 1200 Degasser G1322A  
 Agilent Technologies, 40880 Ratingen, Germany

Autosampler: HTC PAL System  
 CTC Analytics AG  
 4222 Zwingen, Switzerland

Mass Spectrometer: Applied Biosystems API 4000 with turbo ionspray interface mass selective detector (MS/MS)  
Note: Some mass spectrometric conditions are instrument specific. The spectrometric conditions were optimised by a competent operator prior to analysis.

#### 4.1.3 Reagents and Equipment

Column (HPLC): YMC Pro C18 , length 33mm x 4.0mm, Particle size 3µm, pore size 1200A  
Series No.: 90306 14839

Acetonitrile: for HPLC, optigrade  
LGC Promochem GmbH, UN 1648  
46485 Wesel, Germany

Acetic acid (100%): Suprapur  
Merck KGaA, No. 1.00066.1000  
64271 Darmstadt, Germany

Deionized water: purified in a Milli-Q unit  
Milli-Pore GmbH  
65731 Eschborn, Germany

Volumetric flasks, pipettes and other equipment commonly used in the laboratory.

#### 4.1.4 Liquid Chromatographic Conditions

Liquid chromatographic conditions were almost identical to those described in the original method report EL-001-W08-01. Only the HPLC gradient had to be corrected since there were two typographical errors in the original description.

**Table 3: Liquid Chromatographic Conditions**

Column: YMC Pro C18, length 33mm x 4mm, pore size 120A

Particle size: 3 µm

Oven temperature: 60 °C

Injection volume: 80 µL

Run time: 4 minutes

Mobile phase: A: deionized water / acetonitrile (9/1 v/v) (+ 0.01% acetic acid)

B: acetonitrile (+ 0.01% acetic acid)

Retention times: approx. 0.85 min for BYF 14182-3-hydroxy-butyl

approx. 0.93 min for BYF 14182-pyrazolyl-AAP

approx. 1.45 min for BYF 14182

**Table 4: HPLC Gradient**

| Time [min] | Flow [µL/min] | A [%] | B [%] |
|------------|---------------|-------|-------|
| 0.00       | 1000          | 80    | 20    |
| 0.10       | 1000          | 40    | 60    |
| 1.00       | 1000          | 40    | 60    |
| 2.20       | 1000          | 25    | 75    |
| 2.30       | 1000          | 10    | 90    |
| 3.00       | 1000          | 10    | 90    |
| 3.01       | 1000          | 80    | 20    |
| 4.00       | 1000          | 80    | 20    |

**Table 5: Valco Valve Method Properties**

| Time [min] |                                     |
|------------|-------------------------------------|
| 0.0        | switch eluent stream into waste     |
| 0.5        | switch eluent stream into interface |
| 3.8        | switch eluent stream into waste     |

These switching times were also adjusted to the used equipment.

#### 4.1.5 Mass Spectrometric Parameters

MS/MS parameter settings were in general as described in method EL-001-W08-01 but optimized for the instrument being used. The dwell time was reduced to 40 msec for all analytes in the second period.

#### 4.1.6 Calculation

For calculation of the concentrations, six-point calibration curves were used. These curves were calculated using linear regression automatically after each sequence run with the Applied Biosystems quantitation software Analyst (vers. 1.5). Further calculations were performed using the software MS-EXCEL 2003. The results given are rounded values. Thus, rounding "errors" may occur if recalculations are made using the listed figures.

##### 4.1.6.1 Calculation of Analyte Concentrations

NOTE: Evaluation is performed according to the linearity standard procedure.

1. Calculate the response factors (peak area of detected analyte / peak area of the internal standard) of all standard injections and calculate the resulting linearity of the analyte.
2. Determine the response factor (peak area of detected analyte / peak area of the internal standard) for the sample. This value will be used as x.
3. Calculate the residue level in µg/L as follows:

$$R = (x - b) / a$$

R: Determined concentration of analyte in µg/L  
x: Response factor of the sample  
b: Interception from linear regression  
a: Slope from linear regression

##### 4.1.6.2 Calculation of Recoveries

1. Calculate the concentration in the recovery sample according to 4.1.6.1.
2. Calculate the percent recovery as follows:

$$\text{Recovery} = \frac{\text{Concentration} \times 100}{\text{Fortification Level}}$$

Recovery: Recovered concentration of analyte in % found in the fortified sample  
Concentration: Concentration in the fortified sample in µg/L determined according to 4.1.6.1  
Fortification level: Fortified concentration of analyte in µg/L



#### **4.1.7 Deviations from the Method**

Within the analytical procedure for determination of BYF 14182 and its metabolites BYF 14182-3-hydroxy-butyl and BYF 14182-pyrazolyl-AAP only minor details have been changed.

The HPLC gradient had to be corrected since there were two typographical errors in the original description. The dwell time was reduced to 40 msec for all analytes in the second period.