**Test Material:** 2,4-DP-p and its transformation products: 2,4-DCP, 2,4-DCA and 2-EHE

**MRID:** 44243401

Independent Laboratory Method Trials of a Residue Analytical Method Title: for Dichlorprop-P (2,4-DP-p) and Metabolites in Soil.

**EPA PC Code:** 031402

**OCSPP** Guideline: 850.6100

For CDM Smith

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#### Analytical method for 2,4-DP-p and its transformation products, 2,4-DCP, 2,4-DCA and 2-EHE in soil

Reports:	MRID No. 44243401. Bruns, G., S. Nel Method Trials of a Residue Analytical I and Metabolites in Soil. Report prepare Edmonton, Alberta, Canada; sponsored FORCE, BASF Aktiengisellschaft, Lim 95ETL12.PRO. BASF Project No.: TSF Report No.: BSF06.REP. Final report is	Method for Dichlorprop-P (2,4-DP-p) d by Enviro-Test Laboratories, and submitted by 2,4-DP-p TASK burgerhof, Germany. Protocol No.: KF9301 (BASF/Adpen Labs). ETL
<b>Document No.:</b>	MRID 44243401	
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Statements: Classification:	ECM: The study was conducted compli CFR Part 160; Appendix 5, p. 123). Sig Authenticity and Quality Assurance stat pp. 123-124, 139). A No Data Confident ILV: The study was conducted in accor (40 CFR Part 160; p. 3). Signed and dat Quality Assurance statements were prov Authenticity was not included. This analytical method study provides u <b>supplemental</b> . However, performance of requirements for multiple soil/analyte e Only three replicate samples were fortif each test soil in the ECM. In the ILV, the and 2-EHE differed from the nominal L	ned and dated GLP, Certification of tements were provided (Appendix 5, atiality statement was not provided. dance with the USEPA FIFRA GLP ed No Data Confidentiality, GLP and vided (pp. 2-4, 21). A Certification of useful information and is classified as data did not meet guideline xperiments in the ECM and ILV. Fied at the LOQ and 10x LOQ for ne fortification levels for 2,4-DP-p
	soil matrices were inadequately character chromatogram sets were not provided for can be found in comments section.	
PC Code:	031402	
<b>Reviewer:</b>	Faruque Khan, Ph.D. Senior Fate Scientist	Signature: Date: May 12, 2014

## **Executive Summary**

This Analytical Method Number TSKF9301 (from Adpen Labs), is designed for the quantitative determination of 2,4-DP-p and its transformation products 2,4-DCP, 2,4-DCA and 2-EHE in soil using GC/MS. For the ECM, the reviewer could not adequately determine whether the method is quantitative for each analyte for each soil matrix because only three replicates were analyzed at the LOQ and 10x LOQ. However, the mean recovery ( $\pm$ SD) of 2,4-DP-p at the LOQ, across all five soils, was 65.7  $\pm$  4.9% (n=14), which is outside of the guideline requirement. Across all test soils, mean recoveries of 2,4-DCP, 2,4-DCA and 2-EHE at the LOQ and of all analytes at 10x and 200x LOQ were within the guideline requirement for mean recoveries. The lowest toxicological level of concern in soil was not reported; however, 2,4-DCP is listed in the EPA Regional Screening Levels with a screening level of 18 mg/kg in residential soil which is much

higher than the reported LOQ for this method (no other analyte was listed). In the ILV, performance data for all analytes were acceptable at both the LOQ and 10x LOQ in the California, New York and Washington soils (excluding high recovery of 2,4-DCP in California and New York soils at 10x LOQ and high RSD values for 2,4-DP-p in New York soil at the LOQ and 10x LOQ). Recoveries were low for multiple analytes in the Georgia and Indiana soils (see DER Table 3). No major modifications were made by the independent laboratory.

	MRID						Limit of
Analyte(s) by Pesticide	Environmental Chemistry Method	Independent Laboratory Validation	Matrix	Method Date	Registrant	Analysis	Quantitation (LOQ)
2,4-DP-p and its transformation products 2,4- DCP, 2,4- DCA and 2- EHE	44243401	44243401	Soil	10/01/1994	BASF Aktiengisellschaft, Germany	GC/MS	0.01 mg/kg (0.01 ppm)

**Table 1. Analytical Method Summary** 

# I. Principle of the Method

Samples (10.0 g) were measured into 50-mL screw-cap polypropylene centrifuge tubes (Appendix 5, pp. 128, 131-134). The samples were vortexed with 20 mL of 5% acetic acid in methanol at the highest speed for ca. 30 seconds. Cavitation of the samples was performed via sonication in an ultrasonic water bath at room temperature for 20 minutes. After centrifugation at ca. 2000 rpm for ca. 10 minutes, the supernatant was decanted into a 500 mL glass screw-cap bottle. The vortex/sonication/centrifuge/decant procedure was sequentially repeated with 20 mL of 5% acetic acid in methanol:water (1:1, v:v) then 20 mL of 10% acetone in 0.5 M KCl/0.1M NaOH. The combined supernatants were mixed with ca. 430 mL of deionized water then the sample was acidified with 2.5 mL of phosphoric acid (ca. 85%). After shaking by hand for ca. 30 seconds, the pH was measured to ensure that it was less than 2. The sample was purified using a preconditioned C<sub>18</sub> solid phase extraction (SPE) column (1 g, 6 mL; preconditioned with 10 mL methanol then 10 mL of 1.5% phosphoric acid in water). The sample was passed through the column under vacuum, and the eluates were discarded. The analytes were removed from the column via elution with 9 mL of 2% acetone in hexane (Fraction A, containing 2,4-DCP, 2,4-DCA and 2-EHE) and 9 mL of 50% methanol in acetone (Fraction B, containing 2,4-DP-p). Fraction B was mixed with 1.0 mL of hexane then reduced to 0.5-1.0 mL using a N-Evap and a room temperature water bath. The residue of Fraction B was derivitized by mixing with 1.0 mL of 14% BF<sub>3</sub>/methanol solution in a capped tube. The sample tube was transferred to a water bath at  $70 \pm 2^{\circ}$ C for 30 minutes. After cooling, 8 mL of distilled water and 5 mL of hexane were added to the sample. After shaking for 10 minutes at high speed using a mechanical shaker, the aqueous layer was discarded. The remaining hexane layer of Fraction B and the rinse of the transfer pipet were combined with Fraction A. This mixture was reduced to 1 mL using a N-Evap and a room temperature water bath. The residue was vortexed then diluted as necessary for GC/MS analysis.

Samples were analyzed for 2,4-DP-p, 2,4-DCP, 2,4-DCA and 2-EHE by gas chromatography (HP-5 MS, 30 m x 0.25 mm, 0.25- $\mu$ m column) with mass spectrometry (two different MS conditions provided; Appendix 5, Tables IIa-IIb, pp. 142-143). Three or one qualifier ions were monitored for methylated 2,4-DP-p (164, 191 and 248 m/z or only 164 m/z) and 2-EHE (189, 234 and 346 m/z or only 189 m/z). One qualifier ion was monitored for 2,4-DCP (164 m/z) and 2,4-DCA (176 m/z). No confirmation method was used; however, the ECM study authors suggested that a GC column with a different polarity, such as a HP-1701, could be used for confirmation (Appendix 5, p. 135). Injection volume was 1 or 2  $\mu$ L (Appendix 5, p. 134).

In the ECM, the LOQ and LOD were 0.01 mg/kg and 0.005 mg/kg, respectively (Appendix 5, p. 138). In the ILV, the LOQ was reported as 0.01 ppm, but no LOD was reported (p. 20).

# **II. Recovery Findings**

ECM (MRID 44243401): It could not be adequately determined whether the method is quantitative for each analyte for each soil matrix because an insufficient number of replicates were analyzed at the LOQ and 10x LOQ (three replicates each). Based on only three replicates, the method was quantitative (mean 70-120%; RSD  $\leq$ 20%) for 2,4-DP-p at the LOQ in New York soil only and at 10x LOQ in California and New York soils only; for 2,4-DCP at the LOQ in Georgia and Indiana soils and at 10x LOQ in Georgia, Indiana and Washington soils; and for 2,4-DCA and 2-EHE in all soils at the LOQ and 10xLOQ (excluding 2,4-DCA at the LOQ in Indiana soil; Appendix 5, Tables IIIa-IIIc, pp. 144-146; DER Attachment 2). The mean recovery ( $\pm$  SD) of 2,4-DP-p at the LOQ, across all five soils, was 65.7  $\pm$  4.9% (n=14), which is outside of the guideline requirement. Across all test soils, mean recoveries of 2,4-DCP, 2,4-DCA and 2-EHE at the LOQ and of all analytes at 10x and 200x LOQ were within the guideline requirement for mean recoveries. Soil matrix characterization was not reported (Appendix 5, pp. 136-137). Analyte identification was based on the observation of the MS target ions and qualifier ions; no confirmation method was used (Appendix 5, pp. 135, 137).

ILV (MRID 44243401): For 2,4-DP-p, mean recoveries and relative standard deviations (RSD) were within guideline requirements (mean 70-120%; RSD ≤20%) for analysis at 0.0093 mg/kg in the California, Indiana and Washington soils, at 0.093 mg/kg in the California and Washington soils, and at 0.93 mg/kg in the California, New York and Washington soils; however, recoveries at 0.0093 mg/kg (LOQ) in the Georgia and New York soils, at 0.093 mg/kg in the Georgia, Indiana and New York soils, and at 0.93 mg/kg in the Georgia and Indiana soils did not meet guideline requirements (Tables 3A-7B, pp. 22-31). For 2,4-DCP, mean recoveries and relative standard deviations (RSD) were within guideline requirements for analysis at 0.01 mg/kg (LOQ) in the California, Indiana, New York and Washington soils, at 0.10 mg/kg in the Georgia, Indiana and Washington soils, and at 1.0 mg/kg in the California and New York soils; however, recoveries at 0.01 mg/kg (LOQ) in the Georgia soil, at 0.10 mg/kg in the California and New York soils, and at 1.0 mg/kg in the Georgia, Indiana and Washington soils did not meet guideline requirements. For 2,4-DCA, mean recoveries and relative standard deviations (RSD) were within guideline requirements for analysis at 0.01 mg/kg (LOQ) in all five soils, at 0.10 mg/kg in the California, Indiana, New York and Washington soils, and at 1.0 mg/kg in the California, Georgia, New York and Washington soils; however, recoveries at 0.10 mg/kg in the Georgia soil and at 1.0 mg/kg in the Indiana soil did not meet guideline requirements. For 2-EHE, mean

recoveries and relative standard deviations (RSD) were within guideline requirements for analysis at 0.012 mg/kg (LOQ) in the California, Indiana, New York and Washington soils, at 0.12 mg/kg in the California, New York and Washington soils, and at 1.2 mg/kg in the New York soil; however, recoveries at 0.012 mg/kg (LOQ) in the Georgia soil, at 0.12 mg/kg in the Georgia and Indiana soils, and at 1.2 mg/kg in the California, Georgia, Indiana and Washington soils did not meet guideline requirements.

Soil matrices were characterized as follows: loam soil from New York, sand soil from Washington, loamy sand soil from Georgia, sandy loam/loamy sand soil from California and loam/clay loam soil from Indiana; all soils were collected from the 0-12" soil layer from 2,4-DP-p terrestrial field dissipation sites (p. 14). The method was validated with the first trial for the California and New York soils, the second trial for the Georgia and Washington soils and the third trial for the Indiana soil; data from several unacceptable calibration curves was also provided (p. 20; Appendix 6, pp. 164-202).

Analyte		Number		Mean	Standard	<b>Relative Standard</b>
Analyte	Level (mg/kg)	of Tests	Range (%)	Recovery (%)	<b>Deviation</b> (%)	<b>Deviation</b> (%)
			California s	oil		
	0.01 (LOQ)	3	60.9-66.6	63.9	2.9	4.5
Methylated 2,4-DP-p	0.10	3	69.9-90.2	83.1	11.4	13.8
	2.0	2	82.6-92.6	87.6	7.1	8.1
	0.01 (LOQ)	3	122.0-134.0	129.8	6.8	5.2
2,4-DCP	0.10	3	129.0-131.0	130.3	1.2	0.9
	2.0	2	68.8-72.5	70.7	2.6	3.7
	0.01 (LOQ)	3	89.0-96.5	92.0	4.0	4.3
2,4-DCA	0.10	3	100.5-118.5	111.0	9.4	8.4
	2.0	2	99.4-111.9	105.7	8.8	8.4
	0.01 (LOQ)	3	87.5-106.0	93.8	10.5	11.2
2-EHE (2,4-DP-p 2-EHE)	0.10	3	86.5-105.0	97.3	9.6	9.9
(2,4-DГ-р 2-ЕПЕ)	2.0	2	89.4-99.4	94.4	7.1	7.5
	•		Georgia so	il		·
	0.01 (LOQ)	21	59.9-61.4	60.7	1.1	1.7
Methylated 2,4-DP-p	0.10	3	62.8-70.3	66.4	3.8	5.7
	2.0	2	62.5-81.4	72.0	13.4	18.6
	0.01 (LOQ)	3	94.5-98.5	95.8	2.3	2.4
2,4-DCP	0.10	3	95.0-109.5	101.8	7.3	7.2
	2.0	$1^{1}$	91.9	91.9		
	0.01 (LOQ)	3	79.0-88.0	84.3	4.7	5.6
2,4-DCA	0.10	3	84.5-97.0	92.5	6.9	7.5
	2.0	2	73.1-96.9	85.0	16.8	19.8
	0.01 (LOQ)	3	76.5-93.5	84.8	8.5	10.0
2-EHE (2,4-DP-p 2-EHE)	0.10	3	83.5-113.0	96.0	15.3	15.9
(2,4-DP-p 2-EHE)	2.0	2	78.8-91.9	85.4	9.3	10.9
			Indiana so	il		
Mathulated 2.4 DD -	0.01 (LOQ)	3	60.4-64.7	63.1	2.4	3.7
Methylated 2,4-DP-p	0.10	3	62.3-67.5	65.8	3.0	4.6

 Table 2. Initial Validation Method Recoveries for 2,4-DP-p and Its Transformation products in Soil\*

Analyte	Fortification Level (mg/kg)	Number of Tests	Recovery Range (%)	Mean Recovery (%)	Standard Deviation (%)	<b>Relative Standard</b> <b>Deviation (%)</b>
	2.0	2	63.1-70.2	66.7	5.0	7.5
	0.01 (LOQ)	3	111.5-121.5	115.7	5.2	4.5
2,4-DCP	0.10	3	99.0-107.5	103.7	4.3	4.2
	2.0	2	81.3-88.1	84.7	4.8	5.7
	0.01 (LOQ)	3	65.0-72.0	69.3	3.8	5.5
2,4-DCA	0.10	3	79.1-84.6	82.1	2.8	3.4
	2.0	2	83.1-84.3	83.7	0.8	1.0
	0.01 (LOQ)	3	85.0-105.0	95.8	10.1	10.5
2-EHE (2,4-DP-p 2-EHE)	0.10	3	78.5-86.5	82.5	4.0	4.8
(2, <b>4-DI -</b> p 2-EIIE)	2.0	2	72.5-74.4	73.5	1.3	1.8
			New York s	oil		
	0.01 (LOQ)	3	64.7-75.0	70.3	5.2	7.4
Methylated 2,4-DP-p	0.10	3	72.7-76.9	74.6	2.1	2.9
	2.0	2	82.6-84.4	83.5	1.3	1.5
	0.01 (LOQ)	3	124.0-135.0	131.3	6.4	4.8
2,4-DCP	0.10	3	106.5-122.0	115.7	8.1	7.0
	2.0	2	92.5-101.9	97.2	6.6	6.8
	0.01 (LOQ)	3	89.0-107.5	100.7	10.2	10.1
2,4-DCA	0.10	3	91.5-106.0	99.2	7.3	7.3
	2.0	2	92.5-96.3	94.4	2.7	2.8
2-EHE	0.01 (LOQ)	3	93.5-105.0	100.8	6.4	6.3
(2,4-DP-p 2-EHE)	0.10	3	80.5-93.5	87.8	6.7	7.6
(2,1 DI p 2 EIIE)	2.0	2	82.5-87.5	85.0	3.5	4.2
			Washington s	soil		
	0.01 (LOQ)	3	63.2-73.6	68.9	5.3	7.7
Methylated 2,4-DP-p	0.10	3	63.2-65.1	64.2	1.0	1.5
	2.0	2	73.8-77.3	75.6	2.5	3.3
	0.01 (LOQ)	3	116.5-127.0	121.3	5.3	4.4
2,4-DCP	0.10	3	117.3-122.3	119.5	2.6	2.1
	2.0	2	89.9-119.3	104.6	20.8	19.9
	0.01 (LOQ)	3	92.0-98.0	96.0	3.5	3.6
2,4-DCA	0.10	3	92.5-99.5	96.7	3.7	3.8
	2.0	2	95.0-101.3	98.2	4.5	4.5
2-EHE	0.01 (LOQ)	1 <sup>1</sup>	107.5	107.5		
2-EHE (2,4-DP-p 2-EHE)	0.10	3	86.0-91.5	89.5	3.0	3.4
	2.0	2	94.4-96.9	95.7	1.8	1.8

\* Data were obtained from Appendix 5, Tables IIIa-IIIc, pp. 144-146 in the study report. Reported values for mean recovery, standard deviation and relative standard deviation were reviewer-calculated because the study authors only provided these values for the entire data set at each fortification level. The values which were outside of guideline requirements are **bolded**.

1 The number of tests equalled 1 or 2, instead of 3, because 1 or 2 procedural recoveries were not included in the data set in the study report due to matrix interferences.

Analyte	Fortification Level (mg/kg)			Mean Recovery (%)	Standard Deviation (%)	Relative Standard Deviation (%)
			-	fornia; 0-12"; (		Deviation (70)
	$0.0093 (LOQ^1)$	÷	81-118	101 101	16	16
2,4-DP-p	0.093	6	73-105	90	10	11
<b>-</b> ,. <b>D</b> 1 p	0.93	6	86-118	97	12	12
	0.010 (LOQ)	6	100-130	117	10	8.5
2,4-DCP	0.10	6	120-150	135	14	10
	1.0	6	88-130	108	14	13
	0.010 (LOQ)	6	78-96	89	7.3	8.2
2,4-DCA	0.10	6	67-100	81	11	14
2,4-DCA	1.0	6	70-100	80	11	14
	$0.012 (LOQ^1)$	6	76-83	80	2.4	3.0
2-EHE	0.012 (LOQ )	6	65-92	76	9.0	12
(2,4-DP-p 2-EHE)	1.2	6	63-72	<b>68</b>	3.2	4.7
				0-12"; GR948		7.7
	0.0093 (LOQ <sup>1</sup> )	6	55-118	83	24	29
2,4-DP-p	0.093	6	58-73	65	5.3	8.2
2,4 DI p	0.93	6	60-75	<u>69</u>	5.4	7.8
	0.010 (LOQ)	6	47-120	93	32	39
2,4-DCP	0.10	6	57-88	79	11	14
2,4 DCI	1.0	6	52-75	60	9.2	15
	0.010 (LOQ)	4 <sup>2</sup>	79-90	84	5.2	6.2
2,4-DCA	0.10	5 <sup>3</sup>	64-72	60 60	19	32
2,4-DCA	1.0	6	68-81	75	4.9	7
	0.012 (LOQ <sup>1</sup> )	6	42-92	69	20	29
2-EHE	0.012 (LOQ )	6	46-83	64	17	27
(2,4-DP-p 2-EHE)	1.2	6	45-92	65	19	29
				03 a; 0-12"; GR94		27
	0.0093 (LOQ <sup>1</sup> )	5 <sup>4</sup>	62-95	a, 0-12, 0104 77	12	16
2,4-DP-p	0.093	6	52-67	59	4.9	8.3
2, <b>4-D</b> 1-p	0.93	6	44-71	<u> </u>	9.6	16
	0.010 (LOQ)	6	75-87	79	4.9	6.2
2,4-DCP	0.10	6	71-80	73	3.5	4.8
2,4-DCI	1.0	6	63-74	69	4.4	6.4
	0.010 (LOQ)	6	68-81	73	4.9	6.7
2,4-DCA	0.10	6	58-77	73	6.6	9.3
2,4 DCM	1.0	6	59-76	67	6.5	9.7
	0.012 (LOQ <sup>1</sup> )	6	49-83	72	13	18
2-EHE (2,4-DP-p 2-EHE)	0.12 (LOQ )	6	37-83	68 68	19	28
	1.2	6	41-108	80	22	28
				-12"; GR9445)	22	20
	0.0093 (LOQ <sup>1</sup> )	6	68-118	90	23	26
2,4-DP-p	0.093	6	51-96	<b>69</b>	15	20
2,4-рг-р	0.93	6	47-83	72	13	18

# Table 3. Independent Validation Method Recoveries for 2,4-DP-p and Its Transformation products in Soil\*

Analyte	Fortification Level (mg/kg)	Number of Tests		Mean Recovery (%)	Standard Deviation (%)	Relative Standard Deviation (%)
	0.010 (LOQ)	6	70-130	113	23	20
2,4-DCP	0.10	6	110-130	123	8.2	6.7
	1.0	6	78-110	92	13	14
	0.010 (LOQ)	6	75-110	87	14	16
2,4-DCA	0.10	6	63-83	73	8.0	11
	1.0	6	68-97	80	12	15
2-EHE	0.012 (LOQ <sup>1</sup> )	6	62-73	70	6.2	8.9
2-EHE (2,4-DP-p 2-EHE)	0.12	6	64-83	73	6.4	8.8
(2, <b>4</b> -DI -p 2-LIIL)	1.2	6	72-88	78	6.4	8.2
	San	d soil (Wa	nshington; 0	-12"; GR9451)		
	0.0093 (LOQ <sup>1</sup> )	6	74-129	106	20	19
2,4-DP-p	0.093	6	66-77	71	4.1	5.8
	0.93	6	66-84	71	6.6	9.3
	0.010 (LOQ)	6	71-100	83	13	16
2,4-DCP	0.10	6	68-95	82	12	15
	1.0	6	50-71	61	8.6	14
	0.010 (LOQ)	6	70-85	78	6.1	7.8
2,4-DCA	0.10	6	67-82	74	5.9	8.0
	1.0	6	62-82	71	6.8	9.6
	0.012 (LOQ <sup>1</sup> )	6	74-108	94	16	17
2-EHE (2,4-DP-p 2-EHE)	0.12	6	66-100	82	13	16
(2,4-DP-p 2-EHE)	1.2	6	61-117	77	20	26

\* Data were obtained from p. 14 and Tables 3A-7B, pp. 22-31 of the study report. The values which were outside of guideline requirements are **bolded**. 2,4-DP-p was reported in the tables of the study report, as opposed to methylated 2,4-DP-p, since a conversion factor was used.

1 Fortification levels for 2,4-DP-p and 2-EHE differed from the nominal fortification levels of 0.010 (LOQ), 0.1 and 1.0.

2 Outliers of 20% and 24% recovery not included in data set calculations.

3 Outlier of 22% recovery not included in data set calculations.

4 Outlier of 28% recovery not included in data set calculations.

#### **III. Method Characteristics**

The LOQ was the same in the ECM and ILV, 0.01 ppm (0.01 mg/kg). No comparison to baseline noise, other justification or calculation was provided. The ILV study authors noted that the LOQ was an approximate value since five different soil types were investigated (p. 20). The ILV study authors proposed LOQ S/N ranging *ca*. 2-10 for 2,4-DCP and *ca*. 5-20 for the other analytes, depending on instrument sensitivity and column performance. The LOD of 0.005 ppm (0.005 mg/kg) was based on the calibration standards injected at half of the LOQ (Appendix 5, p. 138). No other justification or calculation was provided.

#### **Table 4. Method Characteristics**

			2,4-DP-p					
	California	Georgia	Indiana	New York	Washington			
Limit of Quantitation (LOQ)	0.01 mg/kg	0.01 mg/kg	0.01 mg/kg	0.01 mg/kg	0.01 mg/kg			
Limit of Detection (LOD)	0.005 mg/kg	0.005 mg/kg	0.005 mg/kg	0.005 mg/kg	0.005 mg/kg			
Linearity (calibration curve r <sup>2</sup> and concentration range)		Quadratic equation	tions reported by	y ECM and ILV				
Repeatable	No	No	No	Yes <sup>4</sup>	No			
Reproducible <sup>1</sup>	Yes <sup>2</sup>	No <sup>2</sup>	No <sup>2,3</sup>	No <sup>2</sup>	Yes <sup>2</sup>			
Specific	Yes	Yes	Yes	Yes	Yes			
			2,4-DCP					
	California	Georgia	Indiana	New York	Washington			
Limit of Quantitation (LOQ)	0.01 mg/kg	0.01 mg/kg	0.01 mg/kg	0.01 mg/kg	0.01 mg/kg			
Limit of Detection (LOD)	0.005 mg/kg	0.005 mg/kg	0.005 mg/kg	0.005 mg/kg	0.005 mg/kg			
Linearity (calibration curve r <sup>2</sup> and concentration range)		Quadratic equation	tions reported by	y ECM and ILV				
Repeatable	No	Yes <sup>4</sup>	Yes <sup>4</sup>	No	No			
Reproducible <sup>1</sup>	No <sup>3</sup>	No	Yes <sup>5</sup>	No <sup>3</sup>	Yes <sup>5</sup>			
Specific	Yes	Yes	Yes	Yes	Yes			
	2,4-DCA							
	California	Georgia	Indiana	New York	Washington			
Limit of Quantitation (LOQ)	0.01 mg/kg	0.01 mg/kg	0.01 mg/kg	0.01 mg/kg	0.01 mg/kg			
Limit of Detection (LOD)	0.005 mg/kg	0.005 mg/kg	0.005 mg/kg	0.005 mg/kg	0.005 mg/kg			
Linearity (calibration curve r <sup>2</sup> and concentration range)		Quadratic equation	tions reported by	y ECM and ILV				
Repeatable	Yes <sup>4</sup>	Yes <sup>4</sup>	No	Yes <sup>4</sup>	Yes <sup>4</sup>			
Reproducible <sup>1</sup>	Yes	No <sup>3,6</sup>	Yes <sup>5</sup>	Yes	Yes			
Specific	Yes	Yes	Yes	Yes	Yes			
			2-EHE					
	California	Georgia	Indiana	New York	Washington			
Limit of Quantitation (LOQ)	0.01 mg/kg	0.01 mg/kg	0.01 mg/kg	0.01 mg/kg	0.01 mg/kg			
Limit of Detection (LOD)	0.005 mg/kg	0.005 mg/kg	0.005 mg/kg	0.005 mg/kg	0.005 mg/kg			
Linearity (calibration curve r <sup>2</sup> and concentration range)	Quadratic equations reported by ECM and ILV							
Repeatable	Yes <sup>4</sup>	Yes <sup>4</sup>	Yes <sup>4</sup>	Yes <sup>4</sup>	Yes <sup>4, 7</sup>			
Reproducible <sup>1</sup>	Yes <sup>5,8</sup>	No <sup>8</sup>	No <sup>3,8</sup>	Yes <sup>8</sup>	Yes <sup>5,8</sup>			
Specific	Yes	Yes	Yes	Yes	Yes			

Data were obtained from pp. 8-9, 20; Appendix 2, pp. 85-105; Appendix 5, pp. 134-135, 138; Appendix 5, Table IV, pp. 147-149; Appendix 5, Figure 3, p. 151 of the study report.

1 Although it was not reported as such in the study report, the reviewer compiled this table with the assumption that the five soils of the ECM were the same as the five soils of the ILV (see Comment#5).

2 2,4-DP-p Test samples were fortified at 0.0093, 0.093 and 0.93 mg/kg, instead of 0.01, 0.1 and 1.0 mg/kg.

3 Recoveries at LOQ were acceptable, but recoveries at 10×LOQ did not meet guideline requirements.

4 Based on only three replicates fortified.

5 Recoveries at LOQ and 10×LOQ were acceptable, but recoveries at 100×LOQ did not meet guideline requirements.

6 n = 4 at LOQ fortification level.

7 n = 1 at LOQ fortification level (n = 3 at  $10 \times LOQ$ ).

8 2-EHE Test samples were fortified at 0.012, 0.12 and 1.2 mg/kg, instead of 0.01, 0.1 and 1.0 mg/kg.

#### **IV. Method Deficiencies and Reviewer's Comments**

- 1. Method recoveries did not meet guideline criteria for precision and accuracy (mean 70-120%; RSD  $\leq$ 20%) for multiple soil/analyte experiments in both the ECM and ILV (see DER Table 4). In the ILV, mean recoveries or RSDs at the LOQ or 10×LOQ were not within guideline requirements for analysis of 2,4-DP-p in the Georgia, Indiana and New York test soils, 2,4-DCP in the California, Georgia and New York test soils, 2,4-DCA in the Georgia test soil and 2-EHE in the Georgia and Indiana test soils. The ILV study authors proposed that microbial degradation prior to the extraction step was the cause of the low recoveries in the Indiana and Georgia test soils (p. 20). In the ECM, only three samples were spiked at the LOQ for each soil/analyte (see Comment #2), however, the mean recovery (±SD) of 2,4-DP-p at the LOQ, across all five soils, was 65.7 ± 4.9%, which is outside of the guideline requirement (Appendix 5, Table IIIa, p. 144).
- 2. A minimum of five spiked samples were not analyzed at each concentration for each soil matrix in the ECM. Only three samples were considered for each test soil at a fortification level of LOQ and 10x LOQ (Appendix 5, Tables IIIa-IIIc, pp. 144-146). The OCSPP Guideline 850.6100 criteria requires a minimum of five samples. Therefore, the reviewer could not adequately determine whether the method is quantitative for each analyte for each soil matrix. Across all test soils, mean recoveries of 2,4-DCP, 2,4-DCA and 2-EHE at the LOQ and of all analytes at 10x and 200x LOQ were within the guideline requirement (Appendix 5, Table IIIa-IIIb, pp. 143-145).
- 3. The estimations of the LOQ and LOD in the ECM were not based on scientifically acceptable procedures as defined in 40 CFR Part 136. No calculations were reported; no comparison was made to chromatogram background levels. Detection limits should not be based on the arbitrarily selected lowest concentration in the spiked samples. Additionally, the lowest toxicological level of concern in soil was not reported. An LOQ above toxicological levels of concern results in an unacceptable method classification. 2,4-DP-p, 2,4-DCA and 2-EHE are not listed in the EPA Regional Screening Levels (http://www.epa.gov/region9/superfund/prg/) for residential soil; however, 2,4-DCP is listed with a screening level of 18 mg/kg in residential soil which is much higher than the reported LOQ for this method.
- 4. In the ILV, measured fortification levels differed from the nominal levels of 0.01 mg/kg, 0.10 mg/kg and 1.0 mg/kg. The reported fortification levels were 0.0093 mg/kg, 0.093 mg/kg and 0.93 mg/kg for 2,4-DP-p and 0.012 mg/kg, 0.12 mg/kg and 1.2 mg/kg for 2-EHE. The reviewer evaluated the ILV results based on the nominal levels, but it is noted that the 2-EHE testing was not performed at ≤LOQ.
- 5. The five test soils used in the ECM were not characterized and were only identified by the state in which they were collected (Appendix 5, pp. 136-137). Test soils used in the ILV were characterized by texture and depth of collection; however, percentages of sand, silt, clay, moisture and organic matter were not reported (p. 14). Although the states in which the five soils were collected were the same, there was no statement in either the

ECM or ILV which clarified whether the soils used in the ILV were the same as those used in the ECM.

- 6. In the ECM, the GC/MS chromatograms are provided for reagent blanks, the matrix blank for the New York soil and spiked samples at the LOQ, 10× LOQ and 200× LOQ for all analytes in the New York soil (Appendix 5, Figures 3-6d, pp. 151-163). Chromatograms were not included for matrix blanks of the California, Indiana, Georgia and Washington soils and spiked samples at the LOQ, 10× LOQ and 200× LOQ for all analytes in the California, Indiana, Georgia and Washington soils and spiked samples at the LOQ, 10× LOQ and 200× LOQ for all analytes in the California, Indiana, Georgia and Washington soils. It is preferred that chromatograms are provided for all test soils.
- 7. In the ILV, GC/MS Chromatograms were provided for only three of the five test soils: New York, Indiana and California (Appendix 1, pp. 32-84). The study authors noted that the Indiana soil was the "worst" soil type, the California soil was the "cleanest" soil type and the New York soil was the "average" soil type (p. 8). It is preferred that chromatograms are provided for all test soils.
- 8. The communication between the ILV and ECM was documented (Appendix 3, pp. 106-108). Only two minor modifications of the ECM were performed by the ILV authors: silanized glass wool was not included in the deactivated glass injection port liner and a deactivated fused silica pre-column was installed in front of the analytical capillary column (pp. 10-11). The documentation of approval of these modifications by the ECM authors, ADPEN Labs, was provided (Appendix 3, pp. 106-108). Neither of these modifications required an internal validation to be conducted by the ECM authors or the sponsor. The ILV authors also reported a few critical steps regarding column maintenance (pp. 19-20).
- 9. Calibration curves were calculated by the ECM and ILV study authors using quadratic equations (Appendix 2, pp. 85-105; Appendix 5, Table IV, pp. 147-149; Appendix 5, Figure 3, p. 151). The reviewer did not perform linear regression analysis for the raw data sets.
- It was stated in the ILV that initially there were several trials that were unsuccessful (p. 20). One trial was done on the Washington and Georgia soil types that gave variable recoveries and were unacceptable and two other trials were done on the Indiana soil that were also unacceptable.
- 11. It was reported for the ILV that a single analyst completed a sample set consisting of 12 samples in 14-19 hours (p. 17). The ECM reported that a single analyst completed a sample set consisting of 12 soil samples in *ca*. 12.0 hours (Appendix 5, p. 134).

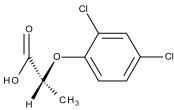
#### V. References

- U.S. Environmental Protection Agency. 2012. Ecological Effects Test Guidelines, OCSPP 850.6100, Environmental Chemistry Methods and Associated Independent Laboratory Validation. Office of Chemical Safety and Pollution Prevention, Washington, DC. EPA 712-C-001.
- 40 CFR Part 136. Appendix B. Definition and Procedure for the Determination of the Method Detection Limit-Revision 1.11, pp. 317-319.

# **Attachment 1: Chemical Names and Structures**

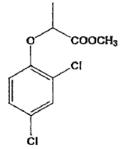
# 2,4-DP-p; Dichlorprop-p; 2,4-Dichlorprop-p

IUPAC Name:	(2R)-2-(2,4-dichlorophenoxy)propanoic acid (+)-(R)-2-(2,4-dichlorophenoxy)propionic acid ("chemical name" in MBD = 11,12)
	MRID p. 11-12)
CAS Name:	Not reported
CAS Number:	120-36-5
SMILES String:	O=C(O)C(Oc(c(cc(c1)Cl)Cl)c1)C



## Methylated 2,4-DP-p; 2,4-DP-p methyl ester

IUPAC Name:	(+)-(R)-2-(2,4-dichlorophenoxy)propionic acid methyl ester ("chemical name" in MRID p. 11-12)
CAS Name:	Not reported
CAS Number:	Not reported
<b>SMILES String:</b>	Not reported
-	



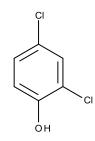
# 2,4-DCA; 2,4-Dichloroanisole

IUPAC Name:	2,4-Dichloro-1-methoxy-benzene 2,4-Dichloroanisole ("chemical name" in MRID p. 11-12)
CAS Name:	Not reported
CAS Number:	553-82-2
SMILES String:	O(c(c(cc(c1)Cl)Cl)c1)C

CI 0 ∙сн₃

# 2,4-DCP; 2,4-Dichlorophenol

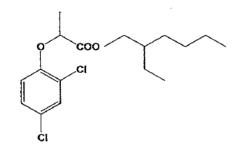
<b>IUPAC Name:</b>	2,4-Dichlorophenol
CAS Name:	Not reported
CAS Number:	120-83-2
SMILES String:	Oc(c(cc(c1)Cl)Cl)c1



# 2-EHE; 2,4-DP-p 2-EHE; 2,4-DP-p 2-Ethylhexyl ester

<b>IUPAC Name:</b>	(+)-(R)-2-(2,4-dichlorophenoxy)propionic acid 2-ethylhexyl ester
	("chemical name" in MRID p. 11-12)
CAS Name:	Not reported

CAS Maine.	Not reported
CAS Number:	Not reported
SMILES String:	Not reported



Chemical: 2,4-DP-p PC: 031402 MRID: 44243401 Guideline: 850.6100 ECM Validation for Determination of 2,4-DP-p in Soil

	ified				P-p (Methyl	ated)		
	ng	Recovery	Mean	SD <sup>1</sup>	RSD <sup>2</sup>	,		
a.i./		(%)	(%)	(%)	(%)	Max	Min	n =
	0/			Califo	ornia			
	0.01	64.2						
LOQ		66.6						
		60.9	63.9	2.9	4.5	66.6	60.9	3
	0.10	90.2						
		89.2						
		69.9	83.1	11.4	13.8	90.2	69.9	3
	2.0	82.6						
		92.6	87.6	7.1	8.1	92.6	82.6	2
				Geo	rgia			
	0.01	61.4						
LOQ		59.9						
			60.7	1.1	1.7	61.4	59.9	2
	0.10	66.1						
		62.8						
		70.3	66.4	3.8	5.7	70.3	62.8	3
	2.0	62.5						_
		81.4	72.0	13.4	18.6	81.4	62.5	2
	0.04	00.4	T	Indi	ana			
	0.01	60.4						
LOQ		64.7	00.4	0.4	0.7	047	00.4	0
	0.40	64.2	63.1	2.4	3.7	64.7	60.4	3
	0.10	67.5						
		62.3 67.5	65.8	3.0	4.6	67.5	62.3	3
	2.0	70.2	05.8	3.0	4.0	07.5	02.3	3
	2.0	63.1	66.7	5.0	7.5	70.2	63.1	2
		05.1	00.7	New		70.2	00.1	2
	0.01	71.3		New	IOIK			
LOQ	0.01	75.0						
		64.7	70.3	5.2	7.4	75.0	64.7	3
	0.10	76.9	, 0.0	0.2	,	, 0.0	0	0
	00	74.1						
		72.7	74.6	2.1	2.9	76.9	72.7	3
	2.0	84.4	-				-	
	-	82.6	83.5	1.3	1.5	84.4	82.6	2
				Washi	ngton			
	0.01	63.2			-			
LOQ		69.9						
		73.6	68.9	5.3	7.7	73.6	63.2	3
	0.10	65.1						
		63.2						
		64.2	64.2	1.0	1.5	65.1	63.2	3
	2.0	77.3						
		73.8	75.6	2.5	3.3	77.3	73.8	2

Results from Appendix 5, Tables IIIa-IIIc, pp. 144-146 of MRID 44243401.

Means and standard deviations calculated using Microsoft program functions =AVERAGE(A1:A2) and =STDEV(A1:A2 Any discrepancies between reviewer calculated values and reported results most likely due to rounding.

1 SD = Standard Deviation; determined using the "unbiased" or "n-1" method.

2 RSD = Relative Standard Deviation; calculated as (SD/mean) x 100.