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AQUATIC LIFE AMBIENT WATER QUALITY CRITERIA FOR CARBARYL - 2012

AQUATIC LIFE
AMBIENT WATER QUALITY CRITERIA FOR
CARBARYL

(CAS Registry Number 63-25-2)

April 2012

U.S. ENVIRONMENTAL PROTECTION AGENCY
OFFICE OF WATER
OFFICE OF SCIENCE AND TECHNOLOGY
HEALTH AND ECOLOGICAL CRITERIA DIVISION
WASHINGTON, D.C.

NOTICES

This document has been reviewed by the Health and Ecological Criteria Division, Office of Science and Technology, U.S. Environmental Protection Agency, and is approved for publication.

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FOREWORD

Section 304(a)(1) of the Clean Water Act of 1977 (P.L. 95-217) requires that the Administrator of the Environmental Protection Agency (EPA) publish water quality criteria that accurately reflect the latest scientific knowledge on the kind and extent of all identifiable effects on health and welfare that might be expected from the presence of pollutants in any body of water, including ground water. This document is a final ambient water quality criteria (AWQC) document for the protection of aquatic life based upon consideration of all available information relating to effects of carbaryl on aquatic organisms.

The term "water quality criteria" is used in two sections of the Clean Water Act, section 304(a)(1) and section 303(c)(2). The term has a different program impact in each section. In section 304, the term represents a non-regulatory, scientific assessment of ecological effects. Criteria presented in this document are such scientific assessments. If water quality criteria associated with specific stream uses are adopted by a state or EPA as water quality standards under section 303, they become maximum acceptable pollutant concentrations for permitting and listing in ambient waters within that state. Water quality criteria adopted in state water quality standards could have the same numerical values as criteria developed under section 304. However, in many situations states might want to adjust water quality criteria developed under section 304 to reflect local environmental conditions and human exposure patterns. Alternatively, states may use different data and assumptions than EPA in deriving numeric criteria that are scientifically defensible and protective of designated uses. It is not until their adoption as part of state water quality standards that criteria become regulatory. Guidelines to assist the states and Indian tribes in modifying the criteria presented in this document are contained in the Water Quality Standards Handbook (U.S. EPA 1994). This handbook and additional guidance on the development of water quality standards and other water-related programs of this agency have been developed by the Office of Water.

This final document is guidance only. It does not establish or affect legal rights or obligations. It does not establish a binding norm and cannot be finally determinative of the issues addressed. Agency decisions in any particular situation will be made by applying the Clean Water Act and EPA regulations on the basis of specific facts presented and scientific information then available.

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This aquatic life criteria document for carbaryl has undergone many revisions and has benefitted from the insights of various reviewers, including members of the Ecological Risk Assessment Branch in the Office of Water, Office of Science and Technology. Additionally, much of the fate and transport information and additional toxicity information have been procured from various ecological risk assessments documents in the Office of Pesticide Programs' Environmental Fate and Effects Division.

EXECUTIVE SUMMARY

Background

Carbaryl (1-naphthyl-N-methylcarbamate; $C_{12}H_{22}NO_2$, CAS #63-25-2) is an insecticide, a molluscicide, and is used to thin fruit in orchards, and it belongs to the N-methyl carbamate class of pesticides. Introduced by the Union Carbide Corporation in 1956, carbaryl acts on animals through contact and ingestion. Carbaryl is registered in the United States for use in controlling more than 160 insect pests on over 115 agricultural and non-crop use applications, including home and garden uses (U.S. EPA 2007a; U.S. EPA 2010a). Major uses include insect control on lawns, home gardens, citrus, fruit, forage and field crops, forests, nuts, ornamentals, rangeland, turf, shade trees, poultry and pets (U.S. EPA 2010a). Agricultural crops with the greatest annual use of carbaryl include apples, pecans, grapes, alfalfa, oranges, and corn. Carbaryl was the third most commonly used conventional pesticide used in homes and gardens in 2005 and 2007 with a range of 4 to 6 million pounds of active ingredient used annually (U.S. EPA 2011).

U.S. Geological Survey's National Water Quality Assessment (NAWQA) Program reported carbaryl as the second most frequently found insecticide in water, with detections in approximately 50% of urban streams. Frequencies of detection in surface water associated with urban uses are commonly higher than those associated with agricultural uses. The majority of these detections are below 0.1 $\mu\text{g/L}$. The NAWQA program analyzed a total of 11,732 water samples from 1999 to 2005 for carbaryl. They reported 29% of all samples with concentrations greater than the minimum detection limit of 0.021 $\mu\text{g/L}$. The mean concentration reported was 0.058 $\mu\text{g/L}$ and the maximum concentration was 33.5 $\mu\text{g/L}$, associated with agricultural land. The highest concentration reported in urban areas was 16 $\mu\text{g/L}$.

Carbaryl Criteria Derivation

U.S. EPA is developing recommended ambient water quality criteria for carbaryl through its authority under section 304(a) of the Clean Water Act (CWA). These water quality criteria may be used by states and authorized tribes to establish water quality standards for carbaryl. The carbaryl aquatic life criteria are developed using peer reviewed methods and data that are acceptable for the derivation of a freshwater and estuarine/marine criteria. Data evaluated for criteria derivation include data submitted in support of the registration of this pesticide and

reviewed by U.S. EPA’s Office of Pesticide Programs as well as studies reported in the open literature.

The resulting recommended ambient water quality criteria indicate that freshwater aquatic organisms would have an appropriate level of protection if the one-hour average concentration does not exceed 2.1 µg/L more than once every three years on average and if the four-day average concentration of carbaryl does not exceed 2.1 µg/L more than once every three years on average (except where a locally important species may be more sensitive).

Estuarine/marine aquatic organisms would have an appropriate level of protection if the one-hour average concentration does not exceed 1.6 µg/L more than once every three years on average (except where a locally important species may be more sensitive).

This document provides guidance to states and tribes authorized to establish water quality standards under the CWA to protect aquatic life from the acute and chronic effects of carbaryl. Under the CWA, states and authorized tribes may adopt water quality criteria into water quality standards to protect designated uses. While this document constitutes U.S. EPA’s scientific recommendations regarding ambient concentrations of carbaryl, this document does not substitute for the CWA or U.S. EPA’s regulations; nor is it a regulation itself. Thus, it cannot impose legally binding requirements on U.S. EPA, states, authorized tribes, or the regulated community, and it might not apply to a particular situation based upon the circumstances. State and tribal decision-makers retain the discretion to adopt approaches on a case-by-case basis that differ from this guidance when appropriate. U.S. EPA may change this guidance in the future.

Table 1. Summary of Aquatic Life Criteria for Carbaryl

	Acute	Chronic
Freshwater	2.1 µg/L	2.1 µg/L
Estuarine/marine	1.6 µg/L	N/A

N/A – not available, unable to calculate estuarine/marine chronic criterion

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I. INTRODUCTION AND BACKGROUND

U.S. EPA is developing ambient water quality criteria for carbaryl through its authority under section 304(a) of the Clean Water Act (CWA). Although not regulatory in themselves, these water quality criteria may be used by states and authorized tribes to establish water quality standards for carbaryl to protect aquatic life designated uses. The criteria presented herein are the agency's best estimate of maximum concentrations of carbaryl to protect most aquatic organisms from unacceptable short- or long-term effects.

Carbaryl (1-naphthyl-N-methylcarbamate; C₁₂H₂₂NO₂, CAS #63-25-2) is an insecticide belonging to the N-methyl carbamate class of pesticides. Carbaryl is registered in the United States for use on over 115 agricultural and non-crop use applications, including home and garden uses (U.S. EPA 2007a; U.S. EPA 2010a). All pesticides distributed or sold in the United States must be registered by U.S. EPA, and the registrations are periodically re-evaluated to determine whether they meet current regulatory standards. The next Registration Review Decision for carbaryl will be made by U.S. EPA Office of Pesticide Programs (OPP) in 2015.

In 2003, OPP completed an ecological risk assessment to support a 2004 Interim Registration Eligibility Decision (IRED) for carbaryl, which identified potential acute risk to fish associated with one use, and acute and chronic risks to aquatic invertebrates associated with most registered uses of carbaryl (U.S. EPA 2004a). The IRED cited a study by the U.S. Geological Survey's National Water Quality Assessment (NAWQA) Program that reported carbaryl as the second most frequently found insecticide in water, with detections in approximately 50% of urban streams (U.S.G.S. 2006). Of the top five most frequently detected insecticides (listed in descending order) – chlorpyrifos, carbaryl, carbofuran, diazinon, and malathion (U.S.G.S. 2006), there are existing ambient water quality criteria for chlorpyrifos, diazinon and malathion; carbofuran tolerances are now cancelled. In 2007 (U.S. EPA 2007b), OPP updated a summary of U.S. surface water monitoring supporting the final Registration Decision. That assessment concluded that carbaryl had relatively high frequencies of detection, with concentrations as high as 33.5 µg/L in an agricultural area and 16.5 µg/L in urban areas. Detections were more frequent in urban areas than agricultural areas, but concentrations were mostly less than 0.1 µg/L. In more recent assessments by OPP, the Agency has made 'may affect' and 'likely to adversely affect' determinations for the California red-legged frog (*Rana aurora draytonii*; U.S. EPA 2007a) and the California Delta smelt (*Hypomesus transpacificus*; U.S. EPA 2010b) from the use

of carbaryl in California. Additionally, the Agency determined that there is the potential for modification of designated critical habitat for both of the Federally-listed species from the use of carbaryl. Based on the conclusions of these assessments, the Agency indicated that formal consultations with the U.S. Fish and Wildlife Service under section 7 of the Endangered Species Act should be initiated for the pesticide registration action.

II. PROBLEM FORMULATION

Problem formulation provides a strategic framework for water quality criteria development by focusing the effects assessment on the most relevant chemical properties and endpoints. The structure of this effects assessment is consistent with U.S. EPA's *Guidance for Ecological Risk Assessment* (U.S. EPA, 1998) and the approach used by U.S. EPA for pesticide effects assessment (U.S. EPA, 2004a).

This ecological effects assessment develops scientifically defensible water quality criteria values for carbaryl under section 304(a)(1) of the Clean Water Act. The goal of the Clean Water Act is to protect and restore the biological, chemical and physical integrity of waters of the U.S. Section 304(a)(1) of the Clean Water Act requires U.S. EPA to develop criteria for water quality that accurately reflect the latest scientific knowledge. These criteria are based solely on data and best professional scientific judgments on toxicological effects. Criteria are developed following the guidance outlined in the Agency's "*Guidelines for Deriving Numerical National Water Quality Criteria for the Protection of Aquatic Organisms and Their Uses*" (Stephan et al. 1985).

Once the section 304(a) water quality criteria are finalized, states and authorized tribes may adopt the criteria into their water quality standards to protect designated uses of water bodies. States and tribes may also modify the criteria to reflect site-specific conditions or use other scientifically defensible methods to develop standards. Water quality standards are subsequently approved by U.S. EPA.

A. Stressor Characteristics

1. Mode of Action and Toxicity

Carbaryl is an insecticide belonging to the N-methyl carbamate class of pesticides. After contact or ingestion, the toxic mode of action of carbaryl (and other carbamate insecticides) for

animals is inhibition of the enzyme acetylcholinesterase (AChE) at synaptic junctions in the nervous system. AChE breaks down the neurotransmitter acetylcholine. Inhibition of AChE results in the accumulation of acetylcholine in the nerve synapses which leads to continual firing of nerve pulses throughout the nervous system. This buildup results in uncontrolled movement, paralysis, convulsions, tetany, and possible death (Gunasekara et al. 2008). Without proper nerve function, the respiratory, circulatory and other vital body systems will fail, ultimately causing death of the organism (Kuhr and Dorough 1976). The acetylcholinesterase inhibition effects of carbaryl are reversible with removal of exposure of the stressor chemical. The primary degradate of carbaryl, 1-naphthol, does not inhibit acetylcholinesterase and its toxic mode of action for animals is thought to be narcosis (Russom et al. 1997). In aquatic organisms, narcosis is a reversible anesthetic effect that is caused by chemicals partitioning into cell membranes and nervous tissue that result in disruption of cell functions including central nervous system function (Barron et al. 2001).

Carbaryl is also used to thin fruit (removing excess fruit so the remaining fruit increase in size and flavor) in orchards; its activity in the abscission (dropping) of flower buds may be related to its structural similarity to plant auxins, such as α -naphthalene acetic acid. The carbaryl degradate 1-naphthol is a known plant auxin (Wood 2010).

Carbaryl is included on the final list of chemicals for initial Tier 1 Screening under U.S. EPA's Endocrine Disruptor Screening Program (EDSP) released on April 2009. This list includes chemicals that the Agency has decided should be tested first, based upon exposure pathway potential to humans; this list should not be construed as a list of known or likely endocrine disruptors. On October 2009, U.S. EPA issued test orders for a data call-in (DCI) from manufacturers/formulators of carbaryl. Test results for carbaryl are due by November 12, 2011.

Water temperature, pH and hardness are factors potentially affecting the toxicity of carbaryl in the aquatic environment. Schoettger and Mauck (1978) found that the toxicity of carbaryl increased with temperature and hardness (that were in the normal range for natural waters) for the brook trout (*Salvelinus fontinalis*). Rainbow trout (*Oncorhynchus mykiss*) and bluegill (*Lepomis macrochirus*) were similarly tested (Sanders et al. 1983). Bluegill showed increased susceptibility to carbaryl with increases in pH, hardness, and temperature, whereas rainbow trout showed no appreciable changes to any of the parameters tested. Midge larva,

Chironomus riparius, showed no significant changes in toxicity when exposed to carbaryl at pHs of 4, 6 and 8; additional testing at elevated temperatures showed that increased temperature resulted in slight increases in carbaryl toxicity at pHs of 4 and 6, but not at 8 (Fisher and Lohner 1986). Thus, the lack of sufficient data showing consistent trends (with the exception of brook trout) for specific physio-chemical parameters precludes the need to adjust water quality criteria for these parameters.

An assessment of the available data regarding the relative toxicity of the degradate 1-naphthol to parent carbaryl compound is inconclusive. Thus, additional acute and chronic aquatic toxicity studies involving fish and invertebrates are needed to complete a more definitive evaluation and to address the uncertainties surrounding the toxicity and effects from 1-naphthol.

2. Overview of Pesticide Usage

Carbaryl (main trade names: Sevin, Arilat, Carbatox, Dicarbam) was first registered by the Union Carbide Corporation in 1956. Currently, carbaryl is nationally registered for over 115 uses in agriculture, professional turf management, ornamental production, and residential settings (U.S. EPA 2010a). Carbaryl's main use is an insecticide in the control of mites, fleas, aphids, fire ants, common garden insects, and other insect pests in orchards and agricultural fields. Carbaryl also is registered for use as a mosquito adulticide and as a molluscicide. Agricultural uses include root crops, tree fruit, nuts, fruit and vegetable, and grain crops. Carbaryl is used by homeowners in residential settings for lawn care, gardening (vegetables and ornamentals), and pet care (pet collars, powders, and dips, in kennels, and on pet sleeping quarters). Carbaryl also is used by nursery, landscape, and golf course industries on turf, annuals, perennials, and shrubs. Carbaryl has forestry and rangeland uses and is used for control of grasshoppers and crickets, adult mosquitoes, ticks and fire ants. Additionally, carbaryl is used to thin fruit in orchards to enhance fruit size and enhance repeat bloom. Carbaryl is available for use in various formulations, including baits, dusts, wettable powders, molasses, oil and water suspensions, pellets and granules.

At the time of the carbaryl IRED (U.S. EPA 2004b), approximately 3.9 million pounds of carbaryl active ingredient were sold annually in the U.S.; with about half used in agriculture and half in non-agricultural settings (per 1998 data). The amount of carbaryl usage per year in agriculture has declined somewhat from an average of 1.9 million pounds of active ingredient in

1992 to 1 to 1.5 million pounds of active ingredient in 2001. **Figure 1** depicts the extent of estimated annual carbaryl agricultural use nationally as of 2002. According to U.S. EPA's October 19, 2009 Screening Level Estimates of Agricultural Uses of Carbaryl (U.S. EPA, 2010a), the amount of carbaryl (total formulated product) use has declined to approximately 1.5 million pounds used annually in 2001. The highest annual application is hay (approximately 600,000 lbs), followed by apples (300,000 lbs), pecans (100,000 lbs), and oranges (100,000 lbs) (U.S. EPA 2010a). Usage of carbaryl in the home and garden sector was ranked third most commonly used pesticide active ingredient with a range of 4 to 6 million pounds active ingredient applied in 2005 and 2007. This was a decrease from the 6 to 9 million pounds active ingredient of carbaryl applied in home and gardens during 2003 (U.S. EPA 2011). Usage data in the home and garden sector is not available to allow development of a map for non-agricultural use similar to **Figure 1**.

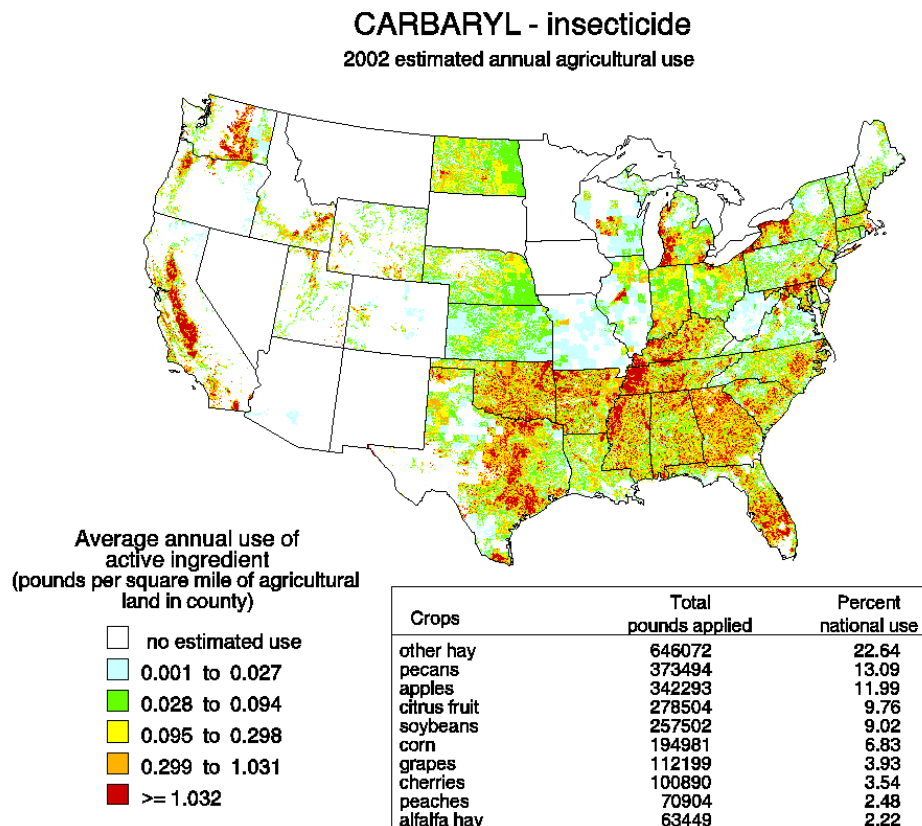


Figure 1: Historical Extent (2002) of carbaryl usage in agriculture.
(Source http://ca.water.usgs.gov/pnsp/pesticide_use_maps/show_map.php?year=02&map=m6006)

3. Environmental Fate and Transport

Technical grade carbaryl is an odorless and colorless to tan crystal (depending on purity), and has a molecular weight of 201.22 g/mole. Carbaryl has a density of 1.21 g/mL at 20°C, a vapor pressure of 1.36×10^{-7} torr at 25°C, and is soluble in water to 32 mg/L at 30°C (U.S. EPA 2010a). It has an n-octanol/water partitioning coefficient (K_{ow}) of 229 and an organic-carbon normalized partition coefficient (K_{oc}) of 196 L/kg_{oc} (U.S. EPA 2010a). Its Henry's law constant of 1.28×10^{-8} atm m³ g/mol @ 25°C suggests it will have minimal volatilization from aqueous solutions (U.S. EPA 2010a).

The hydrolysis of carbaryl is pH related, occurring more rapidly in alkaline solutions (U.S. EPA 2010a). There is no evidence of degradation at a pH of 5 and degradation occurs in neutral (pH 7) and alkaline (pH 9) systems with measured half-lives ($t_{1/2}$) of 12 days and 0.13 days, respectively (U.S. EPA 2010a). Hydrolysis of carbaryl is also evident in saltwater, with reported half-life of one day (pH=7.9) in filtered saltwater (Armbust and Crosby 1991). The major degradate of hydrolysis is 1-naphthol. In aerobic soil and aquatic systems, microbially-mediated degradation (metabolism) of carbaryl occurs fairly rapidly ($t_{1/2}$ = 4.0 and 4.9 days respectively) but more slowly under anaerobic conditions ($t_{1/2}$ = 72 days) (U.S. EPA 2010a). Under both aerobic and anaerobic conditions the major degradate is 1-naphthol.

Aqueous photolysis is another route of degradation for carbaryl, and may occur in the upper water column of an aquatic system, where clearer waters allow light to penetrate. Carbaryl degraded rapidly by aqueous photolysis to 1-naphthol with a half-life of 1.8 days, which in turn degraded very rapidly with a half-life of less than 1 hour.

The primary degradate (*i.e.*, 1-naphthol) in all the degradation studies reviewed, only formed transiently ($t_{1/2}$ ~ <1 hr) and in most cases completely degraded by the end of each study. Data on the primary degradate, 1-naphthol are limited; however, it appears to be less mobile than the parent (discussed below) and is not likely to persist due to fairly rapid degradation. Since 1-naphthol can occur from a variety of natural and anthropogenic processes, its presence cannot be considered indicative of carbaryl use.

Besides 1-naphthol, the only other major degradate reported was 1, 4-naphthoquinone which was found at 17.3% on the third day after study initiation in the aerobic aquatic metabolism study (U.S. EPA 2010a). No additional environmental fate data were available for this degradate.

Carbaryl is considered moderately mobile in soils and sorption of carbaryl is dependent on soil organic matter (U.S. EPA 2010a). Following a high rain event, carbaryl may reach aquatic environments from areas of application in sheet and channel flow runoff. In urban environments, carbaryl present in runoff from yards and gardens could eventually end up in stormwater and wastewater treatment influent. Agricultural applications could have runoff that is subsequently discharged into a surface water body.

Potential transport mechanisms of carbaryl in air include spray drift, and secondary drift of volatilized or soil-bound residues leading to deposition onto nearby or more distant ecosystems. Carbaryl has been shown to be transported and deposited by atmospheric processes (U.S. EPA 2010a); however, given carbaryl's relatively rapid degradation, its potential for long-range atmospheric transport is likely limited.

4. Occurrence

In 2003, the OPP Environmental Fate and Effects Division reported that carbaryl was the second most widely detected insecticide in surface water in the U. S. Geological Survey's (USGS) National Water Quality Assessment (NAWQA) monitoring program. In the 2003 assessment of NAWQA data, 1,067 (21%) out of 5,198 surface water samples had detections greater than the minimum detectable limit (0.021 µg/L). The maximum reported concentration was 5.5 µg/L across all sites. The mean concentration was 0.11 µg/L, with a standard deviation of 0.43 µg/L. In a summary of pesticide occurrence and concentrations for 40 NAWQA stream sites with primarily agricultural basins, carbaryl was detected in 11% of the samples (N = 1,001) with a maximum concentration of 1.5 µg/L.

In a report released in 2006 summarizing pesticide results from NAWQA from 1992 to 2001 (U.S.G.S. 2006), carbaryl is listed as one of the 14 most frequently detected pesticide compounds in surface water and one of the 3 most frequently detected along with diazinon and chlorpyrifos. Carbaryl was detected in 50% of urban samples over this time period. The majority of carbaryl concentrations reported were low with 35% of the urban samples less than 0.1 µg/L. Detection frequencies in agricultural and mixed land use streams were lower (10% and 17%, respectively), and concentrations associated with those land uses were almost all less than 0.1 µg/L.

In 2007, OPP obtained carbaryl data from the USGS NAWQA data warehouse from 1999 to 2005. A total of 11,732 samples were collected in that timeframe and analyzed for carbaryl with 29% of all samples reporting residues greater than the minimum detection limit. For samples with detections, the mean concentration reported was 0.058 µg/L. The maximum concentration reported was 33.5 µg/L at a location associated with agricultural land (mean in agricultural areas: 0.094 µg/L). The detection frequency associated with agricultural uses was lower (19%) than that associated with urban uses (50%). The highest concentration reported in an urban area was 16 µg/L. The higher detection frequency in urban streams (versus agricultural or mixed land uses) is consistent with data summarized in the USGS reports and earlier EPA 2003 assessment (U.S. EPA 2003). The concentrations detected in urban streams (mostly low concentrations, a few detections in the multiple ppb range), are also consistent with earlier data. The relatively high concentration reported associated with agricultural uses is unusual but not outside of the range predicted by modeling done by EPA for pesticide registration (U.S. EPA 2008). Thus, carbaryl was detected less frequently but at higher concentrations in agricultural areas/uses, while carbaryl was detected more frequently but at lower concentrations in urban areas.

Carbaryl's vapor pressure (1.36×10^{-6} torr) and Henry's law constant (1.28×10^{-8} atm-m⁻³ mol⁻¹) are consistent with compounds which are found at least occasionally in the atmosphere. Other than near-field spray drift studies, carbaryl was only measured in a two studies. Carbaryl was detected in 37 percent of rainfall samples collected from Maryland, Indiana, Nebraska and California agricultural watersheds during the 2003 and 2004 growing season, at concentrations ranging from 0.024 to 0.093 µg/L (Vogel et al. 2008). Carbaryl was found in fog in concentrations as high as 0.069 µg/L in six samples collected at three sites along the Pacific Coast in Monterey County, CA (Schomburg et al. 1991). Carbaryl has been also detected in precipitation samples in California (Majewski et al. 2006). Out of 137 rain samples, 93 had detectable carbaryl with a maximum concentration of 0.756 µg/L. Based on these data, it is possible that carbaryl can be deposited on land and aquatic environments in precipitation.

B. Assessment Endpoints and Measures of Effect

U.S. EPA derives ambient water quality criteria for the protection of aquatic life that are protective of the designated uses established for waters of the United States. U.S. EPA is using

the peer-reviewed procedures defined in the Agency's "*Guidelines for Deriving Numerical National Water Quality Criteria for the Protection of Aquatic Organisms and Their Uses*" (Stephan et al. 1985) to derive national Water Quality Criteria for carbaryl.

1. Stressors of Concern

This criteria document quantitatively considers effects of exposures of carbaryl only. Carbaryl degrades into one major degradate: 1-naphthol; however, 1-naphthol is also formed naturally as a degradation product of naphthalene and other polycyclic aromatic hydrocarbons. Available environmental fate data indicate that 1-naphthol degrades more rapidly and is less mobile than carbaryl. Toxicity data indicate that 1-naphthol is roughly equal to or less toxic than the parent compound depending on the species tested (U.S. EPA 2010b). Additional data are needed to better quantify the relative toxicity of the degradate 1-naphthol to parent carbaryl. Since 1-naphthol is less persistent and less mobile than the parent compound, acts through a different mode of action, and is no more toxic than the parent compound, this criteria document focuses on the parent compound alone.

2. Assessment Endpoints

Assessment endpoints are defined as "explicit expressions of the actual environmental value that is to be protected" and are defined by an ecological entity (species, community, or other entity) and its attribute or characteristics (U.S. EPA 1998). Assessment endpoints may be identified at any level of organization (e.g. individual, population, community). In the context of the Clean Water Act, aquatic life criteria rely on the results of growth, reproduction, and survival of the assessed taxa. This information is aggregated into a species sensitivity analyses that evaluates the impact on the aquatic community. Criteria are designed to be protective of the vast majority of aquatic species in an aquatic community (i.e., *5th percentile of tested aquatic animals representing the aquatic community*). As a result, the designated uses (i.e., aquatic life support) and their associated criteria may be considered as assessment endpoints. To develop 304(a) aquatic life criteria under CWA, U.S. EPA typically requires the following:

- Acute toxicity test data for species from a minimum of eight diverse taxonomic groups. The diversity of tested species is intended to ensure protection of various components of an aquatic ecosystem.
 - The acute freshwater requirement is fulfilled with the following 8 minimum data requirements:
 - the family Salmonidae in the class Osteichthyes
 - a second family in the class Osteichthyes, preferably a commercially or recreationally important warmwater species (e.g., bluegill, channel catfish, etc.)
 - a third family in the phylum Chordata (may be in the class Osteichthyes or may be an amphibian, etc.)
 - a planktonic crustacean (e.g., cladoceran, copepod, etc.)
 - a benthic crustacean (e.g., ostracod, isopod, amphipod, crayfish, etc.)
 - an insect (e.g., mayfly, dragonfly, damselfly, stonefly, caddisfly, mosquito, midge, etc.)
 - a family in a phylum other than Arthropoda or Chordata (e.g., Rotifera, Annelida, Mollusca, etc.)
 - a family in any order of insect or any phylum not already represented
 - The acute estuarine/marine requirement is fulfilled with the following 8 minimum data requirements:
 - two families in the phylum Chordata
 - a family in a phylum other than Arthropoda or Chordata
 - either the Mysidae or Penaeidae family
 - three other families not in the phylum Chordata (may include Mysidae or Penaeidae, whichever was not used above)
 - any other family
 - Chronic toxicity test data (longer-term survival, growth, or reproduction) are required for a minimum of three taxa, with at least one chronic test being from an acutely-sensitive species. Acute-chronic ratios can be calculated with data from species of aquatic animals from at least three different families if the following data requirements are met:
 - at least one is a fish
 - at least one is an invertebrate
 - for freshwater chronic criterion: at least one is an acutely sensitive freshwater species (the other two may be estuarine/marine species) or for estuarine/marine chronic criterion: at least one is an acutely sensitive estuarine/marine species (the other two may be freshwater species).

3. Measures of Effect

Each assessment endpoint requires one or more “measures of ecological effect” which are defined as changes in the attributes of an assessment endpoint itself or changes in a surrogate entity or attribute in response to chemical exposure. Ecological effect data are used as measures of direct and indirect effects to biological receptors. The measures of effect selected represent the growth, reproduction, and survival of the organisms.

- The acute measures of effect used for organisms in this document are the LC50, EC50, and IC50. LC stands for “Lethal Concentration” and LC50 is the concentration of a chemical that is estimated to kill 50% of the test organisms. EC stands for “Effective Concentration” and the EC50 is the concentration of a chemical that is estimated to produce a specific effect in 50% of the test organisms. IC stands for “Inhibitory Concentration” and the IC50 is the concentration of a chemical that is estimated to inhibit some biological process (i.e. growth, etc.) by 50% compared to a control organism.
- Endpoints for chronic measures of exposure are the NOEC, LOEC, and MATC. The NOEC (i.e., “No-Observed-Effect-Concentration”) is the highest test concentration at which none of the observed effects were statistically different from the control. The LOEC (i.e., “Lowest-Observed-Effect-Concentration”) is the lowest test concentration at which observed effects were statistically different from the control. The Maximum Acceptable Toxicant Concentration (MATC) is the calculated geometric mean of the NOEC and LOEC.

Data for carbaryl were obtained from studies submitted to EPA’s Office of Pesticide Programs by registrants to support registration and from studies published in the open literature and identified in a literature search using the ECOTOXicology database (ECOTOX) as meeting data quality standards. ECOTOX is a source of high quality toxicity data for aquatic life, terrestrial plants, and wildlife. The database was created and is maintained by the U.S. EPA, Office of Research and Development, and the National Health and Environmental Effects Research Laboratory's Mid-Continent Ecology Division. The latest comprehensive literature search for this document was conducted in May 2009.

The amount of toxicity testing data available for any given pollutant varies significantly, depending primarily on whether it has raised any significant environmental issues and, in the case of a pesticide, how long it has been registered. A further evaluation of available data is performed by EPA to determine test acceptability. Appendix A of *Quality Criteria for Water*

1986 (U.S. EPA 1986) provides an in-depth discussion of the minimum data requirements and data quality requirements for aquatic life criteria development.

The assessment endpoints for aquatic life criteria are based on growth, reproduction, and survival of the assessed taxa. The measures of effect are provided by the acute and chronic toxicity data. These toxicity endpoints [expressed as genus means] are used in the species sensitivity distribution of the aquatic community to derive the aquatic life criteria. Endpoints used in this assessment are listed in **Table 2**.

Table 2. Summary of Assessment Endpoints and Measures of Effect Used in Criteria Derivation

Assessment Endpoints for the Aquatic Community	Measures of Effect
Survival, growth, and reproduction of freshwater fish, other freshwater vertebrates, and invertebrates	For acute effects: LC50, EC50 For chronic effects: NOEC and LOEC, calculated MATC, or EC20
Survival, growth, and reproduction of estuarine/marine fish and invertebrates	For acute effects: LC50, EC50 For chronic effects: NOEC and LOEC, calculated MATC, or EC20
Maintenance and growth of aquatic plants from standing crop or biomass (freshwater and estuarine/marine)	For effects: LOEC, EC20, EC50, IC50, reduced growth rate, cell viability, calculated MATC

MATC = Maximum acceptable toxicant concentration (geometric mean of NOEC and LOEC)
 NOEC = No observed effect concentration
 LOEC = Lowest observed effect concentration
 LC50 = Lethal concentration to 50% of the test population
 EC50/EC20 = Effect concentration to 50%/20% of the test population
 IC50 = Concentration of carbaryl at which growth is inhibited 50% compared to control organism growth.

Table 3 provides a summary of the number of toxicity data available for genera and species that fulfill the minimum dataset requirements for calculation of acute and chronic criteria for both freshwater and estuarine/marine organisms.

Table 3. Summary Table of Number of Species with Acceptable Toxicity Data Separated into the Minimum Data Requirements in the “Guidelines”

	Genus Mean Acute Value (GMAV)	Species Mean Acute Value (SMAV)	Genus Mean Chronic Value (GMCV)	Species Mean Chronic Value (SMCV)
Freshwater				
Family Salmonidae in the class Osteichthyes	3	9	-	-
Second family in the class Osteichthyes, preferably a commercially or recreationally important warmwater species	20	23	3	1
Third family in the phylum Chordata (may be in the class Osteichthyes or may be an amphibian, etc.)	4 ¹	4 ¹	-	-
Planktonic Crustacean	4	6	2	2
Benthic Crustacean	6	8	-	-
Insect	6	6	-	-
Family in a phylum other than Arthropoda or Chordata (<i>e.g.</i> , Rotifera, Annelida, or Mollusca)	3 ²	3 ²	-	-
Family in any order of insect or any phylum not already represented	1 ³	1 ³	-	-
Total	47	60	5	3
Estuarine/Marine				
Family in the phylum Chordata: phylum Chordata, family Gasterosteidae	1	1	-	-
Family in the phylum Chordata: phylum Chordata, family Cyprinodontidae	1	1	-	-
Either the Mysidae or Penaeidae family: family Mysidae	1	1	-	-
Family in a phylum other than Arthropoda or Chordata	1 ⁴	1 ⁴	-	-
Family in a phylum other than Chordata	1 ⁵	1 ⁵	-	-
Family in a phylum other than Chordata	1 ⁶	1 ⁶	-	-
Family in a phylum other than Chordata	1 ⁷	2 ⁷	-	-
Any other family	1 ⁸	1 ⁸	-	-
Any other family	3 ⁹	3 ⁹	-	-
Total	11	12	-	-

¹ Phylum Chordata, Class Amphibia

² Phylum Annelida and Phylum Mollusca

³ Class Insecta, Family Perlidae

⁴ Phylum Mollusca, Family Tellinidae

⁵ Phylum Mollusca, Family Cardiidae

⁶ Phylum Mollusca, Family Veneridae

⁷ Phylum Mollusca, Family Ostreidae

⁸ Phylum Mollusca, Family Mytilidae

⁹ Phylum Arthropoda

Dash indicates not available

The aquatic plant data available for carbaryl indicate that plants are roughly two orders of magnitude less sensitive than the aquatic animals tested (**Appendix E**). Therefore, no further analyses for development of plant criteria were initiated.

C. Conceptual Model

Conceptual models consist of a written description and diagram (U.S. EPA 1998) that illustrate the relationships between human activities, stressors, and ecological effects on assessment endpoints. The conceptual model links exposure characteristics, with the ecological endpoints important for management goals. Under the Clean Water Act, these management goals are established by states and tribes as designated uses of waters of the United States (for example, aquatic life support). In deriving aquatic life criteria, U.S. EPA is developing acceptable thresholds for pollutants that, if not exceeded, are expected to protect designated uses. A state and/or tribe may implement these criteria by adopting them into their respective water quality standards.

1. Conceptual Diagram

Environmental exposure to carbaryl, while ultimately determined by various site specific conditions and processes, is initiated by an application of the pesticide. The environmental fate properties of carbaryl indicate that runoff, ground water recharge, spray drift, and atmospheric deposition represent potential transport mechanisms of carbaryl to surface water which serves as habitat for aquatic organisms. These transport mechanisms are depicted in the conceptual model below (**Figure 2**). The model also depicts exposure pathways for biological receptors of concern (e.g., non-target aquatic animals) and the potential attribute changes (i.e., effects such as reduced survival, growth and reproduction) in the receptors due to carbaryl exposure.

The conceptual model provides a broad overview of how aquatic organisms can potentially be exposed to carbaryl. Transport mechanisms and exposure pathways are not considered in the derivation of aquatic life criteria. Derivation of criteria focuses on effects on survival, growth and reproduction of aquatic organisms. However, the pathways, receptors, and attribute changes depicted in **Figure 2** may be helpful for states and tribes as they adopt criteria into standards and need to evaluate potential exposure pathways affecting designated uses.

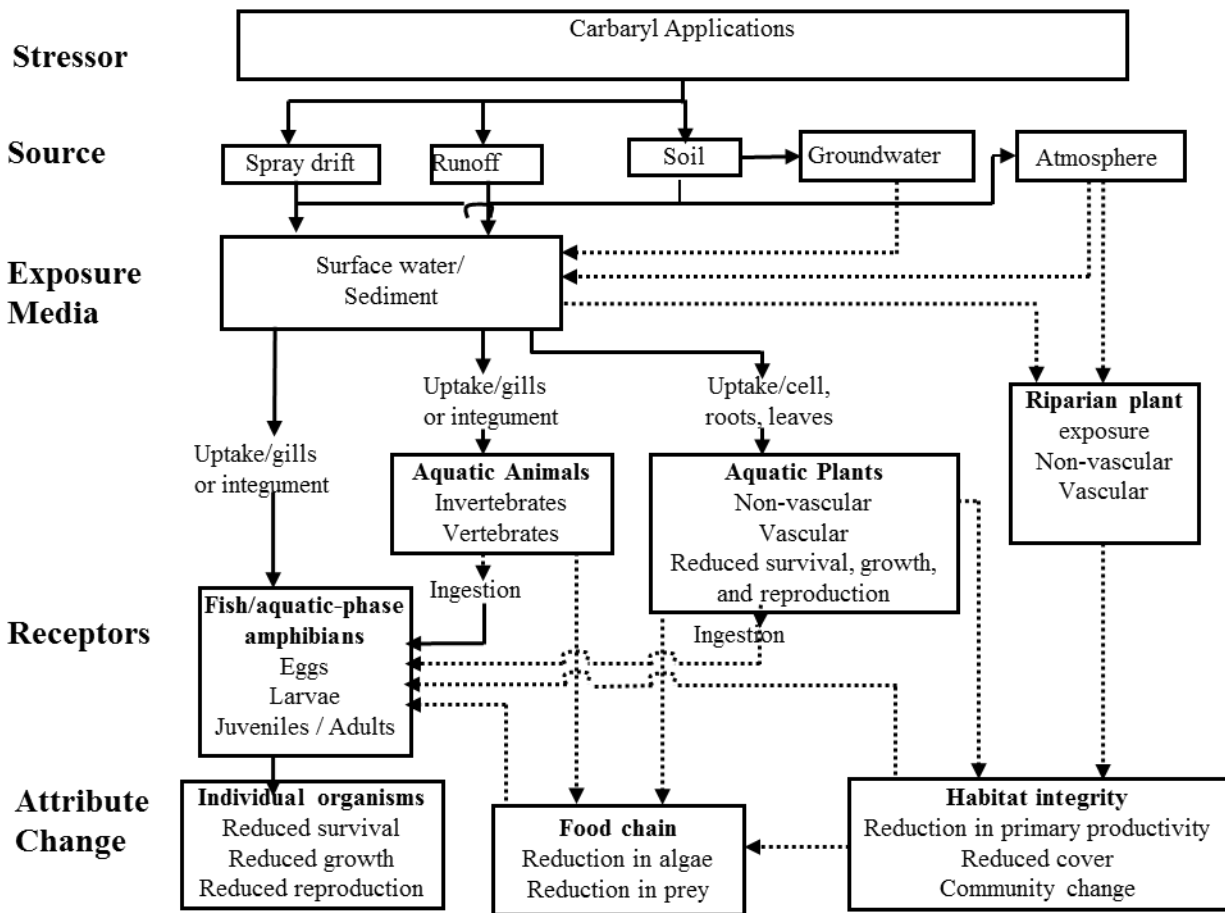


Figure 2: Conceptual model for carbaryl effects on aquatic organisms.

(Dotted lines indicate exposure pathways that have a low likelihood of contributing to ecological effects).

D. Analysis Plan

During development of CWA section 304(a) criteria, U.S. EPA assembles available test data and considers all the relevant data that meet acceptable data quality standard together for all genera. In most cases, data on freshwater and estuarine/marine species are grouped separately to develop separate freshwater and estuarine/marine criteria. Thus, where data allow, four criteria are developed (acute freshwater, acute estuarine/marine, chronic freshwater, and chronic estuarine/marine). If plants are more sensitive than vertebrates and invertebrates, plant criteria are developed.

The CWA criteria are based on a species sensitivity distribution (SSD) comprised of genus mean acute values (GMAVs), calculated from species mean acute values (SMAVs) for acceptable available data. SMAVs are calculated using the geometric mean for all acceptable toxicity tests within a given species (e.g. all tests for *Daphnia magna*). If only one test is available, the SMAV is that test value by default. GMAVs are then calculated using the geometric means of all SMAVs within a given genus (e.g. all SMAVs for genus *Daphnia* - *Daphnia pulex*, *Daphnia magna*). Once again, if only one SMAV is available for a genus, then the GMAV is represented by that value. GMAVs are then rank-ordered by sensitivity from most sensitive to least sensitive. The final acute value (FAV) is determined by regression analysis based on the four most sensitive genera (reflected as GMAVs) in the data set to interpolate or extrapolate (as appropriate) to the 5th percentile of the distribution represented by the tested genera. The acute criterion is the FAV divided by two, which is intended to provide an acute criterion protective of nearly all individuals in such a genus (see “Guidelines”).

Although the aquatic life criteria derivation process relies on selected toxicity endpoints from the sensitive species tested, it does not necessarily mean that the selected toxicity endpoints reflect sensitivity of the most sensitive species existing in a given environment. The intent of the eight minimum data requirements is to serve as a sample representative of the aquatic community. These minimum data requirements represent different ecological, trophic, taxonomic and functional differences observed in the natural aquatic ecosystem. Use of species sensitivity distribution where the criteria values are based on the four most sensitive taxa is reflective of the whole distribution, representing a censored statistical approach that improves estimation of the lower tail when the shape of the whole distribution is uncertain.

The chronic criterion may be determined by one of two methods. If all 8 minimum data requirements are met with acceptable chronic test data, then the chronic criterion is derived using the same method used for the acute criterion. In cases where less chronic data are available (i.e., must have at least three chronic tests from taxa that also have appropriate acute toxicity data) the chronic criterion can be derived by determining an appropriate acute-chronic ratio (ACR). The ACR is a way of relating the acute and chronic toxicities of a material to aquatic organisms. Acute-chronic ratios can be used to derive chronic criteria with data for species of aquatic animals provided that at least three of the minimum data requirements are met and that:

- at least one is a fish
- at least one is an invertebrate
- at least one is an acutely sensitive freshwater species (or estuarine/marine species if estuarine/marine criteria are being derived); the other two species data may be freshwater or estuarine/marine as appropriate to the derivation.

ACRs are calculated by dividing the acute toxicity test values by a “paired” (same lab, same dilution water) chronic test value. Comparisons of ACRs across taxa may elucidate differences and similarities in taxa response. For example, invertebrate ACRs for carbaryl are similar indicating similar taxonomic sensitivity. If variability is greater than ten-fold among calculated ACRs, and no explainable trend exists, then a chronic criterion should not be derived. The Final ACR (FACR) is then derived by calculating the geometric mean of all acceptable ACRs. The Final Chronic Value (FCV) is then estimated by dividing the FAV by the FACR. This serves as the basis for the chronic criterion. Finally, the acute or chronic criterion may be lowered to protect recreationally or commercially important species.

In addition, whenever adequately justified, a state or tribe may replace a national criterion with a site-specific criterion (U.S. EPA 1983a), which may include not only site-specific criterion concentrations, but also site-specific durations of averaging periods and site-specific frequencies of allowed excursions (U.S. EPA 1991). For more information on criteria derivation, see:

http://water.epa.gov/scitech/swguidance/waterquality/standards/upload/2009_01_13_criteria_85g_guidelines.pdf.

The criteria presented herein are the agency’s best estimate of maximum concentrations of carbaryl to protect most aquatic organisms from any unacceptable short- or long-term effects.

Results of such intermediate calculations such as Species Mean Acute Values (**Appendices A and B**) and chronic values (**Appendices C and D**) are specified to four significant figures to prevent round-off error in subsequent calculations and the number of places beyond the decimal point does not reflect the precision of the value.

E. Identification of Data Gaps for Aquatic Organisms

Data gaps were identified for carbaryl. No acceptable aquatic vascular plant data are available. However, given the large difference between toxicity to aquatic plants and animals, it is not likely that aquatic vascular plants would be more sensitive to carbaryl than aquatic animals. Also, additional acute and chronic aquatic toxicity studies using the degradate 1-naphthol on fish and invertebrates are recommended to address lack of information for this degradate. This data gap was identified in EPA's problem formulation for the re-evaluation of carbaryl (U.S. EPA 2010a) thus, in the future, additional degradate data may become available. In addition, supplemental estuarine/marine chronic studies would be beneficial since there is only one study available for evaluation.

III. EFFECTS ANALYSES TO AQUATIC ORGANISMS

A. Acute Toxicity to Aquatic Animals

All available data relating to the acute effects of carbaryl on aquatic animals were considered in deriving the carbaryl criteria. Data that were suitable (in terms of test acceptability and quality), according to the "Guidelines," for the derivation of a freshwater and estuarine/marine FAV are presented in **Appendices A and B, respectively** (most fish and invertebrate data are 96-hr duration, except cladocerans, midges, mysids and certain embryos and larvae of specific estuarine/marine groups are 48-hr duration).

1. Freshwater

Sixty freshwater species representing 47 genera were represented in the dataset of acceptable data for acute toxicity to carbaryl. Species Mean Acute Values (SMAV) ranged from 3.175 µg/L for the stonefly (*Isogenus sp.*) to 27,609 µg/L for the catfish (*Clarias batrachus*).

The second, third, fourth and seventh most sensitive tested species SMAVs are also stoneflies (*Skwala sp.*, *Pteronarcys californica*, *Claassenia sabulosa* and *Pteronarcella badia*), with SMAVs of 3.6, 4.8, 5.6 and 9.163 µg/L, respectively. Cladocerans are the next most sensitive taxon (SMAVs ranged from 5.958 µg/L for *Ceriodaphnia dubia* to 35 µg/L for *Daphnia carinata*), followed by amphipods (*Gammarus pseudolimnaeus*, *G. lacustris*, *Hyaella azteca* and *Pontoporeia hoyi* with SMAVs of 10.12, 18.76, 15.2 and 250 µg/L, respectively). Thus, the ten freshwater genera most sensitive to carbaryl are in the classes Insecta and Crustacea (**Table 4**), as would be expected for an insecticide.

The most sensitive fish SMAV was for brown trout, *Salmo trutta* (SMAV of 700 µg/L), followed by the rainbow trout (SMAV of 860 µg/L) and lake trout (*Salvelinus namaycush*, SMAV of 988.1 µg/L). Tests relating to effects on several endangered fish species were also available: the shortnosed sturgeon, *Acipenser brevirostrum*, with a SMAV of 1,810 µg/L (Dwyer et al. 2000); the razorback sucker, *Xyrauchen texanus*, with a SMAV of 4,350 µg/L (Dwyer et al. 1995); the Gila topminnow, *Poeciliopsis occidentalis*, with a SMAV of >3,000 µg/L (Dwyer et al. 1999b); the bonytail chub, *Gila elegans*, with a SMAV of 2,655 µg/L (Dwyer et al. 1995; Beyers et al. 1994); the Atlantic salmon, *Salmo salar*, with a SMAV of 1,129 µg/L (Mayer and Ellersieck 1986); and the Colorado pikeminnow (formerly squawfish), *Ptychocheilus lucius*, with a SMAV of 2,005 µg/L (Dwyer et al. 1995; Beyers et al. 1994). Tests relating to effects on one threatened fish species was available, i.e., the Apache trout, *Oncorhynchus apache*, with a SMAV of 1,540 µg/L (Dwyer et al. 1995). Tests relating to effects on two threatened/endangered fish species (certain populations threatened, and others endangered) were also available, the Coho salmon, *Oncorhynchus kisutch*, with a SMAV of 1,654 µg/L (Katz 1961; Macek and McAllister 1970; Post and Schroeder 1971; Johnson and Finley 1980; Mayer and Ellersieck 1986) and the Chinook salmon, *Oncorhynchus tshawytscha*, with a SMAV of 2,690 µg/L (Johnson and Finley 1980; Mayer and Ellersieck 1986; Phipps and Holcombe 1985; 1990). Available data indicates endangered fish species tested are not found to be more sensitive than non-endangered fish species.

Aquatic-phase amphibians (*Rana clamitans*, SMAV = 16,296 µg/L; *Bufo boreas*, SMAV = 12,310 µg/L; *Hyla versicolor*, SMAV = 2,470 µg/L and *Xenopus laevis* (frog that is a common test species with fully aquatic life cycle that is non-native to the U.S.), SMAV = 1,730 µg/L), freshwater mussels (*Anodonta imbecillis*, SMAV = 24,632 µg/L) and the aquatic air-breathing

snail (*Aplexa hypnorum*, SMAV = >27,000 µg/L) were comparatively insensitive to carbaryl. The least sensitive species tested with carbaryl is *C. batrachus*, with a SMAV of 27,609 µg/L.

GMAVs for 47 freshwater genera are provided in **Table 4**. The four most sensitive genera were within a factor of 1.8 of one another. The freshwater FAV (the 5th percentile of the species sensitivity distribution) for carbaryl is 4.219 µg/L, calculated using the procedures described in the “Guidelines.” The Final Acute Value is an estimate of the concentration of the material corresponding to a cumulative probability of 0.05 in the acute toxicity values for the genera with which acceptable acute tests have been conducted on the material. The FAV is slightly higher than the GMAVs for two genera of stoneflies, *Isogenus* (3.175 µg/L) and *Skwala* (3.6 µg/L). The FAV is then divided by two to account for the fact that the toxicity tests were designed to assess LC50 values, and the criterion needed to protect aquatic life at levels below which effects on test organisms are indistinguishable from control (unexposed) organisms. Therefore, the FAV/2, which is the freshwater continuous maximum concentration (CMC), for carbaryl is 2.1 µg/L and should be protective for all freshwater organisms potentially exposed to carbaryl under short-term conditions (**Figure 3**).

Table 4. Ranked Freshwater Genus Mean Acute Values

Rank ^a	Genus Mean Acute Value (µg/L)	Species	Species Mean Acute Value ^b (µg/L)
47	27,609	Walking catfish, <i>Clarias batrachus</i>	27,609
46	>27,000	Snail, <i>Aplexa hypnorum</i>	>27,000
45	24,632	Mussel, <i>Anodonta imbecillis</i>	24,632
44	20,000	Black bullhead, <i>Ameiurus melas</i>	20,000
43	16,700	Goldfish, <i>Carassius auratus</i>	16,700
42	16,296	Green frog, <i>Rana clamitans</i>	16,296
41	12,400	Channel catfish, <i>Ictalurus punctatus</i>	12,400
40	12,310	Boreal toad, <i>Bufo boreas</i>	12,310
39	9,039	Green sunfish, <i>Lepomis cyanellus</i>	9,460
	-	Redear sunfish <i>L. microlophus</i>	11,200
	-	Bluegill, <i>L. macrochirus</i>	6,970
38	8,656	European chub, <i>Leuciscus cephalus</i>	8,656
37	8,200	Oligochaete worm, <i>Lumbriculus variegatus</i>	8,200
36	8,012	Fathead minnow, <i>Pimephales promelas</i>	8,012
35	6,400	Largemouth bass, <i>Micropterus salmoides</i>	6,400
34	4,350	Razorback sucker, <i>Xyrauchen texanus</i>	4,350
33	4,153	Common carp, <i>Cyprinus carpio</i>	4,153
32	>3,000	Gila topminnow, <i>Poeciliopsis occidentalis</i>	>3,000
31	2,930	Nile tilapia, <i>Oreochromis niloticus</i>	2,930
30	2,655	Bonytail chub, <i>Gila elegans</i>	2,655
29	2,600	Black crappie, <i>Pomoxis nigromaculatus</i>	2,600
28	2,515	Guppy, <i>Poecilia reticulata</i>	2,515

Table 4. Ranked Freshwater Genus Mean Acute Values

Rank ^a	Genus Mean Acute Value (µg/L)	Species	Species Mean Acute Value ^b (µg/L)
27	2,480	Yellow perch, <i>Perca flavescens</i>	2,480
26	2,470	Gray tree frog, <i>Hyla versicolor</i>	2,470
25	2,462	Crayfish, <i>Orconectes immunis</i>	2,870
	-	Crayfish, <i>O. virilis</i>	2,112
24	2,079	Greenthroat darter, <i>Etheostoma lepidum</i>	2,140
	-	Fountain darter, <i>E. fonticola</i>	2,020
23	2,005	Colorado pikeminnow (formerly squawfish), <i>Ptychocheilus lucius</i>	2,005
22	1,810	Apache trout, <i>Oncorhynchus apache</i>	1,540
	-	Coho salmon, <i>O. kisutch</i>	1,654
	-	Chinook salmon, <i>O. tshawytscha</i>	2,690
	-	Cutthroat trout, <i>O. clarkii</i>	3,300
	-	Rainbow trout, <i>O. mykiss</i>	860
21	1,810	Shortnosed sturgeon, <i>Acipenser brevirostrum</i>	1,810
20	1,730	African clawed frog, <i>Xenopus laevis</i>	1,730
19	1,322	Striped bass, <i>Morone saxatilis</i>	1,322
18	1,269	Brook trout, <i>Salvelinus fontinalis</i>	1,629
	-	Lake trout, <i>S. namaycush</i>	988.1
17	1,000	Crayfish, <i>Procambarus clarkii</i>	1,000
16	889.0	Atlantic salmon, <i>Salmo salar</i>	1,129
	-	Brown trout, <i>S. trutta</i>	700
15	839.6	Crayfish, <i>Cambarus bartoni</i>	839.6

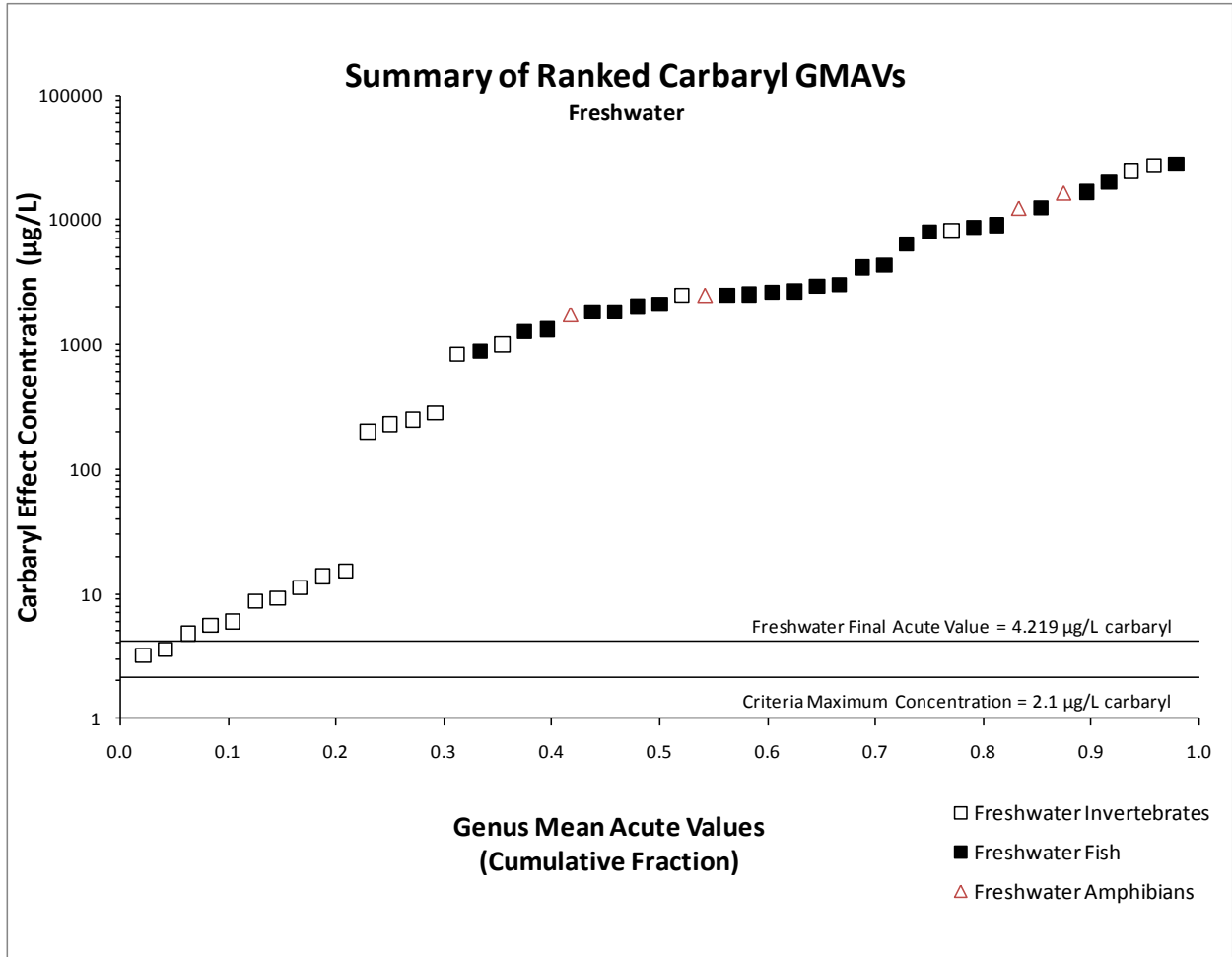
Table 4. Ranked Freshwater Genus Mean Acute Values

Rank ^a	Genus Mean Acute Value (µg/L)	Species	Species Mean Acute Value ^b (µg/L)
14	280	Aquatic sowbug, <i>Asellus brevicaudus</i>	280
13	250	Amphipod, <i>Pontoporeia hoyi</i>	250
12	230	Mysid, <i>Mysis relicta</i>	230
11	200	Backswimmer, <i>Notonecta undulate</i>	200
10	15.2	Amphipod, <i>Hyalella azteca</i>	15.2
9	13.78	Amphipod, <i>Gammarus lacustris</i>	18.76
	-	Amphipod, <i>G. pseudolimnaeus</i>	10.12
8	11.90	Cladoceran, <i>Daphnia carinata</i>	35
	-	Cladoceran, <i>D. magna</i>	7.521
	-	Cladoceran, <i>D. pulex</i>	6.4
7	9.163	Stonefly, <i>Pteronarcella badia</i>	9.163
6	8.781	Cladoceran, <i>Simocephalus serrulatus</i>	8.781
5	5.958	Cladoceran, <i>Ceriodaphnia dubia</i>	5.958
4	5.6	Stonefly, <i>Claassenia sabulosa</i>	5.6
3	4.8	Stonefly, <i>Pteronarcys californica</i>	4.8
2	3.6	Stonefly, <i>Skwala sp.</i>	3.6
1	3.175	Stonefly, <i>Isogenus sp.</i>	3.175

^a Ranked from the most resistant to the most sensitive based on Genus Mean Acute Value.

^b From **Appendix A**.

Figure 3: Ranked Summary of Carbaryl Genus Mean Acute Values (GMAVs) - Freshwater Animals



2. Estuarine/Marine

SMAVs (SMAVs and GMAVs are equal with the exception of the genus *Crassostrea*) for 11 genera representing 12 species of estuarine/marine organisms were calculated for carbaryl (**Table 5**). SMAVs and GMAVs are equal when there is only one species present per genus. The most sensitive genus was the mysid (*Americamysis sp.*), with a SMAV of 7.188 µg/L, followed by the Dungeness crab (*Metacarcinus magister*) with a SMAV of 10 µg/L. The two most tolerant genera were the bent-nosed clam (*Macoma nasuta*) and the threespine stickleback (*Gasterosteus aculeatus*), with SMAVs of 17,000 and 3,990 µg/L, respectively (**Figure 4**).

The four most sensitive estuarine/marine genera are *Americamysis*, *Metacarcinus*, *Callinassa* and *Upogebia* with GMAVs that differ by a factor of approximately 8.3. The ghost shrimp (*Callinassa californiensis*) and mud shrimp (*Upogebia pugettensis*) studies were conducted under static conditions with measured concentrations over a 48-hr duration (Stewart et al. 1967). Although the test duration is not the recommended 96-hr duration, these two datapoints were determined to be acceptable and are included in the estuarine/marine acute criteria calculation. Acetylcholinesterase inhibition studies and recovery studies using carbaryl indicate that results for 48-hr acute toxicity tests with invertebrates would be similar to results from 96-hr tests. Scaps et al. (1997) determined the maximum percentage of acetylcholinesterase activity inhibition occurred at 48 hours after exposure and then remained stable for up to seven days for the estuarine/marine polychaete worm, *Nereis diversicolor*. Therefore, the maximum percentage of acetylcholinesterase activity inhibition at 48 hours would lead to the highest level of immobilization or death for the test organisms. Parsons and Surgeoner (1991) found that third-instar mosquito larvae (*Aedes aegypti*) did not recover from immobilization that occurred after a 24-hr exposure to carbaryl. The ability of larvae to recover from immobilization following exposure to carbaryl decreased with increasing exposure time. These results indicate that a 48-hr acute toxicity test EC50 or LC50 would be similar to those for 96-hr tests and provide justification for the use of 48-hr results in the criterion calculation.

Bivalves are moderately insensitive to carbaryl, with GMAVs ranging from 1,650 µg/L for the bay mussel (*Mytilus trossulus*) to 3,850 µg/L for cockle clam (*Clinocardium nuttallii*). The threespine stickleback (*G. aculeatus*) has a similar SMAV of 3,990 µg/L. The most sensitive genus (*Americamysis*, GMAV of 7.188 µg/L) is greater than 2,365 times more sensitive than the most tolerant genus tested, *M. nasuta* (SMAV = 17,000 µg/L).

The “Guidelines” indicate that eight minimum data requirements are needed to calculate an estuarine/marine FAV; data are available for 11 genera and meet the family requirement outlined above. The estuarine/marine FAV is 3.15 µg/L (**Table 5**). The estuarine/marine CMC (1.58 µg/L) is protective of all estuarine/marine organisms acutely exposed to carbaryl (**Figure 4**).

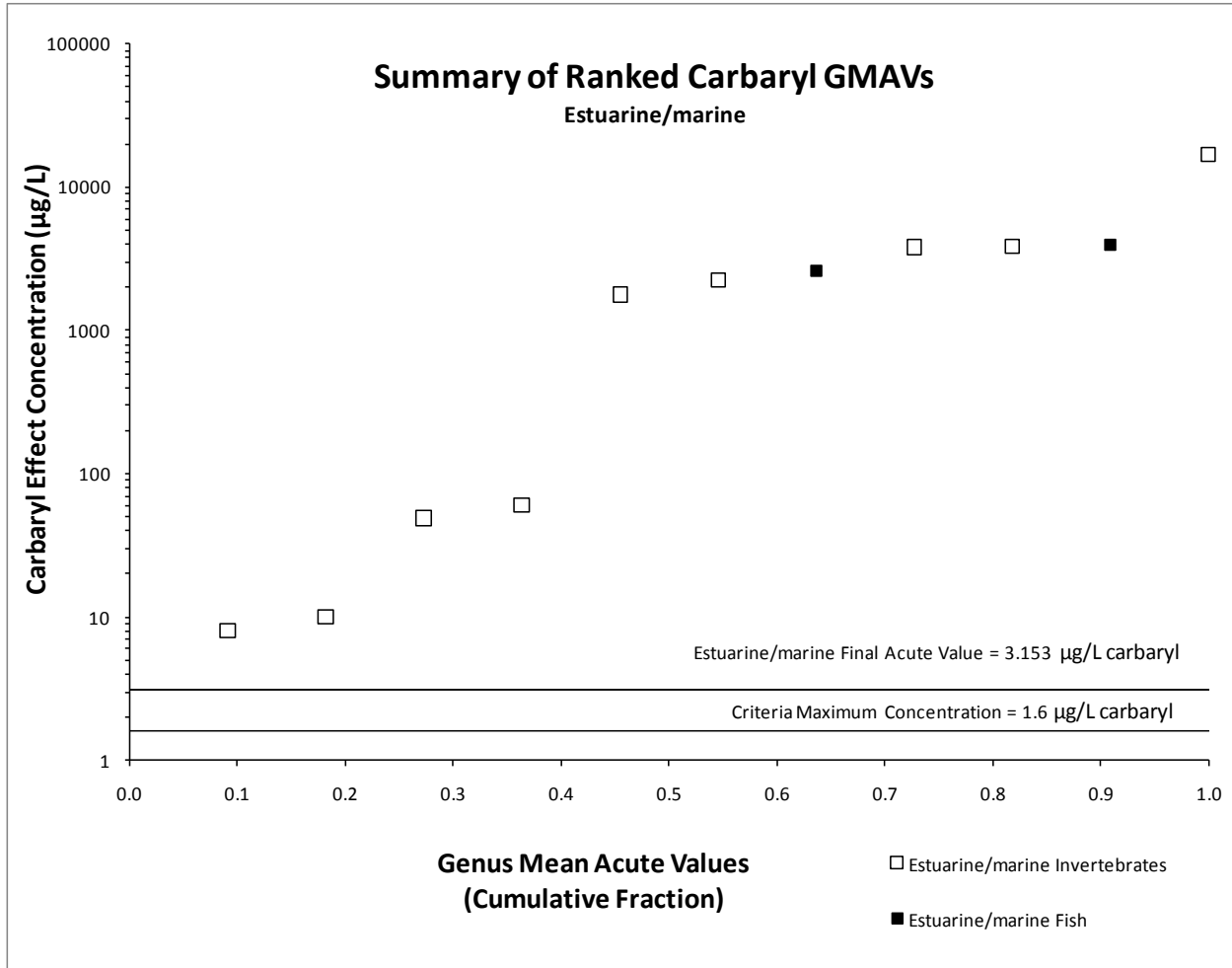
Table 5. Ranked Estuarine/Marine Genus Mean Acute Values

Rank ^a	Genus Mean Acute Value (µg/L)	Species	Species Mean Acute Value ^b (µg/L)
11	17,000	Bent-nosed clam, <i>Macoma nasuta</i>	17,000
10	3,990	Threespine stickleback, <i>Gasterosteus aculeatus</i>	3,990
9	3,850	Cockle clam, <i>Clinocardium nuttalli</i>	3,850
8	3,820	Hard clam, <i>Mercenaria mercenaria</i>	3,820
7	2,600	Sheepshead minnow, <i>Cyprinodon variegatus</i>	2,600
6	2,291	Pacific oyster, <i>Crassostrea gigas</i>	2,200
	-	Eastern oyster, <i>C. virginica</i>	2,386
5	1,650	Bay mussel, <i>Mytilus edulis</i>	1,650
4	60.0	Mud shrimp, <i>Upogebia pugettensis</i>	60.0
3	48.99	Ghost shrimp, <i>Callinassa californiensis</i>	48.99
2	10	Dungeness crab, <i>Metacarcinus magister</i> formerly <i>Cancer magister</i>	10
1	7.188	Mysid, <i>Americamysis bahia</i>	7.188

^a Ranked from the most resistant to the most sensitive based on Genus Mean Acute Value.

^b From **Appendix B**.

Figure 4: Ranked Summary of Carbaryl Genus Mean Acute Values (GMAVs) - Estuarine/Marine Animals



B. Chronic Toxicity to Aquatic Animals

1. Freshwater

Freshwater chronic toxicity data that meet the test acceptability and quality assurance/control criteria in the “Guidelines” are presented in **Appendix C**. All tests were conducted with measured concentrations of carbaryl. Carbaryl chronic toxicity data are available for five species of freshwater organisms: two invertebrate species (cladocerans) and three fish species.

Carbaryl was evaluated by Oris et al. (1991) using the cladoceran, *Ceriodaphnia dubia*. The pH of the seven-day static-renewal test was 8.18. The mean total young per female was the most sensitive endpoint with a final chronic value (geomean of NOEC and LOEC) of 10.6 µg/L. Replicate seven-day reproduction toxicity tests using hypothesis testing and median inhibition concentration interpolation analysis (median IC50 value approximates chronic values) yielded the same final chronic value. Division of the 48-hr LC50 acute value of 11.6 µg/L from the same study (**Appendix A**) by the chronic value of 10.6 µg/L results in an acute-chronic ratio (ACR) of 1.094 for *C. dubia* (**Table 8**).

Two chronic exposures have been conducted with the cladoceran, *Daphnia magna*, to carbaryl (Brooke 1991 and Surprenant 1985b). *Daphnia magna* was exposed for 21 days to five measured concentrations of carbaryl - 0.29, 0.58, 1.07, 2.16, and 4.04 µg/L at a mean pH of 8.2 (Brooke 1991). Neither survival of the adults nor the number of young produced was significantly different from that of the control organisms at concentrations ≤ 4.04 µg/L. Since there was no significant difference at the highest tested concentration of 4.04 µg/L, the “Guidelines” stipulate that a greater than value (>) be assigned. Therefore, the “no-effect” concentration range for *D. magna* exposed to carbaryl was between 4.04 µg/L and the 48-hr EC50 of 10.1 µg/L. Given the 48-hr EC50 value of 10.1 µg/L from the acute toxicity test (Brooke 1991) this would produce a “theoretical” ACR ranging from 1.0 to < 2.5. This theoretical ACR, while not usable in the direct calculation of the chronic criteria, is consistent with the ACR from freshwater invertebrate taxa that is available, 1.094 for *Daphnia magna*. This indicates that acute and chronic toxicities for the invertebrate taxa are closely related.

Another study with the cladoceran, *D. magna*, was conducted by Surprenant (1985b). The author exposed the cladoceran for 21 days in a flow-through measured exposure. The most

sensitive endpoint was an effect of reproduction observed at 3.3 µg/L when compared to the solvent control. The chronic value was calculated to be 2.225 µg/L. An ACR was not derived for this study because there was not an associated acute test.

Two chronic exposures have been conducted with fathead minnows to carbaryl (Carlson 1971 and Norberg-King 1989). Both tests were conducted in the same laboratory by different researchers in different years and with different test durations. Carlson (1971) exposed fathead minnows through a complete life-cycle test beginning with 1- to 5-day old fry at 5 exposure concentrations, i.e., 8, 17, 62, 210, and 680 µg/L. The exposure lasted 9 months resulting in reduced survival at 6 months in the highest exposure concentration of 680 µg/L. After 9 months at the same concentration, fish exhibited reduced egg production and no larvae hatched. No statistically significant effects were noted for survival, growth or reproduction at the 210 µg/L exposure. Thus, the chronic value of 378 µg/L for this study is the geometric mean of the lowest observed effect concentration (LOEC) of 680 µg/L and the no observed effect concentration (NOEC) of 210 µg/L based on reproduction. Division of the 96-hr LC50 of 9,000 µg/L from the same study by the chronic value of 377.9 µg/L results in an ACR of 23.82.

A second chronic test with the fathead minnow was conducted as a 32-day early-life-stage test (Norberg-King 1989). No adverse effects in fish were observed from exposure to carbaryl at concentrations ≤720 µg/L, but adverse effects were measured at 1,600 µg/L. Growth in length, but not weight was reduced at 1,600 µg/L, as was survival. The chronic value for fathead minnows in this test is 1,073 µg/L, which is the geometric mean of the lowest observed effect concentration (LOEC) of 1,600 µg/L and the no observed effect concentration (NOEC) of ≤720 µg/L. Calculation of an ACR from this study is not possible since the authors did not measure the concentration of the exposure in the acute toxicity test. However, two other acute tests for fathead minnows have been conducted at the same laboratory since 1971. Carlson (1971) reported a value of 9,000 µg/L, and Phipps and Holcombe (1985) a value of 5,010 µg/L. The geometric mean of these values is 6,715 µg/L which results in an ACR of 6.258 for the early-life-stage chronic test. The ACR for the complete life cycle test is 23.82, which shows more sensitivity to carbaryl chronic toxicity.

Beyers et al. (1994) conducted individual 32-day early life-stage (ELS) chronic toxicity tests with the endangered Colorado pikeminnow (formerly squawfish), *Ptychocheilus lucius*, and the bonytail chub, *Gila elegans*. In both studies, however, considerable ontogenetic development

of the test organisms had occurred before test initiation whereby the required embryo and protolarva life stages were not present during the exposure period (mesolarval, metalarval and juvenile life stages were present). Thus, the ELS tests were initiated with 41-day old pikeminnow larvae and 48-day old chub larvae. In both tests, growth was as sensitive as or more sensitive than survival as a measure of chronic toxic effects, and was the only value reported. The NOEC and LOEC for Colorado pikeminnow and bonytail chub were 445 and 866 µg/L, and 650 and 1,240 µg/L, respectively. The final chronic value for *P. lucius* was 620.8 µg/L and 897.8 µg/L for *G. elegans*. ACR values could not be determined for these studies because an accompanying flow-through measured acute toxicity value was not available for each species, nor were there any other available appropriate acute toxicity tests for these species.

Table 6. Ranked Freshwater Genus Mean Chronic Values

Rank ^a	Genus Mean Chronic Value (µg/L)	Species	Species Mean Chronic Value ^b (µg/L)
5	897.8	Bonytail chub, <i>Gila elegans</i>	897.8
4	636.8	Fathead minnow, <i>Pimephales promelas</i>	636.8
3	620.8	Colorado pikeminnow (formerly squawfish), <i>Ptychocheilus Lucius</i>	620.8
2	10.6	Cladoceran, <i>Ceriodaphnia dubia</i>	10.6
1	3.770	Cladoceran, <i>Daphnia magna</i>	3.770

^a Ranked from the most resistant to the most sensitive based on Genus Mean Chronic Value.

^b From **Appendix C**.

2. Estuarine/Marine

One carbaryl estuarine/marine chronic toxicity test is available for inclusion in the document. A 28-day flow-through test with measured concentrations initiated with 24-hr old neonates was conducted with the mysid, *Americamysis bahia* (Thursby and Champlin 1991b). The NOEC and LOEC based on survival were 7.18 µg/L and 13.7 µg/L, respectively for a final chronic value of 9.918 µg/L carbaryl. The total percent control survival of 63% is below the acceptable total percent control survival of 70% required in ASTM guide E1191-03a (ASTM

2008). The percent control survival for mating pairs is 75% and exceeds the ASTM requirement of 70%. However, the number of young produced by the first-generation females in the controls was less than three which is an ASTM requirement. Two young were produced by the first-generation females in the control and is below the ASTM requirement of three. Therefore, this study is used qualitatively and used in the discussion of acute-chronic ratios. The calculated ACR of 0.8530 is based on the 96-hr LC50 of 8.46 µg/L reported for the same study.

Table 7. Ranked Estuarine/Marine Genus Mean Chronic Values

Rank ^a	Genus Mean Chronic Value (µg/L)	Species	Species Mean Chronic Value ^b (µg/L)
1	9.918	Mysid, <i>Americamysis bahia</i>	9.918

^a Ranked from the most resistant to the most sensitive based on Genus Mean Chronic Value.

^b From **Appendix D**.

3. Acute-Chronic Ratio

Four valid ACRs are available for carbaryl using the fifth, eighth and thirty-sixth most sensitive genera of freshwater organisms (**Table 4**). Since the difference between the lowest ACR (1.094 for *Ceriodaphnia dubia*) and the highest ACR (23.82 for *Pimephales promelas*) is a factor of 22, and since the “Guidelines” stipulate that if the species mean ACR seems to increase as the SMAV increases, the FACR (final ACR) should be calculated as the geometric mean of the ACRs for species whose SMAVs are close to the FAV. Using this recommendation, the FACR would be the geometric mean of 1.094 (*C. dubia*) and 1.581 (*D. magna*), or 1.315. However, the “Guidelines”, also stipulate that if the most appropriate species mean ACRs are less than 2.0, the FACR should be assumed to be 2.0. Low ACRs may reflect acclimation to the toxicant during the chronic test, or test organisms being fed vs. unfed which may affect bioavailability and susceptibility. Thus, the FACR for carbaryl is assumed to be 2.0 for freshwater organisms.

Dividing the freshwater FAV of 4.219 by the ACR of 2.0 results in a freshwater final chronic value (FCV) of 2.1 µg/L. It is concluded that all freshwater species will be protected

(Figures 3 and 5) from adverse effects due to chronic carbaryl exposure with the freshwater FCV of 2.1 µg/L.

Table 8. Acute-Chronic Ratios

Species	Acute Value (µg/L)	Chronic Value (µg/L)	Ratio	Reference
Freshwater Species				
Cladoceran, <i>Ceriodaphnia dubia</i>	11.6	10.6	1.094	Oris et al. 1991
Cladoceran, <i>Daphnia magna</i>	10.1	6.389 ^a	1.581	Brooke 1991
Fathead minnow, <i>Pimephales promelas</i>	9,000	377.9	23.82	Carlson 1971
Fathead minnow, <i>Pimephales promelas</i>	6,715	1,073	6.258 ^b	Norberg-King 1989

^aThe chronic value of 6.389 µg/L is calculated as the geometric mean of the highest chronic test exposure concentration (4.04 µg/L) which had no adverse impact and the 48-hr EC50 value of 10.1 µg/L from the acute toxicity test.

^b Only data from Carlson (1971) used to calculate ACR for this species.

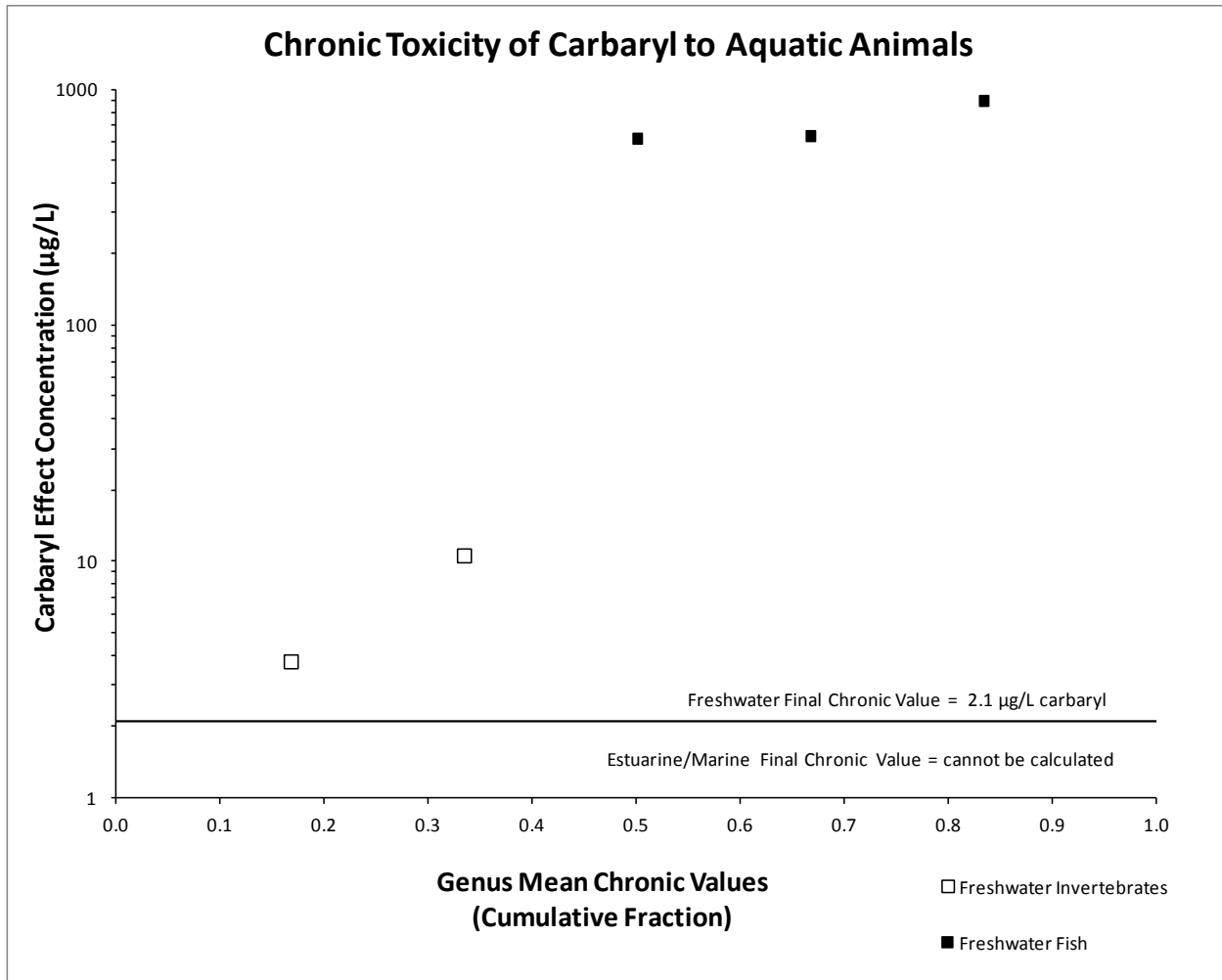
Freshwater

Final Acute Value = 4.219 µg/L

Final Acute-Chronic Ratio = 2.0

Final Chronic Value = (4.219 µg/L)/2.0 = **2.1 µg/L**

Figure 5: Chronic Toxicity of Carbaryl to Aquatic Animals



C. Toxicity to Aquatic Plants

No carbaryl toxicity tests with important aquatic plant species in which the concentrations of test material were measured and the endpoint was biologically important are available in the literature. Therefore, a Final Plant Value cannot be determined. Effects on aquatic plants are discussed qualitatively in the Effects Characterization chapter.

D. Degradate Toxicity

Acute toxicity testing of carbaryl's major degradate 1-naphthol in fish shows that the compound ranged from being moderately to highly toxic (LC50 range 750 to 1,600 µg/L). The toxic mode of action for 1-naphthol is thought to be narcosis (Russom et al. 1997). In aquatic organisms, narcosis is a reversible anesthetic effect that is caused by chemicals partitioning into cell membranes and nervous tissue that result in disruption of central nervous system function (Barron et al. 2001). Chronic exposure of fathead minnows to 1-naphthol reduced larval growth and survival with a NOEC = 100 µg/L (U.S. EPA 2010a). No data are available on the acute or chronic toxicity of 1-naphthol to amphibians. For freshwater invertebrates, 1-naphthol ranged from moderately to highly toxic with an EC50 range: 200 to 3,000 µg/L (U.S. EPA 2010a). Toxicity data are not available for the other major degradate, 1,4-naphthoquinone which is a data gap.

E. Summary of National Criteria

The resulting recommended ambient water quality criteria indicate that freshwater aquatic organisms would have an appropriate level of protection if the one-hour average concentration does not exceed 2.1 µg/L more than once every three years on average and if the four-day average concentration of carbaryl does not exceed 2.1 µg/L more than once every three years on average (except where a locally important species may be more sensitive). Estuarine/marine aquatic organisms would have an appropriate level of protection if the one-hour average concentration does not exceed 1.6 µg/L more than once every three years on average (except where a locally important species may be more sensitive). Note: a Criterion Continuous

Concentration (CCC) could not be calculated for estuarine/marine aquatic organisms due to insufficient data.

Table 9. Summary Table of Aquatic Life Criteria for Carbaryl

	Acute	Chronic
Freshwater	2.1 µg/L	2.1 µg/L
Estuarine/Marine	1.6 µg/L	N/A

N/A – not available, unable to calculate estuarine/marine chronic criterion

IV. EFFECTS CHARACTERIZATION

Carbaryl is expected to be mobile, but degrades rapidly in neutral to alkaline aquatic systems; however, under acidic conditions, carbaryl is more persistent. Its environmental fate characteristics are consistent with other carbamate insecticides (e.g. carbofuran). Based on environmental fate data, potential exposure would be expected to be greater in aquatic environments with pH less than 7. Fate data on the primary degradate, 1-naphthol, has been identified as a data gap. Although 1-naphthol appears to be somewhat mobile, it is not likely to persist due to fairly rapid degradation.

The primary mode of action for carbaryl is acetylcholinesterase inhibition. This effect is reversible with removal of exposure to carbaryl leading to the possibility of recovery of test organisms at sub-lethal concentrations. The following information addresses studies that are not used in the calculation of criteria but provide supporting evidence for the validity of the criteria.

A. Effects on Aquatic Animals

1. Freshwater Acute Toxicity

Acceptable acute toxicity data are available for 60 freshwater species representing 47 genera and represents a dataset supporting development of the acute criterion. In general, technical grade carbaryl is classified as highly toxic to fish, aquatic-phase amphibians and is very highly toxic to aquatic invertebrates on an acute exposure basis (U.S. EPA 2010a). Chronic

exposure to carbaryl resulted in decreased growth in freshwater fish and decreased reproduction in freshwater invertebrates.

Invertebrate species, particularly arthropods (e.g., daphnids, mysids) are the most sensitive, with the 10 lowest SMAVs (and GMAVs) ranging from 3.175 to 15.2 µg/L, and represented by the classes Insecta and Crustacea (**Table 4**). Cladocerans and amphipods showed high sensitivity to carbaryl in acute toxicity tests. The most sensitive genus tested is the stonefly *Isogenus*, which is more than 8,600 times as sensitive as the most resistant genus tested, i.e., the walking catfish *Clarias* (**Figure 3**).

Several studies were identified as not meeting screening guidelines for inclusion in criterion calculations (**Appendix H**), but showed similar ranges of toxicity and are presented here to provide additional supporting evidence of the potential toxicity of carbaryl to aquatic organisms. *Daphnia magna* LC50 values ranged from 0.66 µg/L (Rawash et al. 1975) to 21 µg/L (Wernersson and Dave 1997) for exposures lasting 24 hours. Mayer and Ellersieck (1986) showed similar sensitivities to the amphipod species *Gammarus pseudolimnaeus*, with 48-hr measured LC50 values of 8 and 13 µg/L carbaryl. Several species of Diptera larvae were also exposed to carbaryl; larvae of midges appear to be more sensitive than larvae of mosquitoes. The LC50 values (24-hr) of various midge species ranged from 1.0 to 110 µg/L. These studies were not used in quantitative criterion calculations because test durations were insufficient.

A study of the persistence of carbaryl toxicity was conducted by Mayer and Ellersieck (1986). Stonefly nymphs (*Isogenus sp.*) were exposed to carbaryl solutions which were fresh (**Appendix A**) and others that had been aged 7, 14 or 21 days (**Appendix H**). Aged solutions were mixed and allowed to sit under normal laboratory conditions. The results showed a trend of decreased toxicity with aging of the solutions. The toxicity of carbaryl solutions aged 21 days decreased the toxicity to about one-third that of fresh solutions. These data support conclusions based on environmental fate properties that the “aging” process decreased exposure concentration, thus decreasing toxicity. Results from the aged solutions were not used in the quantitative criterion calculations because the concentrations of carbaryl were not measured at the initiation or completion of the aged solution toxicity tests.

In general, fish are less sensitive to carbaryl than invertebrates. Of the fish tested, brown trout, *Salmo trutta* (SMAV of 700 µg/L) is the most sensitive tested vertebrate species by a factor of approximately 39 times compared to the least sensitive fish tested, i.e., walking catfish,

Clarias batrachus, with a SMAV of 27,609 µg/L. Mayer and Ellersieck (1986) exposed cutthroat trout (*Oncorhynchus clarkii*) to carbaryl solutions aged for 0, 7, 14 and 21 days. Again, results from the aged solutions were not used in the quantitative criterion calculations because the concentrations of carbaryl were not measured at the initiation of the aged solution toxicity tests. Cutthroat trout exposed to fresh carbaryl solutions had a 96-hr LC50 of 6,800 µg/L at pH= 7.3 (**Appendix A**), and a LC50 of 2,300 µg/L at pH= 7.2 when exposed to a solution aged 21 days. The results from this study show enhanced sensitivity to aged carbaryl solutions and are not consistent with the similar “aged” solution study with stonefly nymphs (Mayer and Ellersieck 1986). These data may indicate potential taxa differences or differences in responses to degradates.

Similar to the response of fish, amphibians are also less sensitive to carbaryl than invertebrates. Two different FETAX studies showed divergent values for effects on embryos of the African clawed frog, *Xenopus laevis*. A 24-hr static study of carbaryl with unmeasured concentrations had an LC50 of 4,700 µg/L, while an exposure for the same amount of time and experimental conditions caused developmental abnormalities (localized edema and ventral bending) at 110 µg/L (Elliott-Freeley and Armstrong 1982). However, Bacchetta et al. (2008) reported substantially reduced sensitivity for the species with a LC50 of 20,280 µg/L after a 115-hr static exposure (unmeasured concentrations) of larvae. These studies were not used in quantitative criterion calculations because of insufficient duration of the tests. The SMAV calculated using acceptable LC50 values for *X. laevis* in **Appendix A** is 1,730 µg/L carbaryl, which is lower than reported LC50 values in these qualitative studies.

Acetylcholinesterase inhibition in fish is also demonstrated in the results of Zinkl et al. (1987) with rainbow trout (**Appendix H**). The authors observed decreased brain cholinesterase activity within 24 hours at carbaryl concentrations ≥ 500 µg/L. This study was not included in quantitative criterion calculations because it is an atypical endpoint; while it is reflective of the MOA, it is not directly measuring the typical endpoints of survival, growth, or reproduction.

2. Freshwater Chronic Toxicity

Acceptable chronic toxicity data are available for five freshwater species representing five different genera (two crustaceans and three fish; **Appendix C**). No data were available for coldwater fish. Similar to acute toxicity testing results, invertebrates were more sensitive to

carbaryl than fish on a chronic toxicity basis as well. Chronic toxicity values for cladocerans ranged from 2.2 to 10.6 µg/L, while fish chronic toxicity values ranged from 620.8 to 897.8 µg/L. However, paired acute and chronic toxicity data were only available for three of the freshwater species (two crustaceans and one fish), and these were the only studies used in derivation of the ACR to calculate the FCV (**Table 8**).

An additional fathead minnow study did not meet screening guidelines for inclusion in criterion calculations because of test duration, but the study showed similar ranges of toxicity and provides additional supporting evidence. Larval fathead minnows (<24-hr old) were exposed for seven days (Norberg-King 1989)(**Appendix H**). Exposure for seven days is too short a duration for this test to be used quantitatively for evaluating chronic toxicity. However, the chronic values estimated for four studies ranged from 576 to 1,088 µg/L, which is consistent with the fathead minnow chronic values of the life-cycle toxicity test (378 µg/L) and the 32-day ELS toxicity test (1,073 µg/L).

The use of NOEC/LOEC/MATC approach in criteria derivation can contribute to uncertainty. Potential sources of uncertainty are the range of test concentrations (dilution series) and the sample size used in the test. Where the design is suboptimal, higher NOEC and LOEC values may be reported due to low statistical power and high error variance. Typically, as the accuracy of the test increases, the NOEC decreases. Additional uncertainty is inherent in the calculation of the MATC which is the geometric mean of the NOEC and LOEC. The calculation compounds the fact that higher NOEC and LOEC values may be reported due to poor design. Thus some effects could occur below the calculated MATC due to the inherent uncertainty in the calculation.

Another source of uncertainty for chronic criterion calculations is the use of ACRs. When chronic data are lacking for a particular species' response to a given chemical, ACRs are determined using a formula that relates acute responses (LC50 values) to chronic concentrations using empirical relationships between acute and chronic values for taxonomically similar species for which both acute and chronic data are available. This approach has been incorporated into the 1985 "Guidelines" for evaluating chronic toxicity thresholds where the species mean chronic value (SMCV) is calculated to be the geometric mean between the NOEC and LOEC. For example, the carbaryl FACR is assumed to be 2.0 for freshwater species. The most appropriate calculated species mean ACRs are less than 2.0, and the "Guidelines" stipulate acclimation has

probably occurred during the chronic test and the FACR should be assumed to be 2.0. This assumption is a potential source of uncertainty.

3. Freshwater Field Studies

Field studies have been conducted to measure effects of aerial application of carbaryl for forest pest control upon the non-target aquatic stream and pond organisms. In northern Maine, nine streams, with different substrate, velocity and cover conditions, were studied to determine the effect of aerial application of carbaryl formulated endproduct (Sevin[®]-4-oil) on aquatic stream invertebrates applied at a rate of 0.840 kg active ingredient per hectare (0.840 kg a.i./ha). Drift of aquatic invertebrates (defined as the release, dispersal, and downstream displacement of invertebrates normally inhabiting benthic substrates) increased by 170 times two days post-treatment compared to controls, with drift samples commonly containing Plecoptera (stoneflies), Ephemeroptera (Mayflies), and Diptera (flies/mosquitoes) (Courtemanch and Gibbs 1980). In forest ponds, amphipods were severely impacted following aerial spraying of Sevin-4-oil applied at the same rate (Gibbs et al. 1984). The amphipods *Hyalella azteca* and *Crangonyx richmondensis* were completely eliminated and failed to recolonize 30 months after treatment, with maximum residue concentrations of 254 µg/L found in the water and 53,793 µg/kg (dry weight) in the sediment.

In a 77-day mesocosm study, researchers examined the effects of carbaryl on amphibians in terms of body size, length of larval period, and survival to metamorphosis when exposed to carbaryl (21.3% active ingredient) early in the larval period (Boone and Semlitsch 2002). The study units consisted of fifty 1480-L polyethylene ponds (1.85 m in diameter) containing 1,000 L of well water, 1 kg of leaf litter, and plankton from natural ponds. The study manipulated initial larval density, i.e., low (80) and high (240), pond hydroperiod, (constant or drying), and chemical concentration (absent, 3.5 mg/L, 5.0 mg/L, or 7.0 mg carbaryl /L). Frog species included: Southern leopard frog (*Rana sphenoccephala*), Plains leopard frog (*R. blairi*), green frog (*R. clamitans*), and the Woodhouse's toad (*Bufo woodhousii*). Toads in the high-density larval ponds showed greater survival than those in low-density larval ponds at the highest carbaryl level. For Southern and Plains leopard frogs, carbaryl treatment did not have a significant effect on either species. For *R. clamitans*, carbaryl exposure had a significant effect

($p < 0.05$) on days to metamorphosis with tadpoles in the chemical treatments generally having longer larval periods.

4. Estuarine/Marine Acute Toxicity

Acute toxicity data are available for 12 estuarine/marine species representing 11 genera. These data represent a dataset supporting the development of an estuarine/marine acute criterion.

SMAVs for carbaryl range from 7.188 to 17,000 $\mu\text{g/L}$. The most sensitive genus was the mysid (*Americamysis*), with a GMAV of 7.188 $\mu\text{g/L}$, followed by the Dungeness crab (*Metacarcinus*) with a GMAV of 10 $\mu\text{g/L}$. The two most tolerant genera were the bent-nosed clam (*Macoma*) and the threespine stickleback (*Gasterosteus*), with SMAVs of 17,000 and 3,990 $\mu\text{g/L}$, respectively (**Table 5** and **Figure 4**).

The four most sensitive estuarine/marine genera are *Americamysis*, *Metacarcinus*, *Callinassa* and *Upogebia* with SMAVs of 7.188, 10, 48.99, and 60.0 $\mu\text{g/L}$, respectively. Additional toxicity data on the effect of carbaryl on estuarine/marine species that were not used quantitatively are very similar to acute and chronic ranges of toxicity and provide additional supporting evidence of potential toxicity of carbaryl. Shrimp were found to be more sensitive than bivalves to carbaryl by different researchers (**Appendix I**). Various 24- to 96-hr LC50 values for different species ranged from 1.5 $\mu\text{g/L}$ (*Penaeus aztecus*) to 76 $\mu\text{g/L}$ (*Palaemonetes pugio*). The LC50 values for other test species include: *Crangon septemspinosa* (20 $\mu\text{g/L}$), *Americamysis bahia* (21 $\mu\text{g/L}$), *Penaeus duorarum* (32 $\mu\text{g/L}$), *Callinassa californiensis* (49 $\mu\text{g/L}$), and *Upogebia pugettensis* (40 $\mu\text{g/L}$ for 24-hr test and 60 $\mu\text{g/L}$ for a 48-hr test). These studies were excluded from quantitative calculation of a criterion due to test design. Similarly, sensitive organisms include testing with the prezoa Dungeness crab, *Metacarcinus magister*, where a concentration of 6 $\mu\text{g/L}$ for 24 hours prevented half of the test organisms from molting to zoea (Buchanan et al. 1970). The duration of this test was too short for quantitative use in this assessment. Another sensitive species tested was the embryo of the killifish, *Fundulus heteroclitus*, which at a carbaryl concentration of 10 $\mu\text{g/L}$ for 40 days exhibited slowed development and diminished pigmentation in fry (Crawford and Guarino 1985). This study was excluded from use in quantitative criterion calculations because of test design.

Bivalves are moderately insensitive to carbaryl, with SMAVs ranging from 1,650 $\mu\text{g/L}$ for the bay mussel (*Mytilus*) to 3,850 $\mu\text{g/L}$ for cockle clam (*Clinocardium*). Tests with the blue

mussel (*Mytilus edulis*) at various lifestages for 1 hour exposure had EC50 values for development (e.g., disjunction of blastomeres, reduced development rate and asynchronous and unaligned cleavages) that ranged from 5,300 to 24,000 µg/L (Armstrong and Milleman 1974c). This study was excluded from quantitative assessment due to less than 3 exposure concentrations and a community field exposure test. However, the test results are consistent with longer term tests (10-40 days) on shell growth that had effect concentrations of >1,300 to >2,900 µg/L (Liu and Lee 1975, **Appendix I**).

The sheepshead minnow (*Cyprinodon variegates*) and the threespine stickleback (*Gasterosteus aculeatus*) have SMAVs of 2,600 and 3,990 µg/L, respectively. Acute toxicity of carbaryl to striped bass, *Morone saxatilis* was reported by (Korn and Earnest 1974). The bioassays were 4-day tests using proportional diluters and carbaryl had a 96-hr LC50 = 1,000 µg/l. Although the LC50 for striped bass is more sensitive than the sheepshead minnow and the threespine stickleback, the study was not included in criterion calculations because control survival was not reported.

There were three studies available for estuarine/marine species that evaluated atypical endpoints and were not included in the quantitative criterion calculations. Sheepshead minnows (*Cyprinodon variegatus*) did not avoid water with 100 µg carbaryl/L after 1.5 hour of exposure (Hansen 1969; 1970). Likewise, grass shrimp (*Palaemonetes pugio*) did not avoid water with 100 µg carbaryl/L after a similar 1.5 hour exposure (Hansen et al. 1973). However, Weis and Weis (1974b) observed schooling behavior disrupted in Atlantic silversides (*Menidia menidia*) after 24 hour exposure at 100 µg carbaryl/L.

5. Estuarine/Marine Chronic Toxicity

Only one carbaryl estuarine/marine chronic toxicity test is available for qualitative consideration in this document. Survival was the most sensitive endpoint for the mysid, *A. bahia*, with a chronic value of 9.918 µg/L (Thursby and Champlin 1991b). There is uncertainty associated with this study because reported total control survival was 63% and the number of young produced by the first-generation females in the control was below the accepted ASTM requirement. However, the calculated ACR for this study is similar to the freshwater chronic invertebrate studies and provides an additional line of evidence for the sensitivity of invertebrates to carbaryl.

A ten week flow-through colonization study where exposure concentrations were measured provides qualitative information on the chronic effects of carbaryl on the development of estuarine/marine communities. This study, conducted in laboratory aquaria (Tagatz et al. 1979) showed similar tendencies as the mysid test. The amphipods were exposed to measured carbaryl concentrations of 0, 1.1, 11.1 and 103 µg/L. The number of species collected in the two highest concentrations was approximately half that from the control and lowest concentration, with the Phylum Arthropoda most obviously reduced. The number of amphipods, *Corophium acherusicum*, decreased by about 48 percent relative to the control at 1.1 µg/L, and this amphipod was totally absent at 11.1 and 103 µg/L.

6. Bioaccumulation

No acceptable data on the bioaccumulation of carbaryl in freshwater or estuarine/marine waters are available; however, because of its low octanol/water partition coefficient (229), carbaryl is not expected to bioconcentrate to a significant extent (U.S. EPA 2010a). Reported K_{ow} values range from 65 to 229 (Bracha and O'Brian 1966; Mount and Oehme 1981; Windholz et al. 1976).

The U.S. EPA Office of Water's fish tissue sampling program does not include carbaryl as an analyte, as it is not expected to bioaccumulate or bioconcentrate. No U.S. Food and Drug Administration (FDA) action level or other maximum acceptable concentration in tissue, as defined in the "Guidelines" is available for carbaryl. Therefore, a Final Residue Value cannot be calculated for fish tissue.

B. Effects on Aquatic Plants

In general, freshwater and estuarine/marine plants appear less sensitive than animals to carbaryl. The most sensitive aquatic plant tests are roughly two orders of magnitude less sensitive to carbaryl than the aquatic animal tests (**Appendix E**). For freshwater plants the effect concentrations ranged from 100 µg/L for the green alga, *Scenedesmus quadricaudata* (Lejczak 1977), to 50,000 µg/L for the blue green alga, *Anabaena torulosa* (Obulakondaiah et al. 1993). A planktonic alga mixture was adversely impacted at a concentration of 10,000 µg/L for two weeks (Butler et al. 1975), and growth of the green alga *Ankistrodesmus braunii* was reduced by

a concentration of 25,000 µg/L in a 48-hr test (Kopecek et al. 1975). Other algae were adversely impacted by much lower concentrations. The green alga, *Chlorella pyrenoidosa*, had reduced growth after a four-day exposure to 100 µg/L (Christie 1969), and the blue-green alga, *Microcystis aeruginosa*, began to show adverse effects after eight days of exposure to 1,350 µg/L carbaryl (Bringmann and Kuhn 1978a; 1978b). The only freshwater vascular plant tested is duckweed (*Lemna minor*), which had a 50% inhibition of growth at 23,900 µg/L (Brooke 1993).

Estuarine/marine plant growth effect values varied widely ranging from 100 µg/L for green algae (*Chlorella sp.*, *Dunaliella euchora* and *Protococcus sp.*), diatoms (*Phaeodactylum tricornutum*) and golden algae (*Monochrysis lutheri*) to 100,000 µg/L for the same species. The main difference between the values is that cells were viable at the lower concentrations (the effect of carbaryl was algistatic, i.e., the cells recovered after exposure was terminated), but not at the higher values where the effect was algicidal (i.e., the cells were killed as a result of the exposure) (Ukeles 1962).

V. IMPLEMENTATION

As discussed in the Water Quality Standards Regulation (U.S. EPA 1983a) and the Foreword to this document, a water quality criterion for aquatic life has regulatory impact only when it has been adopted in a state/tribal water quality standard or EPA promulgated standard for a state or tribe. Such a regulatory standard would specify a criterion for a pollutant that would be protective of a particular designated use. With the approval of the U.S. EPA, states/tribes designate one or more uses for waters in their states/tribes and adopt criteria that are protective of those use(s) they have designated (U.S. EPA 1983a;b; 1987; 1994). States/tribes may adopt water quality criteria with the same numerical values as EPA's recommended section 304 criteria. However there are situations where states and authorized tribes might want to adjust water quality criteria developed under section 304 to reflect either statewide or site-specific natural environmental conditions and/or sensitivities of local species. Alternatively, states and tribes may use different data and assumptions than the EPA in deriving numeric criteria when those data have been reviewed and deemed to be scientifically defensible and the resulting criteria value is protective of designated uses. State/tribe water quality standards include both numeric and narrative criteria. A state/tribe may adopt a numeric criterion within

its water quality standards and apply it either to all waters for the use the criterion is designed to protect or to a specific site. A state/tribe may use an indicator characteristic or the national criterion supplemented with other relevant information, to interpret its narrative criteria within its water quality standards to develop a numeric value for use in developing National Pollutant Discharge Elimination System (NPDES) effluent limitations under 40 CRF 122.44(d)(1)(vi).2 (<http://cfr.vlex.com/vid/122-44-establishing-applicable-mpdes-see-123-19811557>).

Site-specific criteria may include not only site-specific criterion concentrations (U.S. EPA 1994), but also site-specific, and possibly pollutant-specific, durations of averaging periods and frequencies of allowed excursions (U.S. EPA 1991). The averaging periods of "one hour" and "four days" were selected by the U.S. EPA on the basis of data concerning how rapidly some aquatic species react to increases in the concentrations of some aquatic pollutants, and "three years" return frequency is the Agency's scientific judgment regarding the average amount of time aquatic ecosystems should be provided to recover between excursions (Stephan et al. 1985; U.S. EPA 1991). However, various species and ecosystems react and recover at greatly differing rates. Therefore, if adequate justification is provided, site-specific and/or pollutant-specific concentrations, durations, and frequencies may be higher or lower than those given in national recommended water quality criteria for aquatic life.

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Appendix A

Appendix A. Acceptable Acute Toxicity Data of Carbaryl to Freshwater Aquatic Animals

Species	Method ^a	Chemical	Hardness (mg/L as CaCO ₃)	LC50 or EC50 (µg/L)	Species Mean Acute Value (µg/L)	Reference
Freshwater Species						
Oligochaete worm, <i>Lumbriculus variegatus</i>	S, U	Analytical	30	8,200	8,200	Bailey and Liu 1980
Snail (adult), <i>Aplexa hypnorum</i>	F, M	-	44.4	>27,000	>27,000	Phipps and Holcombe 1985
Mussel (juvenile; 1-2 d), <i>Anodonta imbecillis</i>	R, U	99%	40-50	23,700	-	Johnson et al. 1993
Mussel (juvenile; 7-10 d), <i>Anodonta imbecillis</i>	R, U	99%	40-50	25,600	24,632	Johnson et al. 1993
Cladoceran (<24 hr), <i>Ceriodaphnia dubia</i>	R, M	98%	169	3.06	-	Brooke 1990; 1991
Cladoceran (<12 hr), <i>Ceriodaphnia dubia</i>	R, M	>99%	57	11.6	5.958	Oris et al. 1991
Cladoceran (adult; 2-2.5 mm), <i>Daphnia carinata</i>	S, U	Technical	-	35	35	Santharam et al. 1976
Cladoceran (5 d), <i>Daphnia magna</i>	S, U	97.6%	-	7.2 ^b	-	Lakota et al. 1981
Cladoceran (<24 hr), <i>Daphnia magna</i>	R, M	99%	40-50	1,900 ^b (LC, not EC value)	-	Johnson et al. 1993
Cladoceran (<24 hr), <i>Daphnia magna</i>	S, U	99.5%	40	5.6	-	Sanders et al. 1983
Cladoceran (<24 hr), <i>Daphnia magna</i>	R, M	98%	181.8	10.1	7.521	Brooke 1991
Cladoceran (<24 hr), <i>Daphnia pulex</i>	S, U	-	40-48	6.4	6.4	Sanders and Cope 1966
Cladoceran (<24 hr), <i>Simocephalus serrulatus</i>	S, U	99.5%	44	11 (10°C)	-	Mayer and Ellersieck 1986
Cladoceran (<24 hr), <i>Simocephalus serrulatus</i>	S, U	-	44	7.6 (16°C)	-	Sanders and Cope 1966

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Cladoceran (<24 hr), <i>Simocephalus serrulatus</i>	S, U	99.5%	44	8.1 (21°C)	8.781	Mayer and Ellersieck 1986
Mysid, <i>Mysis relicta</i>	R, M	98%	139.3	230	230	Landrum and Dupuis 1990
Aquatic sowbug (mature), <i>Asellus brevicaudus</i>	S, U	99.5%	44	280	280	Johnson and Finley 1980; Mayer and Ellersieck 1986
Amphipod (2 mo), <i>Gammarus lacustris</i>	S, U	100%	30.5	16	-	Sanders 1969
Amphipod (mature), <i>Gammarus lacustris</i>	S, U	99.5%	44	22	18.76	Johnson and Finley 1980; Mayer and Ellersieck 1986
Amphipod, <i>Gammarus pseudolimnaeus</i>	S, U	99%	40	13 (pH=6.5)	-	Woodward and Mauck 1980
Amphipod (mature), <i>Gammarus pseudolimnaeus</i>	S, U	99%	40	7 (pH=7.5)	-	Woodward and Mauck 1980
Amphipod (mature), <i>Gammarus pseudolimnaeus</i>	S, U	99%	40	7.2 (pH=8.5)	-	Woodward and Mauck 1980
Amphipod (mature), <i>Gammarus pseudolimnaeus</i>	S, U	99.5%	40	16	10.12	Sanders et al. 1983
Amphipod (14 d), <i>Hyaella azteca</i>	S, U	Technical	280	15.2	15.2	McNulty et al. 1999
Amphipod, <i>Pontoporeia hoyi</i>	R, M	98%	139.3	250	250	Landrum and Dupuis 1990
Crayfish (3-4 cm), <i>Cambarus bartoni</i>	R, U	99.8%	-	839.6	839.6	Simon 1982
Crayfish (3.9 g), <i>Orconectes immunis</i>	F, M	-	44.4	2,870	2,870	Phipps and Holcombe 1985
Crayfish (5-8 cm; males), <i>Orconectes virilis</i>	R, U	99.8%	-	2,112	2,112	Simon 1982

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Crayfish (15-38 g), <i>Procambarus clarkii</i>	S, U	-	250	1,000	1,000	Andreu-Moliner et al. 1986
Stonefly (nymph), <i>Claassenia sabulosa</i>	S, U	Technical	44	5.6	5.6	Sanders and Cope 1968
Stonefly (1st yr class), <i>Isogenus sp.</i>	S, U	99.5%	35	2.8	-	Mayer and Ellersieck 1986
Stonefly (1st yr class), <i>Isogenus sp.</i>	S, U	99.5%	42	3.6	3.175	Mayer and Ellersieck 1986
Stonefly (1st yr class; 15-20 mm), <i>Pteronarcella badia</i>	S, U	Technical	44	1.7	-	Sanders and Cope 1968
Stonefly (1st yr class), <i>Pteronarcella badia</i>	S, U	99%	38	11 (pH=6.5)	-	Woodward and Mauck 1980; Mayer and Ellersieck 1986
Stonefly (1st yr class), <i>Pteronarcella badia</i>	S, U	99%	38	13 (pH=7.5)	-	Woodward and Mauck 1980; Mayer and Ellersieck 1986
Stonefly (1st yr class), <i>Pteronarcella badia</i>	S, U	99%	38	29 (pH=8.5)	9.163	Woodward and Mauck 1980; Mayer and Ellersieck 1986
Stonefly (1st yr class), <i>Pteronarcys californica</i>	S, U	Technical	44	4.8	4.8	Sanders and Cope 1968
Stonefly (naiad), <i>Skwala sp.</i>	S, U	99.5%	-	3.6	3.6	Johnson and Finley 1980
Backswimmer (adult), <i>Notonecta undulata</i>	S, U	94%	-	200	200	Federle and Collins 1976
Apache trout (0.38-0.85 g), <i>Oncorhynchus apache</i>	S, U	99.7%	169	1,540	1,540	Dwyer et al. 1995
Coho salmon (2.7-4.1 g), <i>Oncorhynchus kisutch</i>	S, U	95%	-	997	-	Katz 1961

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Coho salmon, <i>Oncorhynchus kisutch</i>	S, U	99%	40-48	764	-	Macek and McAllister 1970
Coho salmon (1.50 g), <i>Oncorhynchus kisutch</i>	S, U	98%	318-348	1,300	-	Post and Schroeder 1971
Coho salmon (1.0 g), <i>Oncorhynchus kisutch</i>	S, U	99.5%	44	4,340	-	Johnson and Finley 1980; Mayer and Ellersieck 1986
Coho salmon (4.6 g), <i>Oncorhynchus kisutch</i>	S, U	99.5%	42	2,400	-	Mayer and Ellersieck 1986
Coho salmon (5.1 g), <i>Oncorhynchus kisutch</i>	S, U	99.5%	42	1,750	-	Mayer and Ellersieck 1986
Coho salmon (10.10 g), <i>Oncorhynchus kisutch</i>	S, U	99.5%	42	2,700	-	Mayer and Ellersieck 1986
Coho salmon (19.1 g), <i>Oncorhynchus kisutch</i>	S, U	99.5%	42	1,150	1,654	Mayer and Ellersieck 1986
Chinook salmon (fingerling), <i>Oncorhynchus tshawytscha</i>	F, U	99.5%	314	2,400 ^c	-	Johnson and Finley 1980; Mayer and Ellersieck 1986
Chinook salmon (3.0 g), <i>Oncorhynchus tshawytscha</i>	F, M	-	44.4	2,690	2,690	Phipps and Holcombe 1985; 1990
Cutthroat trout (0.37 g), <i>Oncorhynchus clarkii</i>	S, U	98%	318-348	1,500	-	Post and Schroeder 1971
Cutthroat trout (1.30 g), <i>Oncorhynchus clarkii</i>	S, U	98%	318-348	2,169	-	Post and Schroeder 1971
Cutthroat trout (0.5 g), <i>Oncorhynchus clarkii</i>	S, U	99.5%	40	7,100	-	Johnson and Finley 1980; Mayer and Ellersieck 1986
Cutthroat trout (0.6 g), <i>Oncorhynchus clarkii</i>	S, U	99%	40	6,000 (pH=7.5) (7°C)	-	Woodward and Mauck 1980; Mayer and Ellersieck 1986
Cutthroat trout (0.7 g), <i>Oncorhynchus clarkii</i>	S, U	99%	40	5,000 (pH=6.5) (12°C)	-	Woodward and Mauck 1980; Mayer and Ellersieck 1986
Cutthroat trout (0.6 g), <i>Oncorhynchus clarkii</i>	S, U	99%	40	970 (pH=8.5) (12°C)	-	Woodward and Mauck 1980; Mayer and Ellersieck 1986
Cutthroat trout (0.5 g), <i>Oncorhynchus clarkii</i>	S, U	99%	320	3,950 (pH=7.8) (12°C)	-	Woodward and Mauck 1980; Mayer and Ellersieck 1986
Cutthroat trout (0.5 g), <i>Oncorhynchus clarkii</i>	S, U	99.5%	40	6,800 (pH=7.3)	-	Mayer and Ellersieck 1986

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Cutthroat trout (0.9 g), <i>Oncorhynchus clarkii</i>	S, U	99.5%	42	6,700 (pH=7.5)	-	Mayer and Ellersieck 1986
Cutthroat trout, <i>Oncorhynchus clarkii</i>	S, U	99%	40	3,950 (pH=7.5) (12°C)	-	Woodward and Mauck 1980
Greenback cutthroat trout (0.31 g), <i>Oncorhynchus clarkii stomias</i>	S, U	99.7%	169	1,550	-	Dwyer et al. 1995
Lahontan cutthroat trout (0.34-0.57 g), <i>Oncorhynchus clarkii henshawi</i>	S, U	99.7%	169	2,250	3,300	Dwyer et al. 1995
Rainbow trout (3.2 g), <i>Oncorhynchus mykiss</i>	S, U	95%	-	1,350	-	Katz 1961
Rainbow trout, <i>Oncorhynchus mykiss</i>	S, U	99%	40-48	4,340	-	Macek and McAllister 1970
Rainbow trout (1.24 g), <i>Oncorhynchus mykiss</i>	S, U	98%	318-348	1,470	-	Post and Schroeder 1971
Rainbow trout (1.5 g), <i>Oncorhynchus mykiss</i>	S, U	99.5%	42	1,950	-	Johnson and Finley 1980; Mayer and Ellersieck 1986;
Rainbow trout, <i>Oncorhynchus mykiss</i>	S, U	99.5%	40	2,200 (12°C)	-	Sanders et al. 1983
Rainbow trout, <i>Oncorhynchus mykiss</i>	S, U	99.5%	40	2,800 (7°C)	-	Sanders et al. 1983
Rainbow trout, <i>Oncorhynchus mykiss</i>	S, U	99.5%	40	1,100 (pH=6.5)	-	Sanders et al. 1983
Rainbow trout, <i>Oncorhynchus mykiss</i>	S, U	99.5%	40	800 (pH=7.5)	-	Sanders et al. 1983
Rainbow trout, <i>Oncorhynchus mykiss</i>	S, U	99.5%	40	1,500 (pH=8.5)	-	Sanders et al. 1983
Rainbow trout, <i>Oncorhynchus mykiss</i>	S, U	99.5%	40	900	-	Sanders et al. 1983
Rainbow trout, <i>Oncorhynchus mykiss</i>	S, U	99.5%	320	800	-	Sanders et al. 1983
Rainbow trout (1.0 g), <i>Oncorhynchus mykiss</i>	S, U	-	-	935	-	Marking et al. 1984
Rainbow trout (1.0 g), <i>Oncorhynchus mykiss</i>	S, U	-	-	1,000	-	Marking et al. 1984
Rainbow trout (1.0 g), <i>Oncorhynchus mykiss</i>	S, U	-	-	1,400	-	Marking et al. 1984
Rainbow trout (1.0 g), <i>Oncorhynchus mykiss</i>	S, U	-	-	1,000	-	Marking et al. 1984

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Rainbow trout (1.0 g), <i>Oncorhynchus mykiss</i>	S, U	-	-	1,740	-	Marking et al. 1984
Rainbow trout (juvenile), <i>Oncorhynchus mykiss</i>	R, U	99%	-	4,835	-	Douglas et al. 1986
Rainbow trout (1.5 g), <i>Oncorhynchus mykiss</i>	S, U	99.5%	272	1,200	-	Mayer and Ellersieck 1986
Rainbow trout (0.8 g), <i>Oncorhynchus mykiss</i>	S, U	99.5%	40	1,360	-	Mayer and Ellersieck 1986
Rainbow trout (0.8 g), <i>Oncorhynchus mykiss</i>	S, U	99.5%	40	2,080	-	Mayer and Ellersieck 1986
Rainbow trout (1.1 g), <i>Oncorhynchus mykiss</i>	S, U	99.5%	40	1,900	-	Mayer and Ellersieck 1986
Rainbow trout (1.1 g), <i>Oncorhynchus mykiss</i>	S, U	99.5%	40	2,300	-	Mayer and Ellersieck 1986
Rainbow trout (0.5 g), <i>Oncorhynchus mykiss</i>	S, U	99.5%	314	1,330	-	Mayer and Ellersieck 1986
Rainbow trout (0.8 g), <i>Oncorhynchus mykiss</i>	S, U	99.5%	40	<750	-	Mayer and Ellersieck 1986
Rainbow trout (1.1 g), <i>Oncorhynchus mykiss</i>	S, U	99.5%	40	<320	-	Mayer and Ellersieck 1986
Rainbow trout (1.2 g), <i>Oncorhynchus mykiss</i>	S, U	99.5%	40	1,090	-	Mayer and Ellersieck 1986
Rainbow trout (1.1 g), <i>Oncorhynchus mykiss</i>	S, U	99.5%	40	1,460	-	Mayer and Ellersieck 1986
Rainbow trout (1.2 g), <i>Oncorhynchus mykiss</i>	S, U	99.5%	40	3,500	-	Mayer and Ellersieck 1986
Rainbow trout (1.2 g), <i>Oncorhynchus mykiss</i>	S, U	99.5%	320	3,000	-	Mayer and Ellersieck 1986
Rainbow trout (1.0 g), <i>Oncorhynchus mykiss</i>	S, U	99.5%	40	1,600	-	Mayer and Ellersieck 1986
Rainbow trout (1.0 g), <i>Oncorhynchus mykiss</i>	S, U	99.5%	40	1,100	-	Mayer and Ellersieck 1986
Rainbow trout (1.0 g), <i>Oncorhynchus mykiss</i>	S, U	99.5%	40	1,200	-	Mayer and Ellersieck 1986
Rainbow trout (1.0 g), <i>Oncorhynchus mykiss</i>	S, U	99.5%	40	780	-	Mayer and Ellersieck 1986
Rainbow trout (1.0 g), <i>Oncorhynchus mykiss</i>	S, U	99.5%	40	1,450	-	Mayer and Ellersieck 1986
Rainbow trout (0.48-1.25 g), <i>Oncorhynchus mykiss</i>	S, U	99.7%	169	1,880	-	Dwyer et al. 1995

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Rainbow trout (juvenile; 2.7 g), <i>Oncorhynchus mykiss</i>	S, M	99%	-	5,400 ^c	-	Ferrari et al. 2004
Rainbow trout (19.7 g), <i>Oncorhynchus mykiss</i>	F, M	Technical	44.4	860	860	Phipps and Holcombe 1985
Atlantic salmon (0.4 g), <i>Salmo salar</i>	S, U	99.5%	42	4,500	-	Mayer and Ellersieck 1986
Atlantic salmon (0.8 g), <i>Salmo salar</i>	S, U	99.5%	42	2,070	-	Mayer and Ellersieck 1986
Atlantic salmon (0.8 g), <i>Salmo salar</i>	S, U	99.5%	42	1,180	-	Mayer and Ellersieck 1986
Atlantic salmon (0.4 g), <i>Salmo salar</i>	S, U	99.5%	42	905	-	Mayer and Ellersieck 1986
Atlantic salmon (0.8 g), <i>Salmo salar</i>	S, U	99.5%	12	2,010	-	Mayer and Ellersieck 1986
Atlantic salmon (0.8 g), <i>Salmo salar</i>	S, U	99.5%	42	1,430	-	Mayer and Ellersieck 1986
Atlantic salmon (0.2 g), <i>Salmo salar</i>	S, U	99.5%	42	500	-	Mayer and Ellersieck 1986
Atlantic salmon (0.2 g), <i>Salmo salar</i>	S, U	99.5%	42	1,000	-	Mayer and Ellersieck 1986
Atlantic salmon (0.2 g), <i>Salmo salar</i>	S, U	99.5%	42	1,150	-	Mayer and Ellersieck 1986
Atlantic salmon (0.2 g), <i>Salmo salar</i>	S, U	99.5%	42	1,100	-	Mayer and Ellersieck 1986
Atlantic salmon (0.2 g), <i>Salmo salar</i>	S, U	99.5%	42	1,350	-	Mayer and Ellersieck 1986
Atlantic salmon (0.2 g), <i>Salmo salar</i>	S, U	99.5%	42	220	-	Mayer and Ellersieck 1986
Atlantic salmon (0.2 g), <i>Salmo salar</i>	S, U	99.5%	42	900	-	Mayer and Ellersieck 1986
Atlantic salmon (0.2 g), <i>Salmo salar</i>	S, U	99.5%	42	1,000	1,129	Mayer and Ellersieck 1986
Brown trout, <i>Salmo trutta</i>	S, U	99%	40-48	1,950 ^b	-	Macek and McAllister 1970
Brown trout (0.6 g), <i>Salmo trutta</i>	S, U	99.5%	42	6,300 ^b	-	Johnson and Finley 1980; Mayer and Ellersieck 1986
Brown trout (fingerling), <i>Salmo trutta</i>	F, U	99.5%	314	2,000 ^b	-	Mayer and Ellersieck 1986

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Brown trout (fry), <i>Salmo trutta</i>	S, U	97.6%	-	700	700	Lakota et al. 1981
Brook trout (1.15 g), <i>Salvelinus fontinalis</i>	S, U	98%	318-348	1,070	-	Post and Schroeder 1971
Brook trout (2.04 g), <i>Salvelinus fontinalis</i>	S, U	98%	318-348	1,450	-	Post and Schroeder 1971
Brook trout (1.0 g), <i>Salvelinus fontinalis</i>	S, U	99.5%	42	680	-	Mayer and Ellersieck 1986
Brook trout (0.7 g), <i>Salvelinus fontinalis</i>	S, U	99.5%	42	4,560	-	Mayer and Ellersieck 1986
Brook trout (0.7 g), <i>Salvelinus fontinalis</i>	S, U	99.5%	42	2,130	-	Mayer and Ellersieck 1986
Brook trout (0.7 g), <i>Salvelinus fontinalis</i>	S, U	99.5%	42	1,130	-	Mayer and Ellersieck 1986
Brook trout (0.8 g), <i>Salvelinus fontinalis</i>	S, U	99.5%	42	1,200	-	Mayer and Ellersieck 1986
Brook trout (0.8 g), <i>Salvelinus fontinalis</i>	S, U	99.5%	300	1,290	-	Mayer and Ellersieck 1986
Brook trout (1.3 g), <i>Salvelinus fontinalis</i>	S, U	99.5%	42	4,500	1,629	Mayer and Ellersieck 1986
Lake trout (1.7 g), <i>Salvelinus namaycush</i>	S, U	99.5%	40	690	-	Johnson and Finley 1980; Mayer and Ellersieck 1986
Lake trout (1.7 g), <i>Salvelinus namaycush</i>	S, U	99.5%	40	740	-	Mayer and Ellersieck 1986
Lake trout (1.7 g), <i>Salvelinus namaycush</i>	S, U	99.5%	40	920	-	Mayer and Ellersieck 1986
Lake trout (0.5 g), <i>Salvelinus namaycush</i>	S, U	99.5%	162	872	-	Mayer and Ellersieck 1986
Lake trout (2.6 g), <i>Salvelinus namaycush</i>	F, U	99.5%	260	2,300	988.1	Mayer and Ellersieck 1986
Goldfish (0.9 g), <i>Carassius auratus</i>	S, U	99%	40-48	13,200 ^c	-	Macek and McAllister 1970
Goldfish (0.9 g), <i>Carassius auratus</i>	S, U	99.5%	272	12,800 ^c	-	Mayer and Ellersieck 1986
Goldfish (juvenile; 1.3-3.3 g), <i>Carassius auratus</i>	S, U	99.7%	-	17,500 ^c	-	Pfeiffer et al. 1997
Goldfish (14.2 g), <i>Carassius auratus</i>	F, M	-	44.4	16,700	16,700	Phipps and Holcombe 1985

Appendix A. Acceptable Acute Toxicity Data of Carbaryl to Freshwater Aquatic Animals

Species	Method ^a	Chemical	Hardness (mg/L as CaCO ₃)	LC50 or EC50 (µg/L)	Species Mean Acute Value (µg/L)	Reference
Common carp (0.6 g), <i>Cyprinus carpio</i>	S, U	99%	40-48	5,280	-	Macek and McAllister 1970
Common carp (0.38 g), <i>Cyprinus carpio</i>	S, U	94%	22	1,700	-	Chin and Sudderuddin 1979
Common carp (fry), <i>Cyprinus carpio</i>	S, U	97.6%	-	4,220	-	Lakota et al. 1981
Common carp (20-34 mm), <i>Cyprinus carpio</i>	S, U	85%	-	7,850	4,153	de Mel and Pathiratne 2005
European chub (12.43 cm; 18.14 g) <i>Leuciscus cephalus</i>	S,U	85%	61-65	8,656	8,656	Verep 2006
Fathead minnow (0.5 g), <i>Pimephales promelas</i>	S, U	99.5%	42	14,000 ^c	-	Mayer and Ellersieck 1986
Fathead minnow (0.8 g), <i>Pimephales promelas</i>	S, U	99%	40-48	14,600 ^c	-	Macek and McAllister 1970; Sanders et al. 1983
Fathead minnow (0.8 g), <i>Pimephales promelas</i>	S, U	99.5%	272	7,700 ^c	-	Mayer and Ellersieck 1986
Fathead minnow (larvae), <i>Pimephales promelas</i>	R, U	99%	44-49	>1,600 ^c	-	Norberg-King 1989
Fathead minnow (0.32-0.56 g), <i>Pimephales promelas</i>	S, U	99.7%	173	5,210 ^c	-	Dwyer et al. 1995
Fathead minnow (2 mo), <i>Pimephales promelas</i>	F, M	80%	41-49	9,000	-	Carlson 1971
Fathead minnow (0.3 g), <i>Pimephales promelas</i>	F, M	Technical	44.4	5,010	-	Phipps and Holcombe 1985
Fathead minnow (31 d), <i>Pimephales promelas</i>	F, M	99%	43.8	9,470	-	Geiger et al. 1985; 1988
Fathead minnow (28 d), <i>Pimephales promelas</i>	F, M	99%	45.4	8,930	-	Geiger et al. 1985; 1988
Fathead minnow (28 d), <i>Pimephales promelas</i>	F, M	99%	44.1	10,400	-	Geiger et al. 1985; 1988
Fathead minnow (29 d), <i>Pimephales promelas</i>	F, M	99%	45.4	6,670	8,012	Geiger et al. 1985; 1988
Bonytail chub (0.29-0.52 g), <i>Gila elegans</i>	S, U	99.7%	173	3,490	-	Dwyer et al. 1995

Appendix A. Acceptable Acute Toxicity Data of Carbaryl to Freshwater Aquatic Animals

Species	Method ^a	Chemical	Hardness (mg/L as CaCO ₃)	LC50 or EC50 (µg/L)	Species Mean Acute Value (µg/L)	Reference
Bonytail chub (6 d), <i>Gila elegans</i>	R, M	99%	212-216	2,020	2,655	Beyers et al. 1994
Colorado pikeminnow (formerly squawfish) (0.32-0.34 g), <i>Ptychocheilus lucius</i>	S, U	99.7%	173	3,070	-	Dwyer et al. 1995
Colorado pikeminnow (26 d), <i>Ptychocheilus lucius</i>	R, M	99%	212-216	1,310	2,005	Beyers et al. 1994
Razorback sucker (0.31-0.32 g), <i>Xyrauchen texanus</i>	S, U	99.7%	173	4,350	4,350	Dwyer et al. 1995
Black bullhead (1.2 g), <i>Ameiurus melas</i>	S, U	99%	40-48	20,000	20,000	Macek and McAllister 1970
Channel catfish (1.5 g), <i>Ictalurus punctatus</i>	S, U	99%	40-48	15,800 ^c	-	Macek and McAllister 1970
Channel catfish (0.3 g), <i>Ictalurus punctatus</i>	S, U	100%	10	1,300 ^c	-	Brown et al. 1979
Channel catfish (1.5 g), <i>Ictalurus punctatus</i>	S, U	99.5%	272	7,790 ^c	-	Mayer and Ellersieck 1986
Channel catfish (fingerling), <i>Ictalurus punctatus</i>	F, U	99.5%	314	17,300 ^c	-	Mayer and Ellersieck 1986
Channel catfish (27.6 g), <i>Ictalurus punctatus</i>	F, M	-	44.4	12,400	12,400	Phipps and Holcombe 1985
Walking catfish (17-18 cm; 60-70 g), <i>Clarias batrachus</i>	R, U	Technical	-	46,850	-	Tripathi and Shukla 1988
Walking catfish (14 cm; 25 g), <i>Clarias batrachus</i>	S, U	99%	-	16,270	27,609	Lata et al. 2001
Guppy (2.0 cm), <i>Poecilia reticulata</i>	R, M	99%	-	2,515	2,515	Gallo et al. 1995
Gila topminnow (219 mg), <i>Poeciliopsis occidentalis</i>	S, U	99.7%	167	>3,000	>3,000	Dwyer et al. 1999b

Appendix A. Acceptable Acute Toxicity Data of Carbaryl to Freshwater Aquatic Animals

Species	Method ^a	Chemical	Hardness (mg/L as CaCO ₃)	LC50 or EC50 (µg/L)	Species Mean Acute Value (µg/L)	Reference
Striped bass (56 d), <i>Morone saxatilis</i>	S, U	-	40	760	-	Palawski et al. 1985
Striped bass, <i>Morone saxatilis</i>	S, U	-	45.5	2,300	1,322	Palawski et al. 1985
Green sunfish (1.1 g), <i>Lepomis cyanellus</i>	S, U	99.5%	272	9,460	9,460	Mayer and Ellersieck 1986
Redear sunfish (1.1 g), <i>Lepomis microlophus</i>	S, U	99%	40-48	11,200	11,200	Macek and McAllister 1970
Bluegill, <i>Lepomis macrochirus</i>	S, U	99.9%	-	14,000 ^c	-	McCann and Young 1969
Bluegill (1.2 g), <i>Lepomis macrochirus</i>	S, U	99%	40-48	6,760 ^c	-	Macek and McAllister 1970
Bluegill, <i>Lepomis macrochirus</i>	S, U	99.5%	40	16,000 ^c (12°C)	-	Sanders et al. 1983
Bluegill, <i>Lepomis macrochirus</i>	S, U	99.5%	40	8,200 ^c (22°C)	-	Sanders et al. 1983
Bluegill, <i>Lepomis macrochirus</i>	S, U	99.5%	40	5,400 ^c (pH=6.5)	-	Sanders et al. 1983
Bluegill, <i>Lepomis macrochirus</i>	S, U	99.5%	40	5,200 ^c (pH=7.5)	-	Sanders et al. 1983
Bluegill, <i>Lepomis macrochirus</i>	S, U	99.5%	40	1,800 ^c (pH=8.5)	-	Sanders et al. 1983
Bluegill, <i>Lepomis macrochirus</i>	S, U	99.5%	40	2,200 ^c	-	Sanders et al. 1983
Bluegill, <i>Lepomis macrochirus</i>	S, U	99.5%	320	1,000 ^c	-	Sanders et al. 1983
Bluegill (1.2 g), <i>Lepomis macrochirus</i>	S, U	99.5%	272	5,230 ^c	-	Mayer and Ellersieck 1986
Bluegill (0.6 g), <i>Lepomis macrochirus</i>	F, U	99.5%	314	5,047 ^c	-	Mayer and Ellersieck 1986
Bluegill (0.4 g), <i>Lepomis macrochirus</i>	S, U	99.5%	44	7,400 ^c	-	Mayer and Ellersieck 1986
Bluegill (0.4 g), <i>Lepomis macrochirus</i>	S, U	99.5%	44	5,200 ^c	-	Mayer and Ellersieck 1986
Bluegill (0.8 g), <i>Lepomis macrochirus</i>	S, U	99.5%	40	16,000 ^c	-	Mayer and Ellersieck 1986
Bluegill (0.8 g), <i>Lepomis macrochirus</i>	S, U	99.5%	40	7,000 ^c	-	Sanders et al. 1983; Mayer and Ellersieck 1986
Bluegill (0.8 g), <i>Lepomis macrochirus</i>	S, U	99.5%	40	8,200 ^c	-	Mayer and Ellersieck 1986

Appendix A. Acceptable Acute Toxicity Data of Carbaryl to Freshwater Aquatic Animals

Species	Method ^a	Chemical	Hardness (mg/L as CaCO ₃)	LC50 or EC50 (µg/L)	Species Mean Acute Value (µg/L)	Reference
Bluegill (0.4 g), <i>Lepomis macrochirus</i>	S, U	99.5%	320	6,200 ^c	-	Mayer and Ellersieck 1986
Bluegill (0.7 g), <i>Lepomis macrochirus</i>	S, U	99.5%	40	5,400 ^c	-	Mayer and Ellersieck 1986
Bluegill (0.7 g), <i>Lepomis macrochirus</i>	S, U	99.5%	40	5,200 ^c	-	Mayer and Ellersieck 1986
Bluegill (0.7 g), <i>Lepomis macrochirus</i>	S, U	99.5%	40	1,800 ^c	-	Mayer and Ellersieck 1986
Bluegill (0.7 g), <i>Lepomis macrochirus</i>	S, U	99.5%	40	2,600 ^c	-	Mayer and Ellersieck 1986
Bluegill (0.5 g), <i>Lepomis macrochirus</i>	F, M	-	44.4	6,970	6,970	Phipps and Holcombe 1985
Largemouth bass (0.9 g), <i>Micropterus salmoides</i>	S, U	99%	40-48	6,400	6,400	Macek and McAllister 1970
Black crappie (1.0 g), <i>Pomoxis nigromaculatus</i>	S, U	99.5%	40	2,600	2,600	Johnson and Finley 1980; Mayer and Ellersieck 1986;
Greenthroat darter (133 mg), <i>Etheostoma lepidum</i>	S, U	99.7%	167	2,140	2,140	Dwyer et al. 1999b
Fountain darter (62 mg), <i>Etheostoma fonticola</i>	S, U	99.7%	167	2,020	2,020	Dwyer et al. 2005
Yellow perch (1.4 g), <i>Perca flavescens</i>	S, U	99%	40-48	745	-	Macek and McAllister 1970
Yellow perch (0.6 g), <i>Perca flavescens</i>	S, U	99.5%	42	5,100	-	Johnson and Finley 1980; Mayer and Ellersieck 1986
Yellow perch (1.0 g), <i>Perca flavescens</i>	S, U	99.5%	42	13,900	-	Mayer and Ellersieck 1986
Yellow perch (1.0 g), <i>Perca flavescens</i>	S, U	99.5%	42	5,400	-	Mayer and Ellersieck 1986
Yellow perch (1.0 g), <i>Perca flavescens</i>	S, U	99.5%	42	3,400	-	Mayer and Ellersieck 1986
Yellow perch (1.0 g), <i>Perca flavescens</i>	S, U	99.5%	42	1,200	-	Mayer and Ellersieck 1986
Yellow perch (0.9 g), <i>Perca flavescens</i>	S, U	99.5%	42	4,000	-	Mayer and Ellersieck 1986

Appendix A. Acceptable Acute Toxicity Data of Carbaryl to Freshwater Aquatic Animals

Species	Method ^a	Chemical	Hardness (mg/L as CaCO ₃)	LC50 or EC50 (µg/L)	Species Mean Acute Value (µg/L)	Reference
Yellow perch (0.9 g), <i>Perca flavescens</i>	S, U	99.5%	42	4,200	-	Mayer and Ellersieck 1986
Yellow perch (0.9 g), <i>Perca flavescens</i>	S, U	99.5%	42	480	-	Mayer and Ellersieck 1986
Yellow perch (0.9 g), <i>Perca flavescens</i>	S, U	99.5%	42	350	-	Mayer and Ellersieck 1986
Yellow perch (1.0 g), <i>Perca flavescens</i>	S, U	99.5%	42	3,800	-	Mayer and Ellersieck 1986
Yellow perch (1.0 g), <i>Perca flavescens</i>	S, U	99.5%	170	5,000	-	Mayer and Ellersieck 1986
Yellow perch (1.0 g), <i>Perca flavescens</i>	S, U	99.5%	300	3,750	-	Mayer and Ellersieck 1986
Yellow perch (fingerling), <i>Perca flavescens</i>	F, U	99.5%	314	1,420	2,480	Mayer and Ellersieck 1986
Shortnosed sturgeon, <i>Acipenser brevirostrum</i>	S, U	99.7%	170	1,810	1,810	Dwyer et al. 2000
Nile tilapia (45-55 mm; 3.17 g), <i>Oreochromis niloticus</i>	S, U	85%	-	2,930	2,930	dela Cruz and Cagauan 1981
Green frog (Gosner stage 25 tadpole), <i>Rana clamitans</i>	S, U	99.7%	286	22,020 (17°C)	-	Boone and Bridges 1999
Green frog (Gosner stage 25 tadpole), <i>Rana clamitans</i>	S, U	99.7%	286	17,360 (22°C)	-	Boone and Bridges 1999
Green frog (Gosner stage 25 tadpole), <i>Rana clamitans</i>	S, U	99.7%	286	11,320 (27°C)	16,296	Boone and Bridges 1999
Boreal toad (200 mg), <i>Bufo boreas</i>	S, U	99.7%	286	12,310	12,310	Dwyer et al. 1999b
Gray tree frog (tadpole), <i>Hyla versicolor</i>	S, U	99.7%	-	2,470	2,470	Zaga et al. 1998
African clawed frog (embryo), <i>Xenopus laevis</i>	S, U	99.7%	-	15,250 ^b	-	Zaga et al. 1998

Appendix A. Acceptable Acute Toxicity Data of Carbaryl to Freshwater Aquatic Animals

Species	Method ^a	Chemical	Hardness (mg/L as CaCO ₃)	LC50 or EC50 (µg/L)	Species Mean Acute Value (µg/L)	Reference
African clawed frog (tadpole), <i>Xenopus laevis</i>	S, U	99.7%	-	1,730	1,730	Zaga et al. 1998

^a S = static; R = renewal; F = flow-through; M = measured; U = unmeasured.

^b Data not used to calculate SMAV because a more sensitive life stage or endpoint, or definitive value available for the species.

^c Data not used to calculate SMAV because a more sensitive test exposure available for the species.

Dash indicates not available

Appendix B

Appendix B. Acceptable Acute Toxicity Data of Carbaryl to Estuarine/Marine Aquatic Animals

Species	Method ^a	Chemical	Salinity (g/kg)	LC50 or EC50 (µg/L)	Species Mean Acute Value (µg/L)	References
Estuarine/marine Species						
Bay mussel (adult), <i>Mytilus edulis</i>	S, M	99.7%	25	22,700 ^b	-	Liu and Lee 1975
Bay mussel (embryo/larva), <i>Mytilus edulis</i>	S, M	99.7%	25.9	1,480 (22°C)	-	Liu and Lee 1975
Bay mussel (embryo/larva), <i>Mytilus edulis</i>	S, M	99.7%	25.9	1,800 (22°C)	-	Liu and Lee 1975
Bay mussel (embryo/larva), <i>Mytilus edulis</i>	S, M	99.7%	25.9	1,210 (22°C)	-	Liu and Lee 1975
Bay mussel (larva), <i>Mytilus edulis</i>	S, U	80%	25	2,300	1,650	Stewart et al. 1967
Pacific oyster (larva), <i>Crassostrea gigas</i>	S, U	80%	25	2,200	2,200	Stewart et al. 1967
Eastern oyster (embryo/larva), <i>Crassostrea virginica</i>	S, U	-	-	3,000 (24°C)	-	Davis and Hidu 1969
Eastern oyster (juvenile), <i>Crassostrea virginica</i>	F, U	99.7%	27	>2,000 (29°C)	-	Hansen 1980; Mayer 1987
Eastern oyster (juvenile), <i>Crassostrea virginica</i>	F, U	99.7%	17	>2,000 (20°C)	-	Hansen 1980
Eastern oyster (embryo), <i>Crassostrea virginica</i>	S, M	99%	-	2,700	2,386	Surprenant et al. 1985
Cockle clam (2-5 mm), <i>Clinocardium nuttalli</i>	R, U	80%	25	3,850	3,850	Butler et al. 1968
Bent-nosed clam (2.5 g), <i>Macoma nasuta</i>	R, U	80%	25	17,000	17,000	Armstrong and Millemann 1974b
Hard clam (embryo/larva), <i>Mercenaria mercenaria</i>	S, U	-	-	3,820 (24°C)	3,820	Davis and Hidu 1969
Mysid (24 hr) <i>Americamysis bahia</i>	R, U	99%	30	19 ^c	-	Thursby and Champlin 1991a
Mysid (juvenile), <i>Americamysis bahia</i>	F, M	-	-	>7.7	-	Nimmo et al. 1981

Appendix B. Acceptable Acute Toxicity Data of Carbaryl to Estuarine/Marine Aquatic Animals

Species	Method ^a	Chemical	Salinity (g/kg)	LC50 or EC50 (µg/L)	Species Mean Acute Value (µg/L)	References
Mysid (24 hr) <i>Americamysis bahia</i>	F, M	99%	31	8.46 (pH=7.85)	-	Thursby and Champlin 1991b
Mysid (<24 hr) <i>Americamysis bahia</i>	F, M	99.7%	20	5.7	7.188	Lintott 1992a
Mud shrimp (larva; 3 mm), <i>Upogebia pugettensis</i>	S, U	80%	25	90 (16°C)	-	Stewart et al. 1967
Mud shrimp (larva; 3 mm), <i>Upogebia pugettensis</i>	S, U	80%	25	40 (20°C)	60.0	Stewart et al. 1967
Ghost shrimp (larva; 3 mm), <i>Callinassa californiensis</i>	S, U	80%	25	80 (17°C)	-	Stewart et al. 1967
Ghost shrimp (larva; 3 mm), <i>Callinassa californiensis</i>	S, U	80%	25	30 (20°C)	48.99	Stewart et al. 1967
Dungeness crab (9 th stage), <i>Metacarcinus magister</i> (formerly <i>Cancer magister</i>)	R, U	80%	25	300 ^b (10°C)	-	Buchanan et al. 1970
Dungeness crab (9 th stage), <i>Metacarcinus magister</i> (formerly <i>Cancer magister</i>)	R, U	80%	25	280 ^b (10°C)	-	Buchanan et al. 1970
Dungeness crab (adult), <i>Metacarcinus magister</i> (formerly <i>Cancer magister</i>)	R, U	80%	25	180 ^b (18°C)	-	Buchanan et al. 1970
Dungeness crab (adult), <i>Metacarcinus magister</i> (formerly <i>Cancer magister</i>)	R, U	80%	25	260 ^b (11°C)	-	Buchanan et al. 1970
Dungeness crab (zoea), <i>Metacarcinus magister</i> (formerly <i>Cancer magister</i>)	S, U	80%	25	10 (10°C)	10	Buchanan et al. 1970
Sheepshead minnow (10.27 g; 24 mm), <i>Cyprinodon variegatus</i>	S, U	99%	32	2,200 ^c	-	Surprenant 1985a

Appendix B. Acceptable Acute Toxicity Data of Carbaryl to Estuarine/Marine Aquatic Animals

Species	Method ^a	Chemical	Salinity (g/kg)	LC50 or EC50 (µg/L)	Species Mean Acute Value (µg/L)	References
Sheepshead minnow (juvenile; 0.9 g; 29 mm), <i>Cyprinodon variegatus</i>	F, M	99.7%	20	2,600	2,600	Lintott 1992b
Threespine stickleback (0.38-0.77 g; adult), <i>Gasterosteus aculeatus</i>	S, U	95%	5	3,990	-	Katz 1961
Threespine stickleback (0.38-0.77 g; adult), <i>Gasterosteus aculeatus</i>	S, U	95%	25	3,990	3,990	Katz 1961

^a S = static; R = renewal; F = flow-through; M = measured; U = unmeasured.

^b Data not used to calculate SMAV because a more sensitive life stage or endpoint, or definite value available for the species.

^c Data not used to calculate SMAV because a more sensitive test exposure available for the species.

Dash indicates not available

Appendix C

Appendix C. Acceptable Chronic Toxicity Data of Carbaryl to Freshwater Aquatic Animals

Species	Test ^a	Chemical	Hardness (mg/L as CaCO ₃)	Chronic Limits ^b (µg/L)	Chronic Value ^c (µg/L)	Species Mean Chronic Value (µg/L)	Reference
Freshwater Species							
Cladoceran (<12 hr), <i>Ceriodaphnia dubia</i>	LC	>99%	57	-	10.6	10.6	Oris et al. 1991
Cladoceran (≤24 hr), <i>Daphnia magna</i>	LC	99%	140-170	1.5-3.3	2.225	-	Surprenant 1985b
Cladoceran, <i>Daphnia magna</i>	LC	98%	201.5	4.04-10.1 ^d	6.389	3.770	Brooke 1991
Fathead minnow, <i>Pimephales promelas</i>	LC	80%	41-49	210-680	377.9	-	Carlson 1971
Fathead minnow, <i>Pimephales promelas</i>	ELS	99%	44-49	720-1,600	1,073	636.8	Norberg-King 1989
Bonytail chub (48 d), <i>Gila elegans</i>	ELS	99%	344-378	650-1,240	897.8	897.8	Beyers et al. 1994
Colorado pikeminnow (formerly squawfish) (41 d), <i>Ptychocheilus lucius</i>	ELS	99%	344-378	445-866	620.8	620.8	Beyers et al. 1994

^a LC = life-cycle or partial life-cycle; ELS = early life-stage.

^b NOEC is listed first, then the LOEC.

^c Chronic value = calculated geometric mean of NOEC and LOEC

^d LOEC is based on the EC50 value obtained in a parallel acute test conducted in the same study (Brooke 1991).

Chronic value should be considered an upper bound value for the species from this study.

Dash indicates not available

Appendix D

Appendix D. Chronic Toxicity Data of Carbaryl to Estuarine/Marine Aquatic Animals used Qualitatively in the Assessment

Species	Test ^a	Chemical	Salinity (g/kg)	Chronic Limits ^b (µg/L)	Chronic Value ^c (µg/L)	Species Mean Chronic Value (µg/L)	Reference
Estuarine/marine Species							
Mysid (24 hr), <i>Americamysis bahia</i>	LC	99%	31.5	7.18-13.7	9.918	9.918	Thursby and Champlin 1991b

^a LC = life-cycle or partial life-cycle; ELS = early life-stage.

^b NOEC is listed first, then the LOEC.

^c Chronic value = calculated geometric mean of NOEC and LOEC

Appendix E

Appendix E. Acceptable Toxicity Data of Carbaryl to Freshwater Aquatic Plants

Species	Chemical	Method ^a	Hardness (mg/L as CaCO ₃)	Duration (days)	Effect	Concentration (µg/L)	Reference
Freshwater Species							
Blue green alga, <i>Nostoc muscorum</i>	99.9%	S,U	-	4	Chronic Value (growth)	7,071	Bhunja et al. 1994
Blue green alga, <i>Anabaena torulosa</i>	99.9%	S,U	-	21	LOEC (growth)	50,000	Obulakondaiah et al. 1993
Green alga, <i>Pseudokirchneriella subcapitata</i>	Analytical	S,U	-	4	EC20 (growth, cell #)	1,040	Versteeg 1990
Green alga, <i>Pseudokirchneriella subcapitata</i>	99.7%	S, M	-	5	Chronic Value (cell number)	537.2	Lintott 1992c
Green alga, <i>Pseudokirchneriella subcapitata</i>	98%	-	81.9	4	IC50 ^b	490	Brooke 1993
Green alga, <i>Scenedesmus quadricauda</i>	97%	S,U	-	10	LOEC (chlorophyll content)	100	Lejczak 1977
Green alga, <i>Scenedesmus quadricauda</i>	97%	-	-	10	Reduced chlorophyll content	100	Lejczak 1977
Diatom, <i>Navicula pelliculosa</i>	99.7%	S, M	-	5	LOEC (cell number)	400	Lintott 1992d
Duckweed, <i>Lemna minor</i>	98%	-	171	4	IC50 ^b	23,900	Brooke 1993

^a S = static; R = renewal; F = flow-through; M = measured; U = unmeasured.

^b IC50 = concentration of carbaryl at which growth is inhibited 50% compared to control organism growth.

Dash indicates not available

Appendix F

Appendix F. Acceptable Toxicity Data of Carbaryl to Estuarine/Marine Aquatic Plants

Species	Chemical	Method ^a	Salinity (g/kg)	Duration (days)	Effect	Concentration (µg/L)	Reference
Estuarine/marine Species							
Green alga, <i>Chlorococcum sp.</i>	-	S, U	-	4	EC50 (growth)	2,100	Walsh and Alexander 1980
Green alga, <i>Chlorococcum sp.</i>	-	S, U	-	4	EC50 (growth)	1,800	Walsh and Alexander 1980
Green alga, <i>Chlorococcum sp.</i>	-	S, U	-	12	EC50 (growth)	2,700	Walsh and Alexander 1980
Green alga, <i>Chlorococcum sp.</i>	-	S, U	-	12	EC50 (growth)	2,900	Walsh and Alexander 1980
Green alga, <i>Chlorella sp.</i>	-	S, U	-	4	EC50 (growth)	1,000	Walsh and Alexander 1980
Green alga, <i>Chlorella sp.</i>	-	S, U	-	4	EC50 (growth)	600	Walsh and Alexander 1980
Green alga, <i>Chlorella sp.</i>	-	S, U	-	12	EC50 (growth)	1,200	Walsh and Alexander 1980
Green alga, <i>Chlorella sp.</i>	-	S, U	-	12	EC50 (growth)	1,400	Walsh and Alexander 1980
Green alga, <i>Chlorella sp.</i>	95%	S, U	-	10	10% reduced growth rate	100	Ukeles 1962
Green alga, <i>Chlorella sp.</i>	95%	S, U	-	10	20% reduced growth rate	1,000	Ukeles 1962
Green alga, <i>Chlorella sp.</i>	95%	S, U	-	10	100% reduced growth rate, cells not viable	10,000	Ukeles 1962
Green alga, <i>Chlorella sp.</i>	95%	S, U	-	10	100% reduced growth rate, cells not viable	100,000	Ukeles 1962
Green alga, <i>Dunaliella euchora</i>	95%	S, U	-	10	10% reduced growth rate	100	Ukeles 1962
Green alga, <i>Dunaliella euchora</i>	95%	S, U	-	10	35% reduced growth rate	1,000	Ukeles 1962
Green alga, <i>Dunaliella euchora</i>	95%	S, U	-	10	100% reduced growth rate, cell viable	10,000	Ukeles 1962
Green alga, <i>Dunaliella euchora</i>	95%	S, U	-	10	100% reduced growth rate, cells not viable	100,000	Ukeles 1962
Green alga, <i>Protococcus sp.</i>	95%	S, U	-	10	20% reduced growth rate	100	Ukeles 1962
Green alga, <i>Protococcus sp.</i>	95%	S, U	-	10	26% reduced growth rate	1,000	Ukeles 1962
Green alga, <i>Protococcus sp.</i>	95%	S, U	-	10	100% reduced growth rate, cell viable	10,000	Ukeles 1962
Green alga, <i>Protococcus sp.</i>	95%	S, U	-	10	100% reduced growth rate, cells not viable	100,000	Ukeles 1962

Appendix F. Acceptable Toxicity Data of Carbaryl to Estuarine/Marine Aquatic Plants

Species	Chemical	Method ^a	Salinity (g/kg)	Duration (days)	Effect	Concentration (µg/L)	Reference
Diatom, <i>Skeletonema costatum</i>	-	S, U	-	4	EC50 (growth)	1,700	Walsh and Alexander 1980
Diatom, <i>Skeletonema costatum</i>	-	S, U	-	4	EC50 (growth)	900	Walsh and Alexander 1980
Diatom, <i>Skeletonema costatum</i>	-	S, U	-	12	EC50 (growth)	1,800	Walsh and Alexander 1980
Diatom, <i>Skeletonema costatum</i>	-	S, U	-	12	EC50 (growth)	1,600	Walsh and Alexander 1980
Diatom, <i>Nitzschia angularum</i>	-	S, U	-	4	EC50 (growth)	1,500	Walsh and Alexander 1980
Diatom, <i>Nitzschia angularum</i>	-	S, U	-	4	EC50 (growth)	1,000	Walsh and Alexander 1980
Diatom, <i>Nitzschia angularum</i>	-	S, U	-	12	EC50 (growth)	1,600	Walsh and Alexander 1980
Diatom, <i>Nitzschia angularum</i>	-	S, U	-	12	EC50 (growth)	1,500	Walsh and Alexander 1980
Diatom, <i>Phaeodactylum tricornutum</i>	95%	S, U	-	10	100% reduced growth rate, cells viable	100	Ukeles 1962
Diatom, <i>Phaeodactylum tricornutum</i>	95%	S, U	-	10	100% reduced growth rate, cells viable	1,000	Ukeles 1962
Diatom, <i>Phaeodactylum tricornutum</i>	95%	S, U	-	10	100% reduced growth rate, cells viable	10,000	Ukeles 1962
Diatom, <i>Phaeodactylum tricornutum</i>	95%	S, U	-	10	100% reduced growth rate, cells viable	100,000	Ukeles 1962
Chrysophyte, <i>Monochrysis lutheri</i>	95%	S, U	-	10	13% reduced growth rate	100	Ukeles 1962
Chrysophyte, <i>Monochrysis lutheri</i>	95%	S, U	-	10	100% reduced growth rate, cells viable	1,000	Ukeles 1962
Chrysophyte, <i>Monochrysis lutheri</i>	95%	S, U	-	10	100% reduced growth rate, cells not viable	10,000	Ukeles 1962
Chrysophyte, <i>Monochrysis lutheri</i>	95%	S, U	-	10	100% reduced growth rate, cells not viable	100,000	Ukeles 1962

^a S = static; R = renewal; F = flow-through; M = measured; U = unmeasured.

Dash indicates not available

Appendix G

Appendix G. Acceptable Bioaccumulation Data of Carbaryl by Aquatic Organisms

There are no acceptable bioaccumulation data for carbaryl.

Appendix H

Appendix H. Other Data on Effects of Carbaryl to Freshwater Aquatic Organisms

Species	Chemical	Duration	Effect	Concentration (µg/L)	Reference	Reason Other Data
Freshwater Species						
Bacterium, <i>Vibrio fischeri</i>	Analytical	30 min	EC50 (luminescence)	2,440	Hernando et al. 2007	Single-cell organism
Bacteria, <i>Pseudomonas putida</i>	-	-	LOEC (reduced cell growth)	>5,000	Bringmann and Kuhn 1977	Single-cell organism
Planktonic algae mixture	99.7%	14 day	LOEC (growth inhibition)	10,000	Butler et al. 1975	Community exposure
Blue-green algae mixture	-	24 hr	Reduced carbon uptake	500	Vickers and Boyd 1971	Community exposure
Blue-green algae, <i>Microcystis aeruginosa</i>	-	8 day	LOEC (reduced cell growth)	1,350	Bringmann and Kuhn 1978a; b	Lack of exposure details
Green alga, <i>Ankistrodesmus braunii</i>	-	48 hr	Reduced growth	25,000	Kopecek et al. 1975	Duration
Green algae, <i>Chlorella pyrenoidosa</i>	-	96 hr	LOEC (growth)	100	Christie 1969	Lack of exposure details
Green alga, <i>Raphidocelis subcapitata</i>	Analytical	6 day	LOEC (growth)	5,030	Bierkens et al. 1998	Lack of exposure details
Green alga, <i>Raphidocelis subcapitata</i>	Analytical	6 day	LOEC (heat shock protein 70)	2,072	Bierkens et al. 1998	Atypical endpoint
Green algae, <i>Scenedesmus quadricaudata</i>	-	6 day	Increased cell biomass	100	Stadnyk et al. 1971	Lack of exposure details
Green algae, <i>Scenedesmus quadricaudata</i>	-	8 day	Reduced cell growth	1,400	Bringmann and Kuhn 1977; 1978a; b	Lack of exposure details
Ciliate protozoa, <i>Colpidium campylum</i>	-	43 hr	LOEC (growth inhibition)	<10,000	Dive et al. 1980	Duration
Protozoa (7 day), <i>Paramecium aurelia</i>	97.5%	24 hr	LC50	46,000	Edmiston et al. 1984	Single-cell organism
Protozoa (7 day), <i>Paramecium bursaria</i>	97.5%	24 hr	LC50	31,000	Edmiston et al. 1984	Single-cell organism
Protozoa (7 day), <i>Paramecium caudatum</i>	97.5%	24 hr	LC50	10,000	Edmiston et al. 1984	Single-cell organism
Protozoa (7 day), <i>Paramecium multimicronucleatum</i>	97.5%	24 hr	LC50	24,000	Edmiston et al. 1984	Single-cell organism
Protozoa (12 day), <i>Paramecium multimicronucleatum</i>	97.5%	24 hr	LC50	28,000	Edmiston et al. 1985	Single-cell organism
Tubificid worm, <i>Branchiura sowerbyi</i>	-	72 hr	LOEC (mortality)	4,000	Naqvi 1973	Duration

Appendix H. Other Data on Effects of Carbaryl to Freshwater Aquatic Organisms

Species	Chemical	Duration	Effect	Concentration (µg/L)	Reference	Reason Other Data
Snail (< 24 hr), <i>Lymnea stagnalis</i>	97-99%	1 mo	82.5% mortality	2,000	Seuge and Bluzat 1983	Unmeasured exposure
Snail, <i>Physa acuta</i>	Technical	48 hr	LC50	27,000	Hashimoto and Nishiuchi 1981	Duration
Cladoceran (<24 hr), <i>Ceriodaphnia dubia</i>	99.7%	7 day	IC25	<330	Dwyer et al. 1999a	Static, unmeasured exposure
Cladoceran (2 nd instar), <i>Daphnia galeata mendotae</i>	>99%	8-14 hr	LOEC (enlarged helmet)	5	Hanazato and Dodson 1993	Duration
Cladoceran (1 st and 2 nd instar), <i>Daphnia lumholtzi</i>	>99%	8-14 hr	LOEC (enlarged helmet)	10	Hanazato and Dodson 1993	Duration
Cladoceran (<26 hr), <i>Daphnia magna</i>	-	24 hr	LC50	1.1	Gaaboub et al. 1975	Duration
Cladoceran (<24 hr), <i>Daphnia magna</i>	Technical grade	0.5 hr	EC50 (erratic swimming)	12	Parker et al. 1970	Duration
Cladoceran (0-24 hr), <i>Daphnia magna</i>	-	24 hr	LC50	0.66	Rawash et al. 1975	Duration
Cladoceran (4 day), <i>Daphnia magna</i>	-	24 hr	LC50	21	Wernersson and Dave 1997	Duration
Cladoceran (2 nd instar), <i>Daphnia pulex</i>	>99%	8-14 hr	LOEC (enlarged helmet)	15	Hanazato and Dodson 1993	Duration
Cladoceran, <i>Daphnia pulex</i>	-	3 hr	LC50	0.85	Nishiuchi and Hashimoto 1967	Duration
Cladoceran, <i>Daphnia pulex</i>	Technical	3 hr	LC50	50	Hashimoto and Nishiuchi 1981	Duration
Cladoceran (1 st and 2 nd instar), <i>Daphnia retrocurva</i>	>99%	8-14 hr	LOEC (enlarged helmet)	20	Hanazato and Dodson 1993	Duration
Cladoceran, <i>Moina macrocopa</i>	-	3 hr	LC50	1.0	Nishiuchi and Hashimoto 1967	Duration
Cladoceran, <i>Moina macrocopa</i>	Technical	3 hr	LC50	50	Hashimoto and Nishiuchi 1981	Duration
Cladoceran (adult female), <i>Bosmina longirostris</i>	>99%	24 hr	LC50	8.597	Sakamoto et al. 2005	Duration
Cladoceran (juvenile/adult), <i>Leptodora kindtii</i>	>99%	24 hr	LC50	3.477	Sakamoto et al. 2005	Duration
Ostracod, <i>Cyprretta kawatai</i>	99.9%	72 hr	LC50	1,800	Hansen and Kawatski 1976	Duration
Opossum shrimp (adult), <i>Mysis relicta</i>	98%	6 hr	Avg. BCF = 149	7.24 and 15.95	Landrum and Dupuis 1990	Duration
Seed shrimp (mature), <i>Cypridopsis vidua</i>	99.5%	48 hr	EC50	115	Johnson and Finley 1980	Duration

Appendix H. Other Data on Effects of Carbaryl to Freshwater Aquatic Organisms

Species	Chemical	Duration	Effect	Concentration (µg/L)	Reference	Reason Other Data
Amphipod (mature), <i>Pontoporeia hoyi</i>	98%	6 hr	Avg. BCF = 18,700	1.44-3.81	Landrum and Dupuis 1990	Duration
Amphipod, <i>Gammarus pseudolimnaeus</i>	99.5%	96 hr	LC50	16	Schoettger and Mauck 1978	Control survival not reported
Amphipod (mature), <i>Gammarus pseudolimnaeus</i>	99.5%	48 hr	LC50	13	Mayer and Ellersieck 1986	Duration
Amphipod (mature), <i>Gammarus pseudolimnaeus</i>	99.5%	48 hr	LC50	8	Mayer and Ellersieck 1986	Duration
Grass shrimp, <i>Palaemonetes kadiakensis</i>	-	24 hr	LC50	42.5	Naqvi and Ferguson 1970	Duration
Red Crayfish, <i>Procambarus clarkii</i>	Technical	72 hr	LC50	2,000	Muncy and Oliver 1963	Duration
Mayfly (nymph), <i>Cloeon dipterum</i>	Technical	48 hr	LC50	370	Hashimoto and Nishiuchi 1981	Duration
Stonefly (1 st year class), <i>Isogenus sp.</i>	99.5%	96 hr	LC50 (solution aged 7 days)	6.6	Mayer and Ellersieck 1986	Aged solution
Stonefly (1 st year class), <i>Isogenus sp.</i>	99.5%	96 hr	LC50 (solution aged 14 days)	6.6	Mayer and Ellersieck 1986	Aged solution
Stonefly (1 st year class), <i>Isogenus sp.</i>	99.5%	96 hr	LC50 (solution aged 21 days)	12	Mayer and Ellersieck 1986	Aged solution
Stonefly (nymph), <i>Pteronarcys californica</i>	99.5%	96 hr	LC50	2	Schoettger and Mauck 1978	Control survival not reported
Crawling water beetle (5 mg), <i>Peltodytes sp.</i>	≥94%	96 hr	LC50	3,300	Federle and Collins 1976	Three species exposed simultaneously
Blackfly (larva), <i>Simulium vittatum</i>	≥98%	48 hr	LC50	23.72	Overmyer et al. 2003	Duration
Midge (4 th instar), <i>Chironomus plumosus</i>	99.5%	48 hr	EC50	10	Sanders et al. 1983	Duration
Midge (4 th instar), <i>Chironomus riparius</i>	Analytical	24 hr	EC50 (reduced lactate)	1.0	Forcella et al. 2007	Duration
Midge (4 th instar), <i>Chironomus riparius</i>	>98%	24 hr	LC50	110	Lydy et al. 1990	Duration
Midge (4 th instar), <i>Chironomus tentans</i>	99.9%	72 hr	LC50	5,900	Hansen and Kawatski 1976	Duration
Mosquito (2 nd stage), <i>Aedes aegypti</i>	97.6%	36 hr	LC50	336	Lakota et al. 1981	Duration
Mosquito (4 th instar larvae), <i>Culex pipiens</i>	-	24 hr	LC50	75	Rawash et al. 1975	Duration

Appendix H. Other Data on Effects of Carbaryl to Freshwater Aquatic Organisms

Species	Chemical	Duration	Effect	Concentration (µg/L)	Reference	Reason Other Data
Mosquito (4 th instar larvae), <i>Culex pipiens</i>	-	24 hr	LC50	170	Gaoubou et al. 1975	Duration
Apache trout, <i>Oncorhynchus apache</i>	-	96 hr	NOEC (muscarinic cholinergic receptors)	1,300	Jones et al. 1998	Atypical endpoint
Coho salmon (6-18 mo), <i>Oncorhynchus kisutch</i>	-	≤ 30 day	BCF = 3.58 (substantial mortality at 30 days)	2,600	Walsh and Ribelin 1973	Control issues
Coho salmon (4.9 cm; 1.3 g; 4-7 mo), <i>Oncorhynchus kisutch</i>	99%	96 hr	LC50	>150	Laetz et al. 2009	Control survival not reported
Rainbow trout, <i>Oncorhynchus mykiss</i>	99%	96 hr	EC50 (muscular cholinesterase)	270	Ferrari et al. 2007a	Atypical endpoint
Rainbow trout, <i>Oncorhynchus mykiss</i>	99%	96 hr	EC50 (brain cholinesterase)	19.24	Ferrari et al. 2007a	Atypical endpoint
Rainbow trout (2 yr), <i>Oncorhynchus mykiss</i>	-	48 hr	LC0	30,000	Lysak and Marcinek 1972	Duration
Rainbow trout (2 yr), <i>Oncorhynchus mykiss</i>	-	24 hr	LC100	100,000	Lysak and Marcinek 1972	Duration
Rainbow trout (1-1.5 yr), <i>Oncorhynchus mykiss</i>	-	4.5 hr	LOEC (respiration rate)	2,000	Lunn et al. 1976	Duration
Rainbow trout (1-1.5 yr), <i>Oncorhynchus mykiss</i>	-	4.5 hr	NOEC (respiration rate)	1,000	Lunn et al. 1976	Duration
Rainbow trout (1.8 g), <i>Oncorhynchus mykiss</i>	99.5%	24 hr	LC50	4,620	Mayer and Ellersieck 1986	Duration
Rainbow trout (2.2 g), <i>Oncorhynchus mykiss</i>	99.5%	24 hr	LC50	6,090	Mayer and Ellersieck 1986	Duration
Rainbow trout (30-80 g), <i>Oncorhynchus mykiss</i>	Technical	24 hr	LC50	1,410	Zinkl et al. 1987	Duration
Rainbow trout (30-80 g), <i>Oncorhynchus mykiss</i>	Technical	24 hr	LOEC (depression of brain cholinesterase activity)	500	Zinkl et al. 1987	Atypical endpoint
Rainbow trout (0.5-1.0 g), <i>Oncorhynchus mykiss</i>	99.9%	96 hr	LOEC (survival from predation)	10	Little et al. 1990	Atypical endpoint
Rainbow trout, <i>Oncorhynchus mykiss</i>	-	96 hr	NOEC (muscarinic cholinergic receptors)	3,600	Jones et al. 1998	Atypical endpoint

Appendix H. Other Data on Effects of Carbaryl to Freshwater Aquatic Organisms

Species	Chemical	Duration	Effect	Concentration (µg/L)	Reference	Reason Other Data
Lahontan cutthroat trout, <i>Oncorhynchus clarkii henshawi</i>	-	96 hr	NOEC (muscarinic cholinergic receptors)	2,200	Jones et al. 1998	Atypical endpoint
Cutthroat trout (0.5 g), <i>Oncorhynchus clarkii</i>	99.5%	96 hr	LC50 (solution aged 7 days)	4,000 (pH=7.3)	Mayer and Ellersieck 1986	Aged solution
Cutthroat trout (0.5 g), <i>Oncorhynchus clarkii</i>	99.5%	96 hr	LC50 (solution aged 14 days)	3,380 (pH=7.2)	Mayer and Ellersieck 1986	Aged solution
Cutthroat trout (0.5 g), <i>Oncorhynchus clarkii</i>	99.5%	96 hr	LC50 (solution aged 21 days)	2,300 (pH=7.2)	Mayer and Ellersieck 1986	Aged solution
Atlantic salmon (3-4 cm), <i>Salmo salar</i>	-	24 hr	NOEC (temperature preference)	1,000	Peterson 1976	Atypical endpoint
Atlantic salmon (0.4 g), <i>Salmo salar</i>	99.5%	24 hr	LC50 (pH=6.5)	1,400	Mayer and Ellersieck 1986	Duration
Brook trout, <i>Salvelinus fontinalis</i>	99.5%	96 hr	LC50 (pH=7.5) (7°C)	3,000	Schoettger and Mauck 1978	Control survival not reported
Brook trout, <i>Salvelinus fontinalis</i>	99.5%	96 hr	LC50 (pH=7.5) (12°C)	2,100	Schoettger and Mauck 1978	Control survival not reported
Brook trout, <i>Salvelinus fontinalis</i>	99.5%	96 hr	LC50 (pH=7.5) (17°C)	900	Schoettger and Mauck 1978	Control survival not reported
Brook trout, <i>Salvelinus fontinalis</i>	99.5%	96 hr	LC50 (pH=8) (12°C)	5,400	Schoettger and Mauck 1978	Control survival not reported
Brook trout, <i>Salvelinus fontinalis</i>	99.5%	96 hr	LC50 (pH=8) (12°C)	2,500	Schoettger and Mauck 1978	Control survival not reported
Brook trout, <i>Salvelinus fontinalis</i>	99.5%	96 hr	LC50 (pH=6.5) (12°C)	4,600	Schoettger and Mauck 1978	Control survival not reported
Brook trout, <i>Salvelinus fontinalis</i>	99.5%	96 hr	LC50 (pH=7.5) (12°C)	3,700	Schoettger and Mauck 1978	Control survival not reported
Brook trout, <i>Salvelinus fontinalis</i>	99.5%	96 hr	LC50 (pH=8.5) (12°C)	2,100	Schoettger and Mauck 1978	Control survival not reported
Brook trout, <i>Salvelinus fontinalis</i>	99.5%	96 hr	LC50 (pH=9.0) (12°C)	1,100	Schoettger and Mauck 1978	Control survival not reported
Brook trout (0.8 g), <i>Salvelinus fontinalis</i>	99.5%	24 hr	LC50	10,000	Mayer and Ellersieck 1986	Duration

Appendix H. Other Data on Effects of Carbaryl to Freshwater Aquatic Organisms

Species	Chemical	Duration	Effect	Concentration (µg/L)	Reference	Reason Other Data
Lake trout (6-18 month old), <i>Salvelinus namaycush</i>	-	≤ 30 day	BCF = 4.00 (substantial mortality at 30 days)	2,600	Walsh and Ribelin 1973	Control issues
Goldfish, <i>Carassius auratus</i>	-	48 hr	LC50	>10,000	Nishiuchi and Hashimoto 1967	Duration
Goldfish, <i>Carassius auratus</i>	99%	10 day	LC50	>10,000	Shea and Berry 1983	Duration
Goldfish, <i>Carassius auratus</i>	-	7 day	40% with vertebral deformation	5,000	Imada 1976	Duration
Carp, <i>Cyprinus carpio</i>	Technical	48 hr	LC50	13,000	Hashimoto and Nishiuchi 1981	Duration
Common carp (larva; 8 mm; 3 day), <i>Cyprinus carpio</i>	Commercial	96 hr	LC50	2,000	Verma et al. 1981a	Formulation
Common carp (larva; 8 mm; 3 day), <i>Cyprinus carpio</i>	Commercial	60 day	NOEC (growth)	50	Verma et al. 1981a	Formulation
Common carp (larva; 8 mm; 3 day), <i>Cyprinus carpio</i>	Commercial	60 day	LOEC (growth)	80	Verma et al. 1981a	Formulation
Common carp, <i>Cyprinus carpio</i>	-	48 hr	LC50	>10,000	Nishiuchi and Hashimoto 1967	Duration
Common carp (20-34 mm), <i>Cyprinus carpio</i>	85%	7 & 14 day	LOEC (reduced AChE activity in brain)	39	de Mel and Pathiratne 2005	Atypical endpoint
Common carp (egg), <i>Cyprinus carpio</i>	-	55-74 hr	LC50	1,400	Kaur and Toor 1977	Duration
Common carp (egg), <i>Cyprinus carpio</i>	-	55-74 hr	100% mortality	2,500	Kaur and Toor 1977	Duration
Carp (1.2 g), <i>Cyprinus carpio</i>	Technical	72 hr	LC50	13,000	Nishiuchi and Asano 1981	Duration
Carp (3.0 g), <i>Cyprinus carpio</i>	Technical	48 hr	LC50	13,000	Nishiuchi and Asano 1981	Duration
Zebrafish (embryo), <i>Danio rerio</i>	99.9%	24 hr	LC50	44,660	Lin et al. 2007	Duration
Zebrafish (embryo), <i>Danio rerio</i>	99.9%	24 hr	EC50 (pericardial edema, tail malformations)	7,520	Lin et al. 2007	Duration
Fathead minnow (1-2 g), <i>Pimephales promelas</i>	95%	96 hr	LC50	13,000	Henderson et al. 1960	Control survival not reported
Fathead minnow (1-2 g), <i>Pimephales promelas</i>	95%	96 hr	LC50	7,000	Henderson et al. 1960	Control survival not reported
Fathead minnow (larva), <i>Pimephales promelas</i>	99.9%	7 day	Chronic Value (growth)	576	Norberg-King 1989	Duration

Appendix H. Other Data on Effects of Carbaryl to Freshwater Aquatic Organisms

Species	Chemical	Duration	Effect	Concentration (µg/L)	Reference	Reason Other Data
Fathead minnow (larva), <i>Pimephales promelas</i>	99.9%	12 day	Chronic Value (growth)	1,275	Norberg-King 1989	Duration
Fathead minnow (larva), <i>Pimephales promelas</i>	99.9%	7 day	Chronic Value (survival and growth)	1,088	Norberg-King 1989	Duration
Fathead minnow (larva), <i>Pimephales promelas</i>	99.9%	7 day	Chronic Value (survival)	1,018	Norberg-King 1989	Duration
Fathead minnow (larva), <i>Pimephales promelas</i>	99.9%	7 day	Chronic Value (growth)	569	Norberg-King 1989	Duration
Fathead minnow (larva), <i>Pimephales promelas</i>	99.9%	7 day	Chronic Value (survival and growth)	976	Norberg-King 1989	Duration
Fathead minnow (1 day), <i>Pimephales promelas</i>	99.8%	7 day	Chronic Value (growth)	707	Pickering et al. 1996	Duration
Fathead minnow (4 day), <i>Pimephales promelas</i>	99.8%	7 day	Chronic Value (growth)	<250	Pickering et al. 1996	Duration
Fathead minnow (7 day), <i>Pimephales promelas</i>	99.8%	7 day	Chronic Value (growth)	707	Pickering et al. 1996	Duration
Fathead minnow (<24 hr), <i>Pimephales promelas</i>	99.7%	7 day	IC25	420	Dwyer et al. 1999a	Duration
Colorado pikeminnow (formerly squawfish), <i>Ptychocheilus lucius</i>	99%	24 hr	NOEC (AChE inhibition)	29.3	Beyers and Sikoski 1994	Duration
Colorado pikeminnow, <i>Ptychocheilus lucius</i>	99%	24 hr	LOEC (AChE inhibition)	49.1	Beyers and Sikoski 1994	Duration
Colorado pikeminnow (5-6 day), <i>Ptychocheilus lucius</i>	99.7%	7 day	IC25	1,330	Dwyer et al. 1999a	Duration
Bonytail chub (2-7 day), <i>Gila elegans</i>	99.7%	7 day	IC25	250	Dwyer et al. 1999a	Duration
Razorback sucker (6-7 day), <i>Xyrauchen texanus</i>	99.7%	7 day	IC25	2,060	Dwyer et al. 1999a	Duration
Yellow bullhead (adult), <i>Ictalurus natalis</i>	80%	48 hr	LOEC (behavior)	1,000	Morison 1984	Duration
Walking catfish (15-20 cm; 20-30 g), <i>Clarias batrachus</i>	99%	15 day	Decreased glucose and protein	4,000	Sharma 1999	Atypical endpoint

Appendix H. Other Data on Effects of Carbaryl to Freshwater Aquatic Organisms

Species	Chemical	Duration	Effect	Concentration (µg/L)	Reference	Reason Other Data
Walking catfish (15-20 cm; 70-90 g), <i>Clarias batrachus</i>	99%	96 hr	Increased lactic acid in liver and muscle	1,000	Sharma and Gopal 1995	Atypical endpoint
Walking catfish (15-20 cm; 70-90 g), <i>Clarias batrachus</i>	99%	96 hr	Increased lactic acid in heart, gills, kidney and spleen	2,000	Sharma and Gopal 1995	Atypical endpoint
Walking catfish (15-20 cm; 70-90 g), <i>Clarias batrachus</i>	99%	96 hr	Reduced AChE levels in various tissues	1,000	Sharma et al. 1993	Atypical endpoint
Killifish, <i>Fundulus heteroclitus</i>	99%	10 day	40% Mortality	10,000	Shea and Berry 1983	Duration
Mosquitofish (20-45 mm), <i>Gambusia affinis</i>	98%	24 hr	Irritated	10,000	Hansen et al. 1972	Duration
Mosquitofish, <i>Gambusia affinis</i>	Analytical	24 hr	Avoidance	13,000	Krieger and Lee 1973	Duration
Mosquitofish, <i>Gambusia affinis</i>	Analytical	24 hr	LC50	13,000	Krieger and Lee 1973	Duration
Guppy (juvenile), <i>Poecilia reticulata</i>	97.6%	36 hr	LC50	3,840	Lakota et al. 1981	Duration
Bluegill (adult), <i>Lepomis macrochirus</i>	-	96 hr	Increased ventilator response	2,400	Carlson 1990	Atypical endpoint
Bluegill (1-2 g), <i>Lepomis macrochirus</i>	95%	96 hr	LC50	5,600	Henderson et al. 1960	Control survival not reported
Bluegill (0.87 g), <i>Lepomis macrochirus</i>	Technical	96 hr	LC50	2,000	Cope 1965	Control survival not reported
Mozambique tilapia, <i>Tilapia mossambica</i>	Commercial	48 hr	LC50	5,495	Basha et al. 1983	Duration
Plains Leopard frog, <i>Rana blairi</i>	99.7%	24 hr	LOEC (reduced activity)	3,500	Bridges 1997	Duration
Gray tree frog (Gosner stage 25), <i>Hyla versicolor</i>	99.7%	48 hr	LC50	>2,510	Little et al. 2000	Duration
African clawed frog (embryo), <i>Xenopus laevis</i>	-	24 hr	LC50	4,700	Elliott-Freeley and Armstrong 1982	Duration
African clawed frog (tadpole), <i>Xenopus laevis</i>	-	24 hr	Abnormal development	110	Elliott-Freeley and Armstrong 1982	Duration
African clawed frog (embryo), <i>Xenopus laevis</i>	99.8%	115 hr	LC50	20,280	Bacchetta et al. 2008	Duration

Dash indicates not available

Appendix I

Appendix I. Other Data on Effects of Carbaryl to Estuarine/Marine Aquatic Organisms

Species	Chemical	Duration	Effect	Concentration (µg/L)	Reference	Reason Other Data
Estuarine/marine Species						
Bacteria	99%	34 day	Reduction in diversity	100,000	Weber and Rosenberg 1984	Single-cell organism
Bacteria	99%	42 day (field)	No effect on numbers, diversity or filter paper composition	1,000	Weber and Rosenberg 1984	Single-cell organism
Bacteria	99%	42 day (field)	No effect on numbers, diversity or filter paper composition	10,000	Weber and Rosenberg 1984	Single-cell organism
Bacteria	99%	42 day (field)	No effect on numbers, diversity or filter paper composition	100,000	Weber and Rosenberg 1984	Single-cell organism
Bacteria	99%	70 day (lab)	No effect on filter paper decomposition	1,000	Weber and Rosenberg 1984	Single-cell organism
Bacteria	99%	70 day (lab)	No effect on filter paper decomposition	10,000	Weber and Rosenberg 1984	Single-cell organism
Bacteria	99%	70 day (lab)	Complete inhibition of filter paper decomposition	100,000	Weber and Rosenberg 1984	Single-cell organism
Bacterium, <i>Vibrio fischeri</i>	Analytical	30 min	EC50 (luminescence)	1,651	Hernando et al. 2007	Single-cell organism
Green alga, <i>Chlorococcum sp.</i>	-	48 hr	75% reduced growth rate	10,000	Maly and Ruber 1983	Duration
Green alga, <i>Chlorococcum sp.</i>	-	48 hr	40% reduced growth rate	2,000	Maly and Ruber 1983	Duration
Green alga, <i>Chlorococcum sp.</i>	-	48 hr	48% reduced growth rate	1,000	Maly and Ruber 1983	Duration
Green alga, <i>Chlorococcum sp.</i>	-	48 hr	No significant reduction in growth rate	500	Maly and Ruber 1983	Duration
Green alga, <i>Chlorococcum sp.</i>	-	96 hr	>100% reduced growth rate	10,000	Maly and Ruber 1983	Lack of exposure details
Green alga, <i>Chlorococcum sp.</i>	-	96 hr	92% reduction in growth rate between 48 & 96 hr	2,000	Maly and Ruber 1983	Lack of exposure details
Green alga, <i>Chlorococcum sp.</i>	-	96 hr	54% reduction in growth rate between 48 & 96 hr	1,000	Maly and Ruber 1983	Lack of exposure details
Green alga, <i>Chlorococcum sp.</i>	-	96 hr	No sign. reduction in growth rate between 48 and 96 hr	500	Maly and Ruber 1983	Lack of exposure details
Diatom, <i>Nitzschia closterium</i>	-	48 hr	>100% reduced growth rate	10,000	Maly and Ruber 1983	Duration
Diatom, <i>Nitzschia closterium</i>	-	48 hr	69% reduced growth rate	2,000	Maly and Ruber 1983	Duration
Diatom, <i>Nitzschia closterium</i>	-	48 hr	No significant reduction in growth rate	1,000	Maly and Ruber 1983	Duration

Appendix I. Other Data on Effects of Carbaryl to Estuarine/Marine Aquatic Organisms

Species	Chemical	Duration	Effect	Concentration (µg/L)	Reference	Reason Other Data
Diatom, <i>Nitzschia closterium</i>	-	96 hr	No significant reduction in growth rate between 48 and 96 hr	10,000	Maly and Ruber 1983	Lack of exposure details
Diatom, <i>Amphora coffeaformis v. borealis</i>	-	48 hr	>100% reduced growth rate	10,000	Maly and Ruber 1983	Duration
Diatom, <i>Amphora coffeaformis v. borealis</i>	-	48 hr	>100% reduced growth rate	2,000	Maly and Ruber 1983	Duration
Diatom, <i>Amphora coffeaformis v. borealis</i>	-	48 hr	99% reduced growth rate	1,000	Maly and Ruber 1983	Duration
Diatom, <i>Amphora coffeaformis v. borealis</i>	-	48 hr	No significant reduction in growth rate	500	Maly and Ruber 1983	Duration
Diatom, <i>Amphora coffeaformis v. borealis</i>	-	96 hr	69% reduction in growth rate between 48 & 96 hr	10,000	Maly and Ruber 1983	Lack of exposure details
Diatom, <i>Amphora coffeaformis v. borealis</i>	-	96 hr	No significant reduction in growth rate between 48 & 96 hr	2,000	Maly and Ruber 1983	Lack of exposure details
Diatom, <i>Amphiprora sp.</i>	-	48 hr	>100% reduced growth rate	10,000	Maly and Ruber 1983	Duration
Diatom, <i>Amphiprora sp.</i>	-	48 hr	75% reduced growth rate	2,000	Maly and Ruber 1983	Duration
Diatom, <i>Amphiprora sp.</i>	-	48 hr	63% reduced growth rate	1,000	Maly and Ruber 1983	Duration
Diatom, <i>Amphiprora sp.</i>	-	48 hr	No significant reduction in growth rate	500	Maly and Ruber 1983	Duration
Diatom, <i>Amphiprora sp.</i>	-	96 hr	No significant reduction in growth rate between 48 & 96 hr	10,000	Maly and Ruber 1983	Lack of exposure details
Dinoflagellate, <i>Gonyaulax sp.</i>	-	48 hr	No significant reduction in growth rate	10,000	Maly and Ruber 1983	Duration
Dinoflagellate, <i>Gonyaulax sp.</i>	-	96 hr	No significant reduction in growth rate between 48 & 96 hr	10,000	Maly and Ruber 1983	Lack of exposure details
Hermatypic coral, <i>Pocillopora damicornis</i>	Analytical	96 hr	NOEC (survival)	10,000	Acevedo 1991	Duration

Appendix I. Other Data on Effects of Carbaryl to Estuarine/Marine Aquatic Organisms

Species	Chemical	Duration	Effect	Concentration (µg/L)	Reference	Reason Other Data
Polychaete worm, <i>Nereis diversicolor</i>	-	2-8 day	Reduced AChE activity	201.2	Scaps et al. 1997	Atypical endpoint
Lugworm, <i>Arenicola marina</i>	99%	48 hr	LC50	7,200	Conti 1987	Duration
Bay mussel (30-40 mm), <i>Mytilus edulis</i>	98%	72 hr	EC50 (feeding rate)	8,370	Donkin et al. 1997	Atypical endpoint
Bay mussel (embryo), <i>Mytilus edulis</i>	80%	48 hr	EC50 (development)	2,400	Dimick and Breese 1965	Control survival not reported
Blue mussel (unfertilized egg), <i>Mytilus edulis</i>	80%	1 hr	EC50 (development)	20,070	Armstrong and Milleman 1974c	Duration
Blue mussel (1 st polar body), <i>Mytilus edulis</i>	80%	1 hr	EC50 (development)	5,300	Armstrong and Milleman 1974c	Duration
Blue mussel (2 cell), <i>Mytilus edulis</i>	80%	1 hr	EC50 (development)	7,000	Armstrong and Milleman 1974c	Duration
Blue mussel (≥64 cell), <i>Mytilus edulis</i>	80%	1 hr	EC50 (development)	8,300	Armstrong and Milleman 1974c	Duration
Blue mussel (ciliated blastula), <i>Mytilus edulis</i>	80%	1 hr	EC50 (development)	16,000	Armstrong and Milleman 1974c	Duration
Blue mussel (trochophore), <i>Mytilus edulis</i>	80%	1 hr	EC50 (development)	19,000	Armstrong and Milleman 1974c	Duration
Blue mussel (early veliger), <i>Mytilus edulis</i>	80%	1 hr	EC50 (development)	24,000	Armstrong and Milleman 1974c	Duration
Blue mussel, <i>Mytilus edulis</i>	99.7%	10 day	EC50 (shell growth)	>2,610	Liu and Lee 1975	Control issues
Blue mussel, <i>Mytilus edulis</i>	99.7%	20 day	EC50 (shell growth)	>1,300	Liu and Lee 1975	Control issues
Blue mussel, <i>Mytilus edulis</i>	99.7%	20 day	EC50 (metamorphosis with dissoconch shell)	>2,900	Liu and Lee 1975	Control issues
Blue mussel, <i>Mytilus edulis</i>	99.7%	40 day	LC50	>2,900	Liu and Lee 1975	Control issues
Blue mussel, <i>Mytilus edulis</i>	99.7%	40 day	EC50 (shell growth)	>2,900	Liu and Lee 1975	Control issues
Blue mussel, <i>Mytilus edulis</i>	99.7%	20 day	Chronic Value (growth)	463.1	Liu and Lee 1975	Control issues
Blue mussel, <i>Mytilus edulis</i>	-	24 hr	EC50 (# of byssal threads)	>30,000	Roberts 1975	Atypical endpoint
Blue mussel, <i>Mytilus edulis</i>	-	48 hr	EC50 (# of byssal threads)	>30,000	Roberts 1975	Atypical endpoint
Eastern oyster, <i>Crassostrea virginica</i>	>95%	24 hr	LOEC (growth)	1,000	Butler et al. 1960	Duration

Appendix I. Other Data on Effects of Carbaryl to Estuarine/Marine Aquatic Organisms

Species	Chemical	Duration	Effect	Concentration (µg/L)	Reference	Reason Other Data
Eastern oyster (fertilized egg), <i>Crassostrea virginica</i>	-	48 hr	LOEC (development)	1,000	Davis 1961	Lack of exposure details
Eastern oyster, <i>Crassostrea virginica</i>	-	12 day	LC50 (larvae)	3,000	Davis and Hidu 1969	Duration
Eastern oyster, <i>Crassostrea virginica</i>	-	96 hr	19% decrease in shell growth	2,000	Butler 1963	Lack of exposure details
Eastern oyster, <i>Crassostrea virginica</i>	-	96 hr	14% decrease in shell growth	2,000	Butler 1963	Lack of exposure details
Cockle clam (adult), <i>Clinocardium nuttallii</i>	80%	24 hr	EC50	7,300	Stewart et al. 1967	Duration
Hard clam, <i>Mercenaria mercenaria</i>	-	14 day	LC50 (larvae)	>2,500	Davis and Hidu 1969	Lack of exposure details
Quahog clam (fertilized egg), <i>Mercenaria mercenaria</i>	-	48 hr	LOEC (development)	2,500	Davis 1961	Lack of exposure details
Amphipod, <i>Corophium acherusicum</i>	99.5%	10 wk	48% reduction in # of individuals	1.1	Tagatz et al. 1979	Community exposure
Amphipod, <i>Corophium acherusicum</i>	99.5%	10 wk	100% reduction in # of individuals	11.1	Tagatz et al. 1979	Community exposure
Amphipod, <i>Corophium acherusicum</i>	99.5%	10 wk	100% reduction in # of individuals	103	Tagatz et al. 1979	Community exposure
Mysid shrimp (juvenile), <i>Americamysis bahia</i>	99%	72 hr	LC50	8.45	Thursby and Champlin 1991b	96 hr value used
Mysid shrimp (juvenile), <i>Americamysis bahia</i>	99%	24 hr	LC50	21	Thursby and Champlin 1991a	96 hr value used
Mysid shrimp (juvenile), <i>Americamysis bahia</i>	99%	48 hr	LC50	19	Thursby and Champlin 1991a	96 hr value used
Mysid shrimp (juvenile), <i>Americamysis bahia</i>	99%	72 hr	LC50	19	Thursby and Champlin 1991a	96 hr value used
Grass shrimp, <i>Palaemonetes pugio</i>	99%	24 hr	LC50	76	Thursby and Champlin 1991a	Duration
Grass shrimp, <i>Palaemonetes pugio</i>	99%	72 hr	LC50 (fed)	22	Thursby and Champlin 1991a	Test species fed
Grass shrimp, <i>Palaemonetes pugio</i>	99%	96 hr	LC50 (fed)	22	Thursby and Champlin 1991a	Test species fed
Grass shrimp, <i>Palaemonetes pugio</i>	98%	1.5 hr	NOEC (contaminant avoidance)	100	Hansen et al. 1973	Atypical endpoint

Appendix I. Other Data on Effects of Carbaryl to Estuarine/Marine Aquatic Organisms

Species	Chemical	Duration	Effect	Concentration (µg/L)	Reference	Reason Other Data
Grass shrimp, <i>Palaemonetes pugio</i>	98%	1.5 hr	LC50	38	Hansen et al. 1973	Atypical endpoint
Grass shrimp (juvenile), <i>Palaemonetes pugio</i>	99.7%	48 hr	LC50	28	Hansen 1980; Mayer 1987	Duration
Brown shrimp (juvenile), <i>Penaeus aztecus</i>	99.7%	48 hr	LC50	1.5	Hansen 1980; Mayer 1987	Duration
Brown shrimp (adult), <i>Penaeus aztecus</i>	-	48 hr	LC50	2.5	Butler 1963	Duration
Brown shrimp, <i>Penaeus aztecus</i>	-	24 hr	EC50	5.5	Butler 1963	Duration
Pink shrimp (juvenile), <i>Penaeus duorarum</i>	99.7%	48 hr	LC50	32	Hansen 1980; Mayer 1987	Duration
Mud shrimp (larva), <i>Upogebia pugettensis</i>	80%	24 hr	LC50	130	Stewart et al. 1967	Duration
Mud shrimp (larva), <i>Upogebia pugettensis</i>	80%	24 hr	LC50	40	Stewart et al. 1967	Duration
Ghost shrimp (larva), <i>Callinassa californiensis</i>	80%	24 hr	EC50	470	Stewart et al. 1967	Duration
Ghost shrimp (larva), <i>Callinassa californiensis</i>	80%	24 hr	EC50	170	Stewart et al. 1967	Duration
Ghost shrimp (larva), <i>Callinassa californiensis</i>	80%	24 hr	EC50	5,600	Stewart et al. 1967	Duration
Ghost shrimp (adult), <i>Callinassa californiensis</i>	80%	24 hr	EC50	130	Stewart et al. 1967	Duration
Sand shrimp (2.4-4.5 g), <i>Crangon septemspinosa</i>	-	96 hr	Lethal threshold	20	McLeese et al. 1979	Atypical endpoint
American lobster (1 st stage), <i>Homarus americanus</i>	99%	24 hr	LC50	38.73	Champlin and Poucher 1992	Duration
American lobster (1 st stage), <i>Homarus americanus</i>	99%	48 hr	LC50	23.13	Champlin and Poucher 1992	Test species fed
American lobster (1 st stage), <i>Homarus americanus</i>	99%	72 hr	LC50	20.89	Champlin and Poucher 1992	Test species fed
American lobster (1 st stage), <i>Homarus americanus</i>	99%	96 hr	LC50	20.89	Champlin and Poucher 1992	Test species fed
Blue crab (juvenile), <i>Callinectes sapidus</i>	99.7%	48 hr	LC50	320	Hansen 1980; Mayer 1987	Duration

Appendix I. Other Data on Effects of Carbaryl to Estuarine/Marine Aquatic Organisms

Species	Chemical	Duration	Effect	Concentration (µg/L)	Reference	Reason Other Data
Blue crab (juvenile), <i>Callinectes sapidus</i>	-	48 hr	LC50	550	Butler 1963	Duration
Dungeness crab (prezoea), <i>Metacarcinus magister</i> (formerly <i>Cancer magister</i>)	80%	24 hr	NOEC (hatching)	1,000	Buchanan et al. 1970	Duration
Dungeness crab (juvenile male), <i>Metacarcinus magister</i>	80%	24 hr	EC50	600	Stewart et al. 1967	Duration
Dungeness crab (juvenile female), <i>Metacarcinus magister</i>	80%	24 hr	EC50	630	Stewart et al. 1967	Duration
Dungeness crab (egg), <i>Metacarcinus magister</i>	80%	24 hr	NOEC (hatching @ 10°C)	1,000	Buchanan et al. 1970	Duration
Dungeness crab (prezoea), <i>Metacarcinus magister</i>	80%	24 hr	EC50 (molting to zoea @ 10°C)	6	Buchanan et al. 1970	Duration
Dungeness crab (prezoea), <i>Metacarcinus magister</i>	80%	24 hr	EC50 (molting to zoea @ 10°C)	20	Buchanan et al. 1970	Duration
Dungeness crab (prezoea), <i>Metacarcinus magister</i>	80%	24 hr	EC50 (molting to zoea @ 10°C)	30	Buchanan et al. 1970	Duration
Dungeness crab (zoea; 1-2 day), <i>Metacarcinus magister</i>	80%	24 hr	LC50 (17°C)	80	Buchanan et al. 1970	Duration
Dungeness crab (zoeae; 1-2 day), <i>Metacarcinus magister</i>	80%	24 hr	LC50 (17°C)	5	Buchanan et al. 1970	Duration
Dungeness crab (zoeae), <i>Metacarcinus magister</i>	80%	24 hr	EC50 (cessation of swimming) (10°C)	6.5	Buchanan et al. 1970	Duration
Dungeness crab (zoea; 1 day), <i>Metacarcinus magister</i>	80%	24 hr	EC50 (10°C)	>1,000	Buchanan et al. 1970	Duration
Dungeness crab (juvenile), <i>Metacarcinus magister</i>	80%	48 hr	EC50 (14°C)	76	Buchanan et al. 1970	Duration

Appendix I. Other Data on Effects of Carbaryl to Estuarine/Marine Aquatic Organisms

Species	Chemical	Duration	Effect	Concentration (µg/L)	Reference	Reason Other Data
Dungeness crab (juvenile), <i>Metacarcinus magister</i>	80%	48 hr	EC50 (12°C)	57	Buchanan et al. 1970	Duration
Dungeness crab (9 th stage; juvenile), <i>Metacarcinus magister</i>	80%	24 hr	EC50 (18°C)	350	Buchanan et al. 1970	Duration
Dungeness crab (9 th stage; juvenile), <i>Metacarcinus magister</i>	80%	24 hr	EC50 (18°C)	320	Buchanan et al. 1970	Duration
Dungeness crab (9 th stage; juvenile), <i>Metacarcinus magister</i>	80%	48 hr	EC50 (18°C)	620	Buchanan et al. 1970	Duration
Dungeness crab (9 th stage; juvenile), <i>Metacarcinus magister</i>	80%	48 hr	EC50 (18°C)	220	Buchanan et al. 1970	Duration
Dungeness crab (adult), <i>Metacarcinus magister</i>	80%	24 hr	EC50 (11°C)	320	Buchanan et al. 1970	Duration
Dungeness crab (adult), <i>Metacarcinus magister</i>	80%	24 hr	EC50 (18°C)	490	Buchanan et al. 1970	Duration
Shore crab (adult male), <i>Hemigrapsus oregonensis</i>	80%	24 hr	EC50	710	Stewart et al. 1967	Duration
Shore crab (adult female), <i>Hemigrapsus oregonensis</i>	80%	24 hr	EC50	270	Stewart et al. 1967	Duration
Sand dollar, <i>Echinarachnius parma</i>	-	72 hr	NOEC (development)	10,000	Crawford and Guarino 1976	Duration
Sea urchin (embryo/larva), <i>Arbacia punctulata</i>	99%	48 hr	LC50	4,700	Thursby and Champlin 1991a	Duration
Sheepshead minnow, <i>Cyprinodon variegatus</i>	99%	24 hr	LC50	6,300	Thursby and Champlin 1991a	Duration
Sheepshead minnow, <i>Cyprinodon variegatus</i>	99%	48 hr	LC50	6,300	Thursby and Champlin 1991a	Duration
Sheepshead minnow, <i>Cyprinodon variegatus</i>	99%	72 hr	LC50	6,000	Thursby and Champlin 1991a	Duration

Appendix I. Other Data on Effects of Carbaryl to Estuarine/Marine Aquatic Organisms

Species	Chemical	Duration	Effect	Concentration (µg/L)	Reference	Reason Other Data
Sheepshead minnow, <i>Cyprinodon variegatus</i>	99%	96 hr	LC50 (fed)	5,800	Thursby and Champlin 1991a	Test species fed
Sheepshead minnow, <i>Cyprinodon variegatus</i>	98%	24 hr	LC50	2,800	Hansen 1969; 1970	Duration
Sheepshead minnow, <i>Cyprinodon variegatus</i>	98%	1.5 hr	No avoidance	100	Hansen 1969; 1970	Duration
Longnose killifish (juvenile), <i>Fundulus similis</i>	99.7%	48 hr	EC50 (mortality)	1,600	Hansen 1980; Mayer 1987	Duration
Longnose killifish, <i>Fundulus similis</i>	-	24 hr	LC50	1,750	Butler 1963	Duration
Longnose killifish, <i>Fundulus similis</i>	-	48 hr	LC50	1,750	Butler 1963	Duration
Killifish (embryo), <i>Fundulus heteroclitus</i>	-	40 day	LOEC (fry development)	10	Crawford and Guarino 1985	Duration
Killifish, <i>Fundulus heteroclitus</i>	99%	10 day	LC50	>10,000	Shea and Berry 1983	Duration
Mummichog, <i>Fundulus heteroclitus</i>	99.1%	10 day	LC40	10,000	Shea and Berry 1983	Duration
Killifish, (8-16 cell stage), <i>Fundulus heteroclitus</i>	99.2%	72 hr	NOEC (development arrest)	100	Weis and Weis 1974a	Duration
Atlantic silverside (2-3 wk), <i>Menidia beryllina</i>	99%	24 hr	LC50	1,800	Thursby and Champlin 1991a	Duration
Atlantic silverside (2-3 wk), <i>Menidia beryllina</i>	99%	48 hr	LC50	1,700	Thursby and Champlin 1991a	Duration
Atlantic silverside (2-3 wk), <i>Menidia beryllina</i>	99%	72 hr	LC50	1,600	Thursby and Champlin 1991a	Duration
Atlantic silverside (2-3 wk), <i>Menidia beryllina</i>	99%	96 hr	LC50 (fed)	1,600	Thursby and Champlin 1991a	Test species fed
Atlantic silverside (juvenile), <i>Menidia beryllina</i>	99.2%	24 hr	Disruption in schooling behavior	100	Weis and Weis 1974b	Duration
Threespine stickleback (juvenile), <i>Gasterosteus aculeatus</i>	80%	24 hr	EC50	6,700	Stewart et al. 1967	Duration
Threespine stickleback, <i>Gasterosteus aculeatus</i>	95%	48 hr	LC50	10,450	Katz 1961	Duration

Appendix I. Other Data on Effects of Carbaryl to Estuarine/Marine Aquatic Organisms

Species	Chemical	Duration	Effect	Concentration (µg/L)	Reference	Reason Other Data
Threespine stickleback, <i>Gasterosteus aculeatus</i>	95%	72 hr	LC50	4,940	Katz 1961	Duration
Threespine stickleback, <i>Gasterosteus aculeatus</i>	95%	48 hr	LC50	16,625	Katz 1961	Duration
Threespine stickleback, <i>Gasterosteus aculeatus</i>	95%	72 hr	LC50	6,175	Katz 1961	Duration
Striped bass (31 mm; 0.42 g), <i>Morone saxatilis</i>	98%	96 hr	LC50	1,000	Korn and Earnest 1974	Control survival not reported
Shiner perch (juvenile), <i>Cymatogaster aggregate</i>	80%	24 hr	EC50	3,900	Stewart et al. 1967	Duration
Mullet (juvenile), <i>Mugil cephalus</i>	99.7%	48 hr	EC50 (mortality)	2,400	Hansen 1980; Mayer 1987	Duration
White mullet (juvenile), <i>Mugil curema</i>	-	48 hr	LC50	2,500	Butler 1963	Duration
English sole (juvenile), <i>Parophrys vetulus</i>	80%	24 hr	EC50	4,100	Stewart et al. 1967	Duration

Dash indicates not available

Appendix J

Appendix J. List of Carbaryl Studies Not Used in Document Along with Reasons

Author	Title	Date	Organism(s)	Concentration (µg/L)	Reason Unused	Expanded Reason
Abbasi and Soni	Studies on the environmental impact of three common pesticides with respect to toxicity towards a larvivore (channelfish <i>N. denricus</i>).	1991	Channelfish, <i>Nuria denricus</i>	96 hr LC50=34,670	Not North American species	
Adhikary	Effect of pesticides on the growth, photosynthetic oxygen evolution and nitrogen fixation of <i>Westiellopsis prolifica</i> .	1989	Cyanobacteria, <i>Westiellopsis prolifica</i>	-	Formulation	Sevin (50% carbaryl)
Adhikary et al.	Effect of the carbamate insecticide sevin® on <i>Anabaena sp.</i> and <i>Westiellopsis prolifica</i> .	1984	Cyanobacteria, <i>Anabaena sp.</i> <i>Westiellopsis prolifica</i>	-	Formulation	Sevin (50% carbaryl)
Aggarwal et al.	Effect of some carbamate pesticides on nodulation, plant yield and nitrogen fixation by <i>Pisum sativum</i> and <i>Vigna sinensis</i> in the presence of their rhizobia.	1986	Garden pea, <i>Pisum sativum</i> Cow pea, <i>Vigna sinensis</i>	-	Sediment exposure	
Agrawal	The accumulation of biocide residues in a few tissues of <i>Lamellidens marginalis</i> .	1986	Bivalve, <i>Lamellidens marginalis</i>	-	Bioaccumulation: steady state not reached	Static, unmeasured study
Almar et al.	Influence of temperature on several pesticides toxicity to <i>Melanopsis dufouri</i> under laboratory conditions.	1988	Snail, <i>Melanopsis dufouri</i>	96 hr LC50=10,100	Not North American species	
Anbu and Ramaswamy.	Adaptive changes in respiratory movements of an air-breathing fish, <i>Channa striatus</i> (Bleeker) exposed to carbamate pesticide, sevin.	1991	Snakehead catfish, <i>Channa striatus</i>	-	Formulation	Sevin (10% dust)
Ansara-Ross et al.	Probabilistic risk assessment of the environmental impacts of pesticides in the Crocodile (west) Marico catchment, North-West Province.	2008	-	-	Data modeling	
Areekul	Toxicity to fishes of insecticides used in paddy fields and water resources. I. Laboratory experiment.	1986	-	-	Lack of exposure details	Text in foreign language
Ariaratnam and Georghiou	Carbamate resistance in <i>Anopheles albimanus</i> .	1975	Mosquito, <i>Anopheles albimanus</i>	-	Not North American species	Metabolism study
Armstrong and Millemann	Effects of the insecticide carbaryl on clams and some other intertidal mud flat animals.	1974a	-	-	Too few exposure concentrations (<3)	Community field exposure
Arora and Kulshrestha	Comparison of the toxic effects of two pesticides on the testes of a fresh water teleost <i>Channa striatus</i> Bloch.	1984	Snakehead catfish, <i>Channa striatus</i>	-	Not North American species; Formulation	Sevin (50 WP)
Arora and Kulshrestha	Effects of chronic exposure to sublethal doses of two pesticides on alkaline and acid phosphatase activities in the intestine of a fresh water Teleost, <i>Channa striatus</i> Bloch. (Channidae).	1985	Snakehead catfish, <i>Channa striatus</i>	-	Not North American species; Formulation	Sevin (50 WP)

Appendix J. List of Carbaryl Studies Not Used in Document Along with Reasons

Author	Title	Date	Organism(s)	Concentration (µg/L)	Reason Unused	Expanded Reason
Arunachalam and Palanichamy	Sublethal effects of carbaryl on surfacing behaviour and food utilization in the air-breathing fish, <i>Macropodus cupanus</i> .	1982	Paradise fish, <i>Macropodus cupanus</i>	-	Not North American species; Formulation	Carbaryl (50 WP)
Arunachalam et al.	Toxic and sublethal effects of carbaryl on a freshwater catfish, <i>Mystus vittatus</i> (Bloch).	1980	Catfish, <i>Mystus vittatus</i>	-	Not North American species; Formulation	Carbaryl (50 WP)
Arunachalam et al.	Effect of carbaryl on esterases in the air-breathing fish <i>Channa punctatus</i> (Bloch).	1985a	Snakehead catfish, <i>Channa punctatus</i>	-	Not North American species; Only one exposure concentration	
Arunachalam et al.	Sublethal effects of carbaryl on food utilization and oxygen consumption in the air-breathing fish, <i>Channa punctatus</i> (Bloch).	1985b	Snakehead catfish, <i>Channa punctatus</i>	96 hr LC50=6,000	Not North American species	
Arunachalam et al.	The impact of pesticides on the feeding energetics and body composition in the freshwater catfish, <i>Mystus vittatus</i> .	1990	Catfish, <i>Mystus vittatus</i>	-	Not North American species; Formulation	Carbaryl (50 EC)
Ashauer et al.	Simulating toxicity of carbaryl to <i>Gammarus pulex</i> after sequential pulsed exposure.	2007a	Amphipod, <i>Gammarus pulex</i>	-	Not North American species	Pulsed exposure
Ashauer et al.	Modeling combined effects of pulsed exposure to carbaryl and chlorpyrifos on <i>Gammarus pulex</i> .	2007b	Amphipod, <i>Gammarus pulex</i>	-	Not North American species; Mixture; Pulsed exposure	Carbaryl and chlorpyrifos
Atallah and Ishak	Toxicity of some commonly used insecticides to the snail <i>Biomphalaria alexandria</i> , intermediate host of <i>Schistosoma mansoni</i> in Egypt.	1971	Snail, <i>Biomphalaria alexandria</i>	24 hr LC50=47,000	Not North American species	
Bajpai and Perti	Resistance to malathion.	1969	Mosquito, <i>Culex fatigans</i>	-	Prior exposure	Malathion resistant strain
Bakr et al.	Insect growth regulators: I. Biological activity of some IGR's against the susceptible and resistant strains of <i>Culex pipiens</i> larvae: II. Pattern of cross resistance to IGR's in carbaryl-resistant strain.	1989	Mosquito, <i>Culex pipiens</i>	-	Prior exposure	Carbaryl-resistant strain
Balasubramanian and Ramaswami	Effect of pesticide sevin on acetylcholinesterase (AChE) activity in different tissues of <i>Oreochromis mossambicus</i> (Peters).	1991	Mozambique tilapia, <i>Oreochromis mossambicus</i>	24 hr decrease AChE activity in brain and heart at 3,000	Only two exposure concentrations	
Bansal et al.	Pesticide-induced alterations in the oxygen uptake rate of a freshwater major carp <i>Labeo rohita</i> .	1979	Carp, <i>Labeo rohita</i>	-	Not North American species; Formulation	Sevin (50% WP)
Bansal et al.	Predicting long-term toxicity by subacute screening of pesticides with larvae and early juveniles of four species of freshwater major carp.	1980	Carp, <i>Labeo rohita</i> , <i>Cirrhina mrigala</i> , <i>Catla catla</i> , <i>Cyprinus carpio</i>	-	Formulation	Sevin (50% WP)

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Author	Title	Date	Organism(s)	Concentration (µg/L)	Reason Unused	Expanded Reason
Barahona and Sanchez-Fortun.	Toxicity of carbamates to the brine shrimp <i>Artemia salina</i> and the effect of atropine, BW284c51, iso-OMPA and 2-PAM on carbaryl toxicity.	1999	Brine shrimp, <i>Artemia salina</i>		Brine shrimp	
Barker et al.	Relationship between <i>Legionella pneumophila</i> and <i>Acanthamoeba polyphaga</i> : physiological status and susceptibility to chemical inactivation.	1992	-	-	Not applicable	No carbaryl toxicity information
Barry	The effects of a pesticide on inducible phenotypic plasticity in <i>Daphnia</i> .	1999	Cladoceran, <i>Daphnia longicephala</i>	-	Formulation	Yates Carbaryl®
Basak and Konar	Effects of an organophosphorus insecticide, dimethoate, on the survival, behavior, growth and reproduction of fish.	1975	-	-	Not applicable	No carbaryl toxicity information
Basak and Konar	Toxicity of six insecticides to fish.	1976a	Carp, <i>Cyprinus carpio</i> Mozambique tilapia, <i>Oreochromis mossambicus</i> Catfish, <i>Heteropneustes fossilis</i>	-	Formulation	Sevin (50% WP)
Basak and Konar	Pollution of water by pesticides and protection of fishes: parathion.	1976b	Carp, <i>Cyprinus carpio</i> Mozambique tilapia, <i>Oreochromis mossambicus</i> Catfish, <i>Heteropneustes fossilis</i>	-	Formulation	Sevin (50% WP)
Basha et al.	Respiratory potentials of the fish (<i>Tilapia mossambica</i>) under malathion, carbaryl and lindane intoxication.	1984	Mozambique tilapia, <i>Oreochromis mossambicus</i>	48 hr decreased oxygen consumption at 1,832	Only one exposure concentration	
Basol et al.	Comparative toxicity of some pesticides on human health and some aquatic species.	1980	Electric eel	-	Altered test species; No scientific name given	
Basso et al.	Alterations of <i>Aplysia</i> feeding behavior following acute carbamate intoxication.	1986	Marine snails, <i>Aplysia depilans</i> , <i>Aplysia punctata</i> , <i>Aplysia fasciata</i>	-	Formulation	AS 50, Sipcam (47.5% carbaryl)
Beauvais et al.	Cholinergic and behavioral neurotoxicity of carbaryl and cadmium to larval rainbow trout (<i>Oncorhynchus mykiss</i>).	2001	Rainbow trout, <i>Oncorhynchus mykiss</i>	-	Excessive solvent	0.75 mL/L acetone
Beyers et al.	Effects of rangeland aerial application of sevin-4-oil® on fish and aquatic invertebrate drift in Little Missouri River, North Dakota.	1995	-	-	Formulation	Sevin-4-oil®
Bhatia	Toxicity of some pesticides to <i>Puntius ticto</i> (Hamilton).	1971a	Ticto barb, <i>Puntius ticto</i>	96 hr TLm=70,000	Not North American species	

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Author	Title	Date	Organism(s)	Concentration (µg/L)	Reason Unused	Expanded Reason
Bhatia	Toxicity of some insecticides to <i>Cirrhinus mrigala</i> (Hamilton) and <i>Colisa fasciata</i> (Bloch & Schneider).	1971b	Mrigal, <i>Cirrhinus mrigala</i> Kholisa, <i>Colisa fasciata</i>	96 hr LC50=3,168 LC50=1,237	Not North American species	
Bhattacharya	Target and non-target effects of anticholinesterase pesticides in fish.	1993	Snakehead catfish, <i>Channa punctatus</i>	-	Formulation	50% carbaryl
Bhattacharya	Stress response to pesticides and heavy metals in fish and other vertebrates.	2001	-	-	Review of previous studies	
Bhavan and Geraldine	Carbaryl-induced alterations in biochemical metabolism of the prawn, <i>Macrobrachium malcolmsonii</i> .	2002	Prawn, <i>Macrobrachium malcolmsonii</i>	-	Not North American species; Formulation	Sevin (50% carbaryl)
Bhunja et al.	Carbaryl-induced effects of glutathione content, glutathione reductase and superoxide dismutase activity of the cyanobacterium <i>Nostoc muscorum</i> .	1993	Cyanobacterium, <i>Nostoc muscorum</i>	-	Altered test species	Excised tissue
Bhunja and Sahoo	Genotoxic potential of carbaryl in the peripheral blood erythrocytes of <i>Anabas testudineus</i> .	2004	Climbing perch, <i>Anabas testudineus</i>	-	Injected toxicant; Formulation	Sevin (50% carbaryl)
Bielecki	The effect of phoschlorine, carbatox and copper sulphate on the development of eggs and hatching of miracidia in <i>Fasciola hepatica</i> L.	1987	Liver fluke, <i>Fasciola hepatica</i>	3 hr inhibit egg development at 3,000,000	Only one exposure concentration	
Binelli et al.	New evidence for old biomarkers: effects of several xenobiotics on EROD and AChE activities in zebra mussel (<i>Dreissena polymorpha</i>).	2006	Zebra mussel, <i>Dreissena polymorpha</i>	96 hr Decrease AChE activity at 100,000	Only one exposure concentration	
Bluzat and Seuge	Effects of three insecticides (lindane, fenthion, and carbaryl) on the acute toxicity to four aquatic invertebrate species and the chronic toxicity.	1979	Amphipod, <i>Gammarus pulex</i> Mayfly, <i>Cleon sp.</i> Midge, <i>Chaoborus sp.</i> Snail, <i>Lymnaea stagnalis</i>	48 hr LC50=29 LC50=480 LC50=296 LC50=21,000	Excessive solvent used; Text in foreign language	1% solvent
Bogacka and Groba	Toxicity and biodegradation of chlorfenvinphos, carbaryl and propoxur in water environment.	1980	Cladoceran, <i>Daphnia magna</i>	48 hr LC50=1	Lack of detail; text in foreign language	
Bogaerts et al.	Use of ciliated protozoan <i>Tetrahymena pyriformis</i> for the assessment of toxicity and quantitative structure-activity relationships of xenobiotics: comparison with the microtox test.	2001	Protozoan, <i>Tetrahymena pyriformis</i>	-	Excessive solvent used	DMSO (0.12%)
Bonning and Hemingway	The efficacy of acetylcholinesterase in organophosphorus and carbamate resistance in <i>Culex pipiens</i> L. from Italy.	1991	Mosquito, <i>Culex pipiens</i>	-	Prior exposure	Carbaryl resistant strain

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Author	Title	Date	Organism(s)	Concentration (µg/L)	Reason Unused	Expanded Reason
Boone	Examining the single and interactive effects of three insecticides on amphibian metamorphosis.	2008	American toad, <i>Bufo americanus</i> Green frog, <i>Rana clamitans</i>	-	Formulation; Only two exposure concentrations	Sevin (22.5% carbaryl)
Boone and Bridges	Effects of carbaryl on green frog (<i>Rana clamitans</i>) tadpoles: timing of exposure versus multiple exposures.	2003	Green frog, <i>Rana clamitans</i>	-	Pulsed exposure	
Boone and James	Interactions of an insecticide, herbicide, and natural stressors in amphibian community mesocosms.	2003	American toad, <i>Bufo americanus</i> Southern leopard frog, <i>Rana sphenocephala</i> Spotted salamander, <i>Ambystoma maculatum</i> Small-mouth salamander, <i>A. texanum</i>	-	Only two exposure concentrations; Formulation	Sevin (21.3% carbaryl)
Boone and Semlitsch	Interactions of an insecticide with larval density and predation in experimental amphibian communities.	2001	Woodhouse toad, <i>Bufo woodhouse</i> Gray treefrog, <i>Hyla versicolor</i> Green frog, <i>Rana clamitans</i>	-	Formulation	Sevin (21.3% carbaryl)
Boone and Semlitsch	Interactions of bullfrog tadpole predators and an insecticide: predation release and facilitation.	2003	Bullfrog, <i>Rana catesbeiana</i> Red-spotted newt, <i>Notophthalmus viridescens</i> Bluegill, <i>Lepomis macrochirus</i> Crayfish, <i>Orconectes sp.</i>	-	Only two exposure concentrations; Formulation	Sevin (21.3% carbaryl)
Boone et al.	Growth and development of larval green frogs (<i>Rana clamitans</i>) exposed to multiple doses of an insecticide.	2001	Green frog, <i>Rana clamitans</i>	-	Formulation	Sevin (21.3% carbaryl)
Boone et al.	Effects of an insecticide on amphibians in large-scale experimental ponds.	2004	Woodhouse toad, <i>Bufo woodhouse</i> Southern leopard frog, <i>Rana sphenocephala</i>	-	Formulation	Sevin (21.3% carbaryl)
Boone et al.	Multiple sublethal chemicals negatively affect tadpoles of the green frog, <i>Rana clamitans</i> .	2005	Green frog, <i>Rana clamitans</i>	-	Only one exposure concentration; Formulation	Sevin (22.5% carbaryl)

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Author	Title	Date	Organism(s)	Concentration (µg/L)	Reason Unused	Expanded Reason
Boone et al.	Multiple stressors in amphibian communities: Effects of chemical contamination, bullfrogs, and fish.	2007	American toad, <i>Bufo americanus</i> Southern leopard frog, <i>Rana sphenocephala</i> Bullfrog, <i>Rana catesbeiana</i> Spotted salamander, <i>Ambystoma maculatum</i> Bluegill, <i>Lepomis macrochirus</i>	-	Formulation	Sevin (22.5% carbaryl)
Boran et al.	Acute toxicity of carbaryl, methiocarb and carbosulfan to the rainbow trout (<i>Oncorhynchus mykiss</i>) and guppy (<i>Poecilia reticulata</i>).	2007	Rainbow trout, <i>Oncorhynchus mykiss</i> Guppy, <i>Poecilia reticulata</i>	96 hr LC50=522 LC50=1,383	Excessive solvent used	0.2%
Bradbury et al.	Use of respiratory-cardiovascular responses of rainbow trout (<i>Oncorhynchus mykiss</i>) in identifying acute toxicity syndromes in fish: part 4. Central nervous system seizure agents.	1991	Rainbow trout, <i>Oncorhynchus mykiss</i>	-	Too few test organisms; Surgically altered test species	Spinally transected trout
Bridges	Effects of a pesticide on tadpole activity and predatory avoidance behavior.	1999a	Gray treefrog, <i>Hyla versicolor</i>	24 hr decrease tadpole activity at 2,500	Only two exposure concentrations	
Bridges	Predator-prey interactions between two amphibian species: effects of insecticide exposure.	1999b	Southern leopard frog, <i>Rana sphenocephala</i> Red-spotted newt, <i>Notophthalmus viridescens</i>	1 hr decrease newt consumption of tadpoles at 2,500	Only one exposure concentration	
Bridges	Long-term effects of pesticide exposure at various life stages of the southern leopard frog (<i>Rana sphenocephala</i>).	2000	Southern leopard frog, <i>Rana sphenocephala</i>	-	Unmeasured chronic exposure	
Bridges and Boone	The interactive effects of UV-B and insecticide exposure on tadpole survival, growth and development.	2003	Southern leopard frog, <i>Rana sphenocephala</i>	-	Formulation	Sevin ® (22.5% carbaryl)
Bridges and Semlitsch	Variation in pesticide tolerance of tadpoles among and within species of ranidae and patterns of amphibian decline.	2000	-	-	Only one exposure concentration	
Bridges and Semlitsch	Genetic variation in insecticide tolerance in a population of southern leopard frogs (<i>Rana sphenocephala</i>): implications for amphibian conservation	2001	Southern leopard frog, <i>Rana sphenocephala</i>	-	Only one exposure concentration	
Bridges et al.	Comparative contaminant toxicity: are amphibian larvae more sensitive than fish?	2002	-	-	Review of previous studies	

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Author	Title	Date	Organism(s)	Concentration (µg/L)	Reason Unused	Expanded Reason
Brown	Effects of split applications of sevin-4-oil® on aquatic invertebrate drift.	1980	-	-	Formulation	Sevin-4-oil®
Bruner and Fisher	The effects of temperature, pH, and sediment on the fate and toxicity of 1-naphthol to the midge larvae <i>Chironomus riparius</i> .	1993	Midge, <i>Chironomus riparius</i>	-	Not applicable	No carbaryl toxicity information
Burdick et al.	Effect of sevin upon the aquatic environment.	1960	-	-	Formulation	Sevin
Burdick et al.	Toxicity of sevin (carbaryl) to fingerling brown trout.	1965	Brown trout	-	Lack of detail; Formulation	Sevin; Species name not given
Butler	Pesticide-wildlife studies, 1964. A review of Fish and Wildlife Service investigations during the calendar year.	1964	Marine shrimp and drills	-	Formulation	Sevin (10%, 3% and 2%)
Cajaraville et al.	Acute toxicity of two hydroxylated hydrocarbons to the prosobranch gastropod <i>Littorina littorea</i> .	1989a	Snail, <i>Littorina littorea</i>	-	Not applicable	No carbaryl toxicity information
Cajaraville et al.	A stereological survey of lysosomal structure alterations in <i>Littorina littorea</i> exposed to 1-naphthol.	1989b	Snail, <i>Littorina littorea</i>	-	Not applicable	No carbaryl toxicity information
Cajaraville et al.	Short-term toxic effects of 1-naphthol on the digestive gland-gonad complex of the marine prosobranch <i>Littorina littorea</i> (L): a light microscopic study.	1990	Snail, <i>Littorina littorea</i>	-	Not applicable	No carbaryl toxicity information
Calapaj	Chemical pollution of Mytilus. I. Radiostrontium, radiocesium, inorganic mercury, hexavalent chromium.	1973	-	-	Lack of detail	Text in foreign language
Campero et al.	Ecological relevance and sensitivity depending on the exposure time for two biomarkers.	2007a	Damselfly, <i>Coenagrion puella</i>	7d Growth NOEC=5 LOEC=40	Not North American species	
Campero et al.	Sublethal pesticide concentrations and predation jointly shape life history: Behavioral and physiological mechanisms.	2007b	Damselfly, <i>Coenagrion puella</i>	-	Not North American species	Predator present in exposure medium
Capaldo	Effects of carbaryl (SEVIN) on the stage I zoeae of the red-jointed fiddler crab, <i>Uca minax</i> (LeConte).	1987	Red-jointed fiddler crab, <i>Uca minax</i>	-	Formulation	Sevin (27% carbaryl)
Carlson et al.	Neurological effects on startle response and escape from predation by medaka exposed to organic chemicals.	1998	Medaka, <i>Oryzias latipes</i>	48 hr LC50=9,400	Not North American species	
Carter	'In vivo' studies of brain acetylcholinesterase inhibition by organophosphate and carbamate insecticides in fish.	1971	-	-	Excessive solvent	
Carter and Graves	Measuring effects of insecticides on aquatic animals.	1972	-	-	Lack of exposure details	No scientific names given

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Author	Title	Date	Organism(s)	Concentration (µg/L)	Reason Unused	Expanded Reason
Chaiyarach et al.	Acute toxicity of the insecticides toxaphene and carbaryl and the herbicides propanil and molinate to four species of aquatic organisms.	1975	Mosquitofish, <i>Gambusia affinis</i> Grass shrimp, <i>Palaemonetes kadiakensis</i> Crayfish, <i>Procambarus simulans</i> Clam, <i>Rangia cuneata</i>	96 hr LC50=31,800 LC50=120 LC50=2,430 LC50=125,000	Prior exposure	
Chakrawarti and Chaurasia	Toxicity of some organophosphate, chlorinated and carbamate pesticides to some fresh water fishes.	1981	Walking catfish, <i>Clarias batrachus</i> Catfish, <i>Heteropneustes fossilis</i>	96 hr LC50=>1,200,000 for both species	Lack of exposure details	Dilution water not characterized
Chambers	Investigation of chemical control of ghost shrimp on oyster grounds 1960-1963.	1969	Ghost shrimp, <i>Callinassa californiensis</i>	-	Formulation	Sevin
Chandran et al.	Method for the estimation of safe experimental concentration for intermediate toxicity test.	1991	Catfish, <i>Mystus tengara</i> Catfish, <i>Heteropneustes fossilis</i> Catfish, <i>Anabas testudineus</i>	96 hr LC50=18,000 LC50=58,000 LC50=20,500	Not North American species	
Chang et al.	Impact of pesticide application on zooplankton communities with different densities of invertebrate predators: an experimental analysis using small-scale mesocosms.	2005	-	-	Only one exposure concentration	
Chari	A rapid bioassay procedure to determine the toxicity of pesticides to <i>Channa punctatus</i> Bloch.	1992	Snakehead catfish, <i>Channa punctatus</i>	-	Formulation	Sevin
Cheah et al.	Some effects of rice pesticides on crawfish.	1980a	Crayfish, <i>Procambarus clarkii</i>	96 hr LC50=500	Lack of exposure details	
Cheah et al.	Acute toxicity of selected rice pesticides to crayfish, <i>Procambarus clarkii</i> .	1980b	Crayfish, <i>Procambarus clarkii</i>	-	Formulation	Sevin
Chen et al.	Laboratory studies on the susceptibility of mosquito-eating fish, <i>Lebistes reticulatus</i> and the larvae of <i>Culex pipiens fatigans</i> to insecticides.	1971	Guppy, <i>Lebistes reticulatus</i> Mosquito, <i>Culex pipiens fatigans</i>	24 hr LC50=2,600 LC50=400	Excessive solvent used; Lack of exposure details	0.4%; Dilution water not characterized
Chen and Sudderuddin	Toxicological studies of insecticides on <i>Culex quinquefasciatus</i> Say and <i>Aedes aegypti</i> (L.).	1978	Mosquito, <i>Culex quinquefasciatus</i> Mosquito, <i>Aedes aegypti</i>	24 hr LC50=680 LC50=690	Lack of exposure details	Dilution water not characterized
Chitra and Pillai	Development of organophosphorus and carbamate-resistance in Indian strains of <i>Anopheles stephensi</i> Liston.	1984	Mosquito, <i>Anopheles stephensi</i>	-	Excessive solvent	4 mL/L ethanol

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Author	Title	Date	Organism(s)	Concentration (µg/L)	Reason Unused	Expanded Reason
Choudhury et al.	Non lethal concentrations of pesticide impair ovarian function in the freshwater perch, <i>Anabas testudineus</i> .	1993	Perch, <i>Anabas testudineus</i>	-	Not North American species; Only one exposure concentration	
Christensen and Tucker	Effects of selected water toxicants on the in vitro activity of fish carbonic anhydrase.	1976	Channel catfish, <i>Ictalurus punctatus</i>	-	Surgically altered test species	Red blood cells
Christoffers and Ernst	The in-vivo fluorescence of <i>Chlorella fusca</i> as a biological test for the inhibition of photosynthesis.	1983	Green alga, <i>Chlorella fusca</i>	-	Only one exposure concentration; Excessive solvent	1% acetone
Chung et al.	Toxicity of carbaryl, diquat dibromide, and fluoranthene, individually and in mixture, to larval grass shrimp, <i>Palaemonetes pugio</i> .	2008	Grass shrimp, <i>Palaemonetes pugio</i>	96 hr LC50=43.02	Excessive solvent used	0.1%
Cocks	The effect of aldrin on water balance in the freshwater pulmonate gastropod (<i>Biomphalaria glabrata</i>).	1973	Snail, <i>Biomphalaria glabrata</i>	24 hr heart beat 31% of control at 64,390	Not North American species	
Connors and Black	Evaluation of lethality and genotoxicity in the freshwater mussel <i>Utterbackia imbecillis</i> (Bivalvia: Unionidae) exposed singly and in combination to chemicals used in lawn care.	2004	Mussel, <i>Utterbackia imbecillis</i>	-	Formulation	Sevin (22.5% carbaryl)
Cook et al.	Succession of microfungi in estuarine microcosms perturbed by carbaryl, methyl parathion and pentachlorophenol.	1980	-	-	Only one exposure concentration	
Coors and De Meester	Synergistic, antagonistic and additive effects of multiple stressors: Predation threat, parasitism and pesticide exposure in <i>Daphnia magna</i> .	2008	Cladoceran, <i>Daphnia magna</i>	21 d reduced reproduction at 5.6	Only one exposure concentration; Lack of exposure details	Dilution water not characterized
Coppage	Anticholinesterase action of pesticidal carbamates in the central nervous system of poisoned fishes.	1977	Sailfin molly, <i>Poecilia latipinna</i>	15 hr reduced survival at 1,333	Only one exposure concentration; Lack of exposure details	Dilution water not characterized
Courtemanch and Gibbs	The effects of sevin-4-oil® on lentic communities.	1978	-	-	Formulation	Sevin-4-oil®
Cutkomp et al.	ATPase activity in fish tissue homogenates and inhibitory effects of DDT and related compounds.	1971	Bluegill, <i>Lepomis macrochirus</i>	-	Surgically altered test species	Homogenized tissue
Dahlberg	Toxicity to acrolein to barnacles (<i>Balanus eburneus</i>).	1971	Barnacle, <i>Balanus eburneus</i>	-	Not applicable	No carbaryl toxicity information
Damalas et al.	Bispyribac-sodium efficacy on early watergrass (<i>Echinochloa oryzoides</i>) and late watergrass (<i>Echinochloa phyllopogon</i>) as affected by coapplication of selected rice herbicides and insecticides.	2008	Early Watergrass, <i>Echinochloa oryzoides</i> Late Watergrass, <i>Echinochloa phyllopogon</i>	-	Mixture	Bispyribac-sodium and carbaryl

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Das and Adhikary	Toxicity of three pesticides to several rice-field cyanobacteria.	1996	Cyanobacteria	-	Formulation	Sevin (50% WP)
Das and Kumar	Toxicity of carbaryl on α -amylase of the fish <i>Colisa fasciatus</i> .	1993	Giant gourami, <i>Colisa fasciatus</i>	48 hr LC50=112	Not North American species; Lack of detail	No procedure information provided
Das and Rajagopalan	Susceptibility of larvae of <i>Culex fatigans</i> (Wiedmann), <i>Anopheles stephensi</i> (Liston) and <i>Aedes aegypti</i> (Linn.) to insecticides in Pondicherry.	1979	Mosquito, <i>Culex fatigans</i> Mosquito, <i>Anopheles stephensi</i> Mosquito, <i>Aedes aegypti</i>	-	Formulation	Sevimol (40% carbaryl)
Davey et al.	Toxicity of five ricefield pesticides to the mosquitofish, <i>Gambusia affinis</i> , and green sunfish, <i>Lepomis cyanellus</i> , under laboratory and field conditions in Arkansas.	1976	Mosquitofish, <i>Gambusia affinis</i> Green sunfish, <i>Lepomis cyanellus</i>	-	Formulation	
Davidson et al.	Effects of chytrid and carbaryl exposure on survival, growth and skin peptide defenses in foothill yellow-legged frogs.	2007	Foothill yellow-legged frog, <i>Rana boylei</i>	-	Formulation; Only one exposure concentration	
Deshmukh and Keshavan	Acute toxicity of DDT and sevin to the bullfrog, <i>Rana tigrina</i> .	1984	Bullfrog, <i>Rana tigrina</i>	96 hr LC50=70,000	Not North American species	
Dhanapakiam and Premlatha	Histopathological changes in the kidney of <i>Cyprinus carpio</i> exposed to malathion and sevin.	1994	Carp, <i>Cyprinus carpio</i>	-	Formulation	Sevin (commercial grade)
Dimayuga et al.	Insecticide-induced accumulation of melanomacrophage centers (MMCs) in Nile tilapia (<i>Oreochromis niloticus</i> Linn.).	2008	Nile tilapia, <i>Oreochromis niloticus</i>	-	Formulation	Sevin
Dodson et al.	Behavioral responses of <i>Daphnia pulex</i> exposed to carbaryl and <i>Chaoborus kairomone</i>	1995	Cladoceran, <i>Daphnia pulex</i>	-	Only two exposure concentrations	
Downing et al.	Community and ecosystem responses to a pulsed pesticide disturbance in freshwater ecosystems.	2008	Aquatic mesocosm	-	Formulation	Sevin (5% carbaryl)
Dumbauld et al.	Efficacy of the pesticide carbaryl for thalassinid shrimp control in Washington State oyster (<i>Crassostrea gigas</i> , Thunberg, 1793) aquaculture.	1997	-	-	Mixture, Field exposure; Formulation	
Dumbauld et al.	Response of an estuarine benthic community to application of the pesticide carbaryl and cultivation of Pacific oysters (<i>Crassostrea gigas</i>) in Willapa Bay, Washington.	2001	Estuarine benthic community	-	Formulation; Mixture	Field exposure
Dumbauld et al.	An integrated pest management program for burrowing shrimp control in oyster aquaculture.	2006	-	-	Mixture	

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Durfey and Simpson	Control of two burrowing shrimp species, ghost shrimp, <i>Callinassa californiensis</i> and mud shrimp, <i>Upogebia pugettensis</i> , using subsurface injection of carbaryl ("sevin") as an alternative to aerial application in preparation of oyster beds for seeding.	1995	Ghost shrimp, <i>Callinassa californiensis</i> Mud shrimp, <i>Upogebia pugettensis</i>	-	Sediment exposure	
El-Magid	Effects of some pesticides on the growth of blue-green alga <i>Spirulina platensis</i> .	1986	Blue-green alga, <i>Spirulina platensis</i>	5 d Growth NOEC=1,000 LOEC=5,000	Not North American species	
Epstein and Legator	The mutagenicity of pesticides concepts and evaluation.	1971	-	-	Review of previous studies	
Estenik and Collins	In vivo and in vitro studies of mixed-function oxidase in an aquatic insect, <i>Chironomus riparius</i> . In: Pesticide and xenobiotic metabolism in aquatic organisms.	1979	Midge, <i>Chironomus riparius</i>	24 hr LC50=104.5	Excessive solvent used	0.1%
European Commission DG Environment	Endocrine disrupters: Study on gathering information on 435 substances with insufficient data.	2002	-	-	Review of previous studies	
Feldhaus et al.	Interactive effects of pesticide mixtures on the neurobehavioral responses and AChE levels of the planaria.	1998	Brown planaria, <i>Dugesia tigrina</i>	-	Dilution water is deionized water without the proper salts added	
Fernandez et al.	Amphibian micronucleus test(s): A simple and reliable method for evaluating in vivo genotoxic effects of freshwater pollutants and radiations: Initial assessment.	1993	Newt, <i>Pleurodeles waltl</i>	8 d increased micronucleus frequency at 2,500	Only one exposure concentration	
Fernandez-Alba et al.	Toxicity of pesticides in wastewater: a comparative assessment of rapid bioassays.	2001	Bacterium, <i>Vibrio fischeri</i> Cladoceran, <i>Daphnia magna</i>	-	Not applicable	No carbaryl toxicity information
Ferrari et al.	Effects of carbaryl and azinphos methyl on juvenile rainbow trout (<i>Oncorhynchus mykiss</i>) detoxifying enzymes.	2007b	Rainbow trout, <i>Oncorhynchus mykiss</i>	96 hr decreased liver CbE at 1,000	Only two exposure concentrations	
Ferrari et al.	Antioxidant responses to azinphos methyl and carbaryl during the embryonic development of the toad <i>Rhinella (Bufo) arenarum</i> Hensel.	2009	Toad, <i>Rhinella (Bufo) arenarum</i>	Increased embryonic development malformations at 20,000	Not North American species; Only one exposure concentration	
Fischer et al.	The effect of benzimidazole, carbamate and organophosphorous pesticides on the oxygen-dependent nuclear volume alterations in the chloragocytes of <i>Tubifex tubifex</i> Mull.	1982	Tubificid worm, <i>Tubifex tubifex</i>	5 d did not induce nuclear swelling within the physiological range at 1,000	Only one exposure concentration	
Fisher and Lohner	Studies on the environmental fate of carbaryl as a function of pH.	1986	Midge, <i>Chironomus riparius</i>	24 hr LC50=103	Excessive solvent used	1 mL/L

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Author	Title	Date	Organism(s)	Concentration (µg/L)	Reason Unused	Expanded Reason
Fisher et al.	Quantitative structure-activity relationships for predicting the toxicity of pesticides in aquatic systems with sediment.	1993	Midge, <i>Chironomus riparius</i>	-	Excessive solvent used; Sediment in exposure media	1 mL/500 mL acetone
Fitzgerald et al.	Studies on chemicals with selective toxicity to blue-green algae.	1952	-	-	Not applicable	No carbaryl toxicity information
Fleeger et al.	Indirect effects of contaminants in aquatic ecosystems.	2003	-	-	Review of previous studies	
Foster and Tullis	A quantitative structure-activity relationship between partition coefficients and the acute toxicity of naphthalene derivatives in <i>Artemia salina</i> nauplii.	1984	Brine shrimp, <i>Artemia salina</i>	-	Brine shrimp	
Freitag et al.	Ecotoxicological profile analysis. VII. Screening chemicals for their environment behavior by comparative evaluation	1982	Green alga, <i>Chlorella fusca</i> var. <i>vacuolated</i>	-	Bioaccumulation study: steady state not reached, static exposure	
Frempong-Boadu	A laboratory study of the effectiveness of methoxychlor, fenthion and carbaryl against blackfly larvae (Diptera: Simuliidae).	1966	Blackflies, <i>Prosimulium mixtum</i> , <i>P. magnum</i> , <i>Simulium venustum</i> , <i>S. tuberosum</i>	-	Formulation	1% carbaryl
Galindo Reyes et al.	Effects of pesticides on DNA and protein of shrimp larvae <i>Litopenaeus stylirostris</i> of the California Gulf.	2002	Shrimp larvae, <i>Litopenaeus stylirostris</i>	-	Formulation	Sevin
Garcia-Ripoll et al.	Confirming <i>Pseudomonas putida</i> as a reliable bioassay for demonstrating biocompatibility enhancement by solar photo-oxidative process of a biorecalcitrant effluent.	2009	Bacteria, <i>Pseudomonas putida</i>	-	Sediment exposure	
Ghosh	Interrelationship of acetylcholinesterase-acetylcholine, triiodothyronine-thyroxine and gonadotropin-gonadotropin releasing hormone in pesticide treated murrel, <i>Channa punctatus</i> (Bloch).	1990	Snakehead catfish, <i>Channa punctatus</i>	-	Lack of detail (procedures); Not North American species; Only one exposure concentration	
Ghosh and Bhattacharya	Elevation of c-reactive protein in serum of <i>Channa punctatus</i> as an indicator of water pollution.	1992	Snakehead catfish, <i>Channa punctatus</i>	-	Not North American species; Only one exposure concentration	
Ghosh et al.	Impact of nonlethal levels of metacid-50 and carbaryl on thyroid function and cholinergic system of <i>Channa punctatus</i> .	1989	Snakehead catfish, <i>Channa punctatus</i>	-	Not North American species; Only one exposure concentration	
Ghosh et al.	Impairment of the regulation of gonadal function in <i>Channa punctatus</i> by the metacid-50 and carbaryl under laboratory and field conditions.	1990	Snakehead catfish, <i>Channa punctatus</i>	-	Not North American species; Only one exposure concentration	

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Author	Title	Date	Organism(s)	Concentration (µg/L)	Reason Unused	Expanded Reason
Ghosh et al.	Glutathione depletion in the liver and kidney of <i>Channa punctatus</i> exposed to carbaryl and metacid-50.	1993	Snakehead catfish, <i>Channa punctatus</i>	-	Not North American species; Only one exposure concentration	
Gibbs	Effects of a split application of sevin-4-oil on aquatic organisms.	1979	-	-	Formulation	Sevin-4-oil
Gibbs et al.	The effects on pond macroinvertebrates from forest spraying of carbaryl (sevin-4-oil) and its persistence in water and sediment.	1981	-	-	Formulation	Sevin-4-oil
Gibbs et al.	The effects in 1982 on pond macroinvertebrates from forest spraying of carbaryl, sevin-4-oil, in 1980.	1982	-	-	Formulation	Sevin-4-oil
Gibbs et al.	Persistence of carbaryl (sevin-4-oil) in woodland ponds and its effects on pond macroinvertebrates following forest spraying.	1984	Spruce budworm, <i>Choristoneura fumiferana</i>	-	Formulation	Sevin-4-oil (49% carbaryl)
Gilbert et al.	Rapid assessment of metabolic activity in marine microalgae: application in ecotoxicological test and evaluation of water quality.	1992	Green alga, <i>Tetraselmis suecica</i> <i>Skeletonema costatum</i> <i>Prorocentrum lima</i>	-	Lack of details (procedure, purity, exposure media)	
Gill et al.	Gill, liver, and kidney lesions associated with experimental exposures to carbaryl and dimethoate in the fish (<i>Puntius conchoni</i> Ham.).	1988	Barb, <i>Puntius conchoni</i>	-	Not North American species; Formulation	Sevin (50% WP)
Gillott et al.	The role of sediment as a modifying factor in pesticide-algae interactions.	1975	Alga, <i>Euglena gracilis</i>	24 hr C-14 incorporation NOEC=5 LOEC=10	Excessive solvent used	1 mL/L
Goel and Srivastava	Laboratory evaluation of some molluscicides against fresh water snails, <i>Indoplanorbis</i> and <i>Lymnaea</i> species.	1981	Snail, <i>Indoplanorbis exustus</i> Snail, <i>Lymnaea acuminata</i>	24 hr LC50=30,000 LC50=15,980	Not North American species; Lack of exposure details	Dilution water not characterized
Gouda et al.	Toxicity of dimecron, sevin and lindex to <i>Anabas scandens</i> and <i>Heteropneustes fossilis</i> .	1981	Climbing perch, <i>Anabas scandens</i> Catfish, <i>Heteropneustes fossilis</i>	-	Not North American species; Formulation	Sevin (50% carbaryl)
Gray et al.	Emerging issues: The effects of endocrine disrupters on reproductive development.	1996	-	-	Not applicable	No carbaryl toxicity information
Groba and Trzcinska	Effect of selected organophosphorous and carbamate insecticides on rainbow trout (<i>Salmo gairdneri</i> R.).	1979	Rainbow trout, <i>Oncorhynchus mykiss</i>	-	Only two exposure concentrations; Lack of exposure details	Text in foreign language
Grosch	Reproduction tests: the toxicity for <i>Artemia</i> of derivatives from non-persistent pesticides.	1973	Brine shrimp, <i>Artemia salina</i>	-	Brine shrimp	

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Author	Title	Date	Organism(s)	Concentration (µg/L)	Reason Unused	Expanded Reason
Gruber and Munn	Organophosphate and carbamate insecticides in agricultural waters and cholinesterase (ChE) inhibition in common carp (<i>Cyprinus carpio</i>).	1998	Common carp, <i>Cyprinus carpio</i>	-	Mixture	
Gupta and Sahai	Qualitative detection of organochlorine and carbamate residues in the brain of catfish, <i>Heteropenustes fossilis</i> (Bloch) by thin layer chromatography	1989	Catfish, <i>Heteropenustes fossilis</i>	-	Lack of detail (procedure)	
Gupta and Sundararaman	Correlation between burrowing capability and AChE activity in the earthworm, <i>Pheretima posthuma</i> , on exposure to carbaryl.	1991	Earthworm, <i>Pheretima posthuma</i>	-	Dilution water is distilled water without proper salts added	
Haines	Effect of an aerial application of carbaryl on brook trout (<i>Salvelinus fontinalis</i>)	1981	Brook trout, <i>Salvelinus fontinalis</i>	-	Lack of details (procedure)	
Han Il et al.	Studies on control effects of pesticide applications against the vector mosquito larvae in rice fields in Korea.	1981	Mosquito, <i>Culex tritaeniorhynchus</i> <i>Anopheles sinensis</i>	-	Formulation	Sevin (50% carbaryl)
Hanazato	Effects of long- and short-term exposure to carbaryl on survival, growth and reproduction of <i>Daphnia ambigua</i> .	1991a	Cladoceran, <i>Daphnia ambigua</i>	-	Lack of details (procedure, duration)	
Hanazato	Effects of repeated application of carbaryl on zooplankton communities in experimental ponds with or without the predator <i>Chaoborus</i> .	1991b	Zooplankton community	-	Only two exposure concentrations	
Hanazato	Pesticides as chemical agents inducing helmet formation on <i>Daphnia ambigua</i> .	1991c	Cladoceran, <i>Daphnia ambigua</i>	-	Excessive solvent used	3.5 mL/L ethanol
Hanazato	Insecticide inducing helmet development in <i>Daphnia ambigua</i> .	1992	Cladoceran, <i>Daphnia ambigua</i>	-	Only two exposure concentrations	
Hanazato	Combined effect of the insecticide carbaryl and the <i>Chaoborus</i> kairomone on helmet development in <i>Daphnia ambigua</i> .	1995	Cladoceran, <i>Daphnia ambigua</i>	-	Lack of details (procedure, duration)	
Hanazato and Dodson	Complex effects of a kairomone of <i>Chaoborus</i> and an insecticide on <i>Daphnia pulex</i> .	1992	Cladoceran, <i>Daphnia pulex</i>	-	Lack of details (procedure, duration)	
Hanazato and Hirokawa	Changes in vulnerability of <i>Daphnia</i> to an insecticide application depending on the population phase.	2004	Cladoceran, <i>Daphnia pulex</i>	-	Only one exposure concentration	
Hanazato and Yasuno	Effects of a carbamate insecticide, carbaryl, on the summer phyto- and zooplankton communities in ponds.	1987	Zooplankton community	-	Only one exposure concentration	

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Hanazato and Yasuno	Influence of overwintering <i>Daphnia</i> on spring zooplankton communities: an experimental study.	1989a	Zooplankton community	-	Only one exposure concentration; Possible mixture used	
Hanazato and Yasuno	Effects of carbaryl on the spring zooplankton communities in ponds.	1989b	Zooplankton community	-	Only one exposure concentration	
Hanazato and Yasuno	Influence of persistence period of an insecticide on recovery patterns of a zooplankton community in experimental ponds.	1990a	Zooplankton community	-	Formulation	50% carbaryl
Hanazato and Yasuno	Influence of time of application of an insecticide on recovery patterns of a zooplankton community in experimental ponds.	1990b	Zooplankton community	-	Formulation	50% carbaryl
Hanazato and Yasuno	Influence of <i>Chaoborus</i> density on the effects of an insecticide on zooplankton communities in ponds.	1990c	Zooplankton community	-	Formulation	50% carbaryl
Hardersen and Wratten	The sensitivity of the nymphs of two New Zealand damselfly species (Odonata: Zygoptera) to azinphos-methyl and carbaryl.	1996	Damselfly, <i>Xanthocnemis zealandia</i> <i>Austrolestes colenonis</i>	48 hr LC50=600 LC50=3,130	Not North American species	
Hardersen and Wratten	The effects of carbaryl exposure on the penultimate larval instars of <i>Xanthocnemis zealandica</i> on emergence and fluctuating asymmetry.	1998	Damselfly, <i>Xanthocnemis zealandia</i>	-	Not North American species; Exposure concentrations fluctuated widely	
Harilal and Sahai	Qualitative identification of metabolites of carbaryl in the gonads of catfish <i>Heteropneustes fossilis</i> (Bloch).	1990	Catfish, <i>Heteropneustes fossilis</i>	-	Bioaccumulation study: steady state not reached; Not North American species	
Havens	An experimental comparison of the effects of two chemical stressors on a freshwater zooplankton assemblage.	1994	Zooplankton community	-	Dilution water not characterized; Formulation	Commercial grade
Havens	Insecticide (carbaryl, 1-naphthyl-n-methylcarbamate) effects on a freshwater plankton community: zooplankton size, biomass, and algal abundance.	1995	Plankton community	-	Formulation	Commercial grade
Haynes et al.	The toxicity of sevin to goldfish.	1958	Goldfish, <i>Carassius auratus</i>	-	Formulation	Sevin (50% carbaryl)
Heldal et al.	Toxic responses of the green alga <i>Dunaliella bioculata</i> (Chlorophyceae, Volvocales) to selected oxidized hydrocarbons.	1984	Green alga, <i>Dunaliella bioculata</i>	-	Not applicable	No carbaryl toxicity information
Hemingway and Georghiou	Studies on the acetylcholinesterase of <i>Anopheles albimanus</i> resistant and susceptible to organophosphate and carbamate insecticides.	1983	Mosquito, <i>Anopheles albimanus</i>	24 hr LC50=890	Excessive solvent used; Waxed cups used	1 mL/100 mL

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Hendrick et al.	Effects of some insecticides on the survival, reproduction, and growth of the Louisiana red crawfish.	1966	Crawfish, <i>Procambarus clarkia</i>	-	Formulation; Field application	
Hermann	Routine testing of new Bayer pesticides for fish toxicity, as part of the product development programme.	1975	-	-	Review of previous studies	
Hernandez et al.	Toxicity of ethyl-parathion and carbaryl on early stages of the development of sea urchin.	1986	Sea urchin, <i>Pseudechinus magellanicus</i>	-	Text in foreign language; Not North American species	
Hernandez et al.	Toxicity of ethyl-parathion and carbaryl on early development of sea urchin.	1990	Sea urchin, <i>Pseudechinus magellanicus</i>	96 hr EC50=92.5	Not North American species	
Hidaka et al.	Avoidance of pesticides with medakas (<i>Oryzias latipes</i>).	1984	Medaka, <i>Oryzias latipes</i>	-	Not North American species; Lack of exposure details	Text in foreign language
Hiltibran	Oxygen and phosphate metabolism of bluegill liver mitochondria in the presence of some insecticides.	1974	Bluegill, <i>Lepomis macrochirus</i>	-	Surgically altered test species	Liver mitochondria
Hirose and Kitsukawa	Acute toxicity of agricultural chemical to seawater teleosts, with special respect to TLM and the vertebral abnormality.	1976	Medaka, <i>Oryzias latipes</i>	-	Text in foreign language	
Holcombe et al.	The acute toxicity of selected substituted phenols, benzenes and benzoic acid esters to fathead minnows <i>Pimephales promelas</i> .	1984	Fathead minnows, <i>Pimephales promelas</i>	-	Not applicable	No carbaryl toxicity information
Hopf and Muller	Laboratory breeding and testing of <i>Australorbis glabratus</i> for molluscicidal screening.	1962	Snail, <i>Australorbis glabratus</i>	-	Lack of details (procedure)	
Hopkins and Winne	Influence of body size on swimming performance of four species of neonatal natricine snakes acutely exposed to a cholinesterase-inhibiting pesticide.	2006	Semi-aquatic snakes, <i>Nerodia taxispilota</i> , <i>N. rhombifer</i> , <i>N. fasciata</i> , <i>Seminatrix pygaea</i>	-	Formulation	Sevin (commercial grade)
Hopkins et al.	Differential swimming performance of two natricine snakes exposed to a cholinesterase-inhibiting pesticide.	2005	Black swamp snake, <i>Seminatrix pygaea</i> Diamondback water snake, <i>Nerodia rhombifer</i>	-	Only two exposure concentrations; Formulation	Sevin®
Hota et al.	Metabolic effects of kilex carbaryl on a fresh water teleost, <i>Channa punctatus</i> (Bloch).	1993	Snakehead catfish, <i>Channa punctatus</i>	24 hr decline in protein concentration in liver at 2,000	Only one exposure concentration; Not North American species	
Hulbert	Effects of sevin, a spruce budworm insecticide on fish and invertebrates in the Mattawamkeag River in 1976.	1978	-	-	Formulation	Sevin-4-oil

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Huque	Preliminary report on the residues of carbaryl granules in rice plants.	1972	-	-	Formulation; Bioaccumulation study: steady state not reached	Carbaryl 10% granules
Hydorn et al.	Effect of forest spraying with acephate insecticide on consumption of spiders by brook trout (<i>Salvelinus fontinalis</i>).	1979	-	-	Not applicable	No carbaryl toxicity information
Ishii and Hashimoto	Metabolic fate of carbaryl (1-naphthyl N-methyl carbamate) orally administered to carp, <i>Cyprinus carpio</i> .	1970	Carp, <i>Cyprinus carpio</i>	-	Dietary exposure; Lack of exposure details	Text in foreign language
Jacob et al.	Toxicity of certain pesticides found in the habitat to the larvivorous fishes <i>Aplocheilus lineatus</i> (Cuv. & Val.) and <i>Macropodus cupanus</i> (Cuv. & Val.).	1982	Panchax, <i>Aplocheilus lineatus</i> Paradise fish, <i>Macropodus cupanus</i>	-	Not North American species; Formulation	Sevin (50% WP)
Jadhv et al.	Effect of pesticides on amylase activity in digestive gland of fresh water bivalve <i>Corbicula striatella</i> .	1995	Bivalve, <i>Corbicula striatella</i>	96 hr decrease amylase activity at 2,500	Not North American species; Only one exposure concentration	
Jadhv et al.	Carbaryl toxicity to freshwater bivalve <i>Corbicula striatella</i> .	1996	Bivalve, <i>Corbicula striatella</i>	96 hr LC50=5,100	Not North American species	
James and Sampath	Combined toxic effects of carbaryl and methyl parathion on survival, growth, and respiratory metabolism in <i>Heteropneustes fossilis</i> (Bloch).	1994	Catfish, <i>Heteropneustes fossilis</i>	-	Not North American species; Formulation	Carbaryl (50% WDP)
Jamback and Frempong-Boadu	Testing blackfly larvicides in the laboratory and in streams.	1966	Blackfly, <i>Simulium sp.</i>	5 min. 47% detachment at 400	Only two exposure concentrations; Lack of exposure details	Dilution water not characterized
Jauhar and Kulshrestha	Histopathological changes induced by the sublethal doses of endosulfan and carbaryl in the intestine of <i>Channa striatus</i> Bloch.	1983	Snakehead catfish, <i>Channa striatus</i>	-	Not North American species; Formulation	Sevin (50 WP)
Jauhar and Kulshrestha	Histopathological effects induced by sublethal doses of sevin and thiodan on the gills of <i>Channa striatus</i> Bloch. (Pisces, Channidae).	1985	Snakehead catfish, <i>Channa striatus</i>	-	Not North American species; Formulation	Sevin (50 WP)
Jayaprada and Ramana Rao	Carbaryl toxicity on tissue acetylcholinesterase in the penaeid prawn, <i>Metapenaeus monceros</i> (Fabricius) a monitoring study.	1991	Prawn, <i>Metapenaeus monceros</i>	96 hr LC50=24.87	Not North American species	
Jeyasingam et al.	The relative toxicities of insecticides on aquatic insect <i>Eretes sticticus</i> (Linn.) (Coleoptera: Dytiscidae).	1978	Diving beetle, <i>Eretes sticticus</i>	48 hr TLm=890	Not North American species	

Appendix J. List of Carbaryl Studies Not Used in Document Along with Reasons

Author	Title	Date	Organism(s)	Concentration (µg/L)	Reason Unused	Expanded Reason
John	Alteration of certain blood parameters of freshwater teleost <i>Mystus vittatus</i> after chronic exposure to metasystox and sevin.	2007	Catfish, <i>Mystus vittatus</i>	30 d decreased blood ESR and Hb% at 7,000	Not North American species; Only one exposure concentration; Lack of exposure details	Dilution water not characterized
John and Prakash	Acute toxicity of metasystox and sevin to <i>Mystus vittatus</i> .	1998	Catfish, <i>Mystus vittatus</i>	-	Not North American species; Formulation	Sevin® (505 carbaryl)
Joshi and Kumar	Acid and alkaline phosphates activity in different tissues of fresh water crab, <i>Paratelphusa masoniana</i> (Henderson) to pesticide exposure.	2001	Crab, <i>Paratelphusa masoniana</i>	-	Lack of details (procedure)	
Juchelka and Snell	Rapid toxicity assessment using ingestion rate of cladocerans and ciliates.	1995	-	-	Not applicable	No carbaryl toxicity information
Juhnke and Luedemann	Results of the investigation of 200 chemical compounds for acute fish toxicity with the golden orfe test.	1978	Golden orfe, <i>Leuciscus idus melanotus</i>	48 hr LC50=20,000	Lack of exposure details	Dilution water not characterized
Jyothi and Narayan	Toxic effects of carbaryl on gonads of freshwater fish, <i>Clarias batrachus</i> (Linnaeus).	1999a	Walking catfish, <i>Clarias batrachus</i>	-	Only one exposure concentration; Formulation	Commercial grade
Jyothi and Narayan	Certain pesticide-induced carbohydrate metabolic disorders in the serum of freshwater fish <i>Clarias batrachus</i> (Linn.).	1999b	Walking catfish, <i>Clarias batrachus</i>	-	Only one exposure concentration; Formulation	Sevin (50% WDP powder)
Jyothi and Narayan	Pesticide induced alterations of non-protein nitrogenous constituents in the serum of a fresh water cat fish, <i>Clarias batrachus</i> (Linn.).	2000	Walking catfish, <i>Clarias batrachus</i>	-	Formulation	Sevin (50% WDP powder)
Jyothi and Narayan	Effect of pesticides carbaryl and phorate on serum cholesterol level in fish, <i>Clarias batrachus</i> (Linn.).	2001	Walking catfish, <i>Clarias batrachus</i>	-	Only one exposure concentration; Formulation	Commercial grade
Kader et al.	The relative toxicities of ten biocides on <i>Spicodiantomus chelospinus</i> Rajendran (1973) [Copepoda: Calanoida].	1976	Copepod, <i>Spicodiantomus chelospinus</i>	48 hr TLm=130	Not North American species; Lack of exposure details	Dilution water not characterized
Kallander et al.	Recovery following pulsed exposure to organophosphorus and carbamate insecticides in the midge, <i>Chironomus riparius</i> .	1997	Midge, <i>Chironomus riparius</i>	-	Excessive solvent used	1.0 mL/L acetone
Kanazawa	Uptake and excretion of organophosphorus and carbamate insecticides by fresh water fish, motsugo, <i>Pseudorasbora parva</i> .	1975	Motsugo, <i>Pseudorasbora parva</i>	-	Not North American species; Bioaccumulation study: steady state not reached	
Kanazawa	Prediction of biological concentration potential of pesticides in aquatic organisms.	1980	Topmouth gudgeon, <i>Pseudorasbora parva</i>	14 d BCF=9 (whole body)	Not North American species	
Kanazawa	Bioconcentration potential of pesticides by aquatic organisms.	1981	-	-	Review of previous studies	

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Author	Title	Date	Organism(s)	Concentration (µg/L)	Reason Unused	Expanded Reason
Kanazawa	In vitro and in vivo effects of organophosphorus and carbamate insecticides on brain acetylcholinesterase activity of fresh-water fish, topmouth gudgeon.	1983a	Topmouth gudgeon, <i>Pseudorasbora parva</i>	24 hr reduced brain AChE by 72-78% at 4,000	Not North American species	
Kanazawa	A method of predicting the bioconcentration potential of pesticides by using fish.	1983b	Topmouth gudgeon, <i>Pseudorasbora parva</i>	-	Not North American species; Bioaccumulation: steady state not reached	
Kanazawa et al.	Distribution of carbaryl and 3,5-xyllyl methylcarbamate in an aquatic model ecosystem.	1975	Community model ecosystem	-	Sediment present in test media	
Karnak and Collins	The susceptibility to selected insecticides and acetylcholinesterase activity in a laboratory colony of midge larvae, <i>Chironomus tentans</i> (Diptera: Chironomidae).	1974	Midge, <i>Chironomus tentans</i>	24 hr LC50=2.5	Excessive solvent used; Test species fed	
Kasai et al.	P450 monooxygenases are an important mechanism of permethrin resistance in <i>Culex quinquefasciatus</i> Say larvae.	1998	Mosquito, <i>Culex quinquefasciatus</i>	-	Dilution water is distilled water without proper salts added	
Kashiwada et al.	Stage-dependent differences in effects of carbaryl on population growth rate in Japanese medaka (<i>Oryzias latipes</i>).	2008	Medaka, <i>Oryzias latipes</i>	14 d decreased embryo growth at 5,000	Not North American species; Only two exposure concentrations; Excessive solvent	0.1%
Kaur and Dhawan	Variable sensitivity of <i>Cyprinus carpio</i> eggs, larvae, and fry to pesticides.	1993	Common carp, <i>Cyprinus carpio</i>	-	Formulation	Sevin (50 WP)
Kaur and Dhawan	Effect of carbaryl on tissue composition, maturation, and breeding potential of <i>Cirrhina mrigala</i> (Ham.).	1996	Carp, <i>Cirrhina mrigala</i>	-	Formulation	50% WP
Kaur and Toor	Role of abiotic factors in the embryonic development of scale carp.	1980	Common carp, <i>Cyprinus carpio</i>	-	Lack of details (procedure)	
Kaur and Toor	Toxicity of some insecticides to the fingerlings of Indian major carp, <i>Cirrhina mrigala</i> (Hamilton).	1995	Carp, <i>Cirrhina mrigala</i>	-	Formulation	Carbaryl 50 WP
Kaur and Toor	Histopathological changes in the liver of fingerlings of Indian major carp, <i>Cirrhina mrigala</i> (Hamilton) exposed to some biocides.	1997	Carp, <i>Cirrhina mrigala</i>	-	Only one exposure concentration	
Kaushik and Kumar	Susceptibility of the freshwater crab <i>Paratelphusa masoniana</i> (Henserson) to three pesticides, singly and in combination.	1993	Crab, <i>Paratelphusa masoniana</i>	-	Not North American species; Formulation	Carbaryl 50% WP
Kaushik and Kumar	Midgut pathology of aldrin, monocrotophos, and carbaryl in the freshwater crab, <i>Paratelphusa masoniana</i> (Henderson).	1998	Crab, <i>Paratelphusa masoniana</i>	-	Not North American species; only one exposure concentration	

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Author	Title	Date	Organism(s)	Concentration (µg/L)	Reason Unused	Expanded Reason
Kay	Toxicology of pesticides: Recent advances.	1973	-	-	Review of previous studies	
Kem et al.	Inhibition of barnacle larval settlement and crustacean toxicity of some hoplonemertine pyridyl alkaloids.	2003	Barnacle, <i>Balanus amphitrite</i>	-	Not applicable	No carbaryl toxicity information
Khalil et al.	Filarial infectivity rate of <i>Culex pipiens molestus</i> subjected to sublethal concentrations of insecticides abate and sevin and distribution of infective filaria larvae in mosquito body regions.	1974	-	-	Lack of exposure details; Mixture	Dilution water not characterized
Khan and Nelson	Adverse effects of some selected agrochemicals and pharmaceuticals in aquatic environment with reference to amphibians and fish.	2005	-	-	Review of previous studies	
Khangarot et al.	Man and biosphere - studies on the Sikkim Himalayas. Part 6: Toxicity of selected pesticides to frog tadpole <i>Rana hexadactyla</i> (Lesson).	1985	Frog, <i>Rana hexadactyla</i>	-	Not North American species; Formulation	Kelex (50% carbaryl)
Khillare and Wagh	Chronic effects of endosulfan, malathion and sevin in the fresh water fish, <i>Barbus stigma</i> testis histopathology.	1987a	Barb, <i>Barbus stigma</i>	-	Not North American species; Only one exposure concentration	
Khillare and Wagh	Developmental abnormalities induced by the pesticides in the fish, <i>Barbus stigma</i> (Ham.).	1987b	Barb, <i>Barbus stigma</i>	-	Not North American species; Only one exposure concentration	
Khillare and Wagh	Long-term effects of pesticides endosulfan, malathion and sevin on the fish, <i>Puntius stigma</i> .	1988a	Fish, <i>Puntius stigma</i>	-	Not North American species; Lack of details (procedure)	
Khillare and Wagh	Acute toxicity of pesticides in the freshwater fish <i>Barbus stigma</i> : histopathology of the stomach.	1988b	Barb, <i>Barbus stigma</i>	-	Not North American species; Lack of details (procedure)	
Khillare and Wagh	Effect of certain pesticides on spermatogenesis in fish <i>Barbus stigma</i> (Ham.).	1989	Barb, <i>Barbus stigma</i>	-	Not North American species; Only one exposure concentration	
Kikuchi et al.	Screening of organophosphate insecticide pollution in water by using <i>Daphnia magna</i> .	2000	-	-	Not applicable	No carbaryl toxicity information
Kimura and Keegan	Toxicity of some insecticides and molluscicides for the Asian blood sucking leech, <i>Hirudo nipponia</i> Whitman.	1966	Leech, <i>Hirudo nipponia</i>	48 hr LC50=5,500	Not North American species	
Klassen et al.	Toxicities of certain larvicides to resistant and susceptible <i>Aedes aegypti</i> (L.)	1965	Mosquito, <i>Aedes aegypti</i>	LC50=4,400	Prior exposure; Lack of exposure details	Dilution water not characterized; Duration not given
Kolankaya	Organochlorine pesticide residues and their toxic effects on the environment and organisms in Turkey.	2006	-	-	Not applicable	No carbaryl toxicity information

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Author	Title	Date	Organism(s)	Concentration (µg/L)	Reason Unused	Expanded Reason
Korn	The uptake and persistence of carbaryl in channel catfish.	1973	Channel catfish, <i>Ictalurus punctatus</i>	-	Bioaccumulation: steady state not reached	
Koundinya and Murthi	Haematological studies in <i>Sarotherodon (Tilapia) mossambica</i> (Peters) exposed to lethal (LC50/48 hrs) concentration of sumithion and sevin.	1979	Mozambique tilapia, <i>Oreochromis mossambica</i>	-	Formulation	Sevin (commercial grade)
Koundinya and Ramamurthi	Comparative study of inhibition of acetylcholinesterase activity in the freshwater teleost <i>Sarotherodon (Tilapia) mossambica</i> (Peters) by sevin (carbamate) and sumithion (organophosphate).	1979	Mozambique tilapia, <i>Oreochromis mossambica</i>	-	Formulation	Sevin (commercial grade)
Koundinya and Ramamurthi	Effect of sub-lethal concentration of sumithion and sevin on certain hematological values of <i>Sarotherodon mossambicus</i> (Peters).	1980a	Mozambique tilapia, <i>Oreochromis mossambica</i>	-	Formulation	Carbaryl (50% WP)
Koundinya and Ramamurthi	Toxicity of sumithion and sevin to the freshwater fish, <i>Sarotherodon mossambicus</i> (Peters).	1980b	Mozambique tilapia, <i>Oreochromis mossambica</i>	-	Formulation	Sevin (50% WP)
Koundinya and Ramamurthi	Tissue respiration in <i>Sarotherodon mossambicus</i> (Peters) exposed to sub-lethal concentration of sumithion and sevin.	1981	Mozambique tilapia, <i>Oreochromis mossambica</i>	-	Formulation	Sevin
Koval'Chuk et al.	Acute toxicity of yalan, eptam and sevin for <i>Daphnia magna</i> .	1971	Cladoceran, <i>Daphnia magna</i>	-	Lack of exposure details	Text in foreign language
Krishnan and Chockalingam	Toxic and sublethal effects of endosulfan and carbaryl on growth and egg production of <i>Moina micrura</i> Kurz (Cladocera: Moinidae).	1989	Cladoceran, <i>Moina micrura</i>	24 hr LC50=119.6	Not North American species	
Kulshrestha and Arora	Effect of sublethal doses of carbaryl and endosulfan on the skin of <i>Channa striatus</i> Bloch.	1984a	Snakehead catfish, <i>Channa striatus</i>	-	Not North American species; Formulation	Sevin (50% WP)
Kulshrestha and Arora	Impairments induced by sublethal doses of two pesticides in the ovaries of a freshwater teleost <i>Channa striatus</i> Bloch.	1984b	Snakehead catfish, <i>Channa striatus</i>	-	Not North American species; Formulation	Sevin (50 WP)
Kulshrestha and Jauhar	Impairments induced by sublethal doses of sevin and thiodan on the brain of a freshwater teleost <i>Channa striatus</i> Bloch. (Channidae).	1986	Snakehead catfish, <i>Channa striatus</i>	-	Not North American species; Formulation	Sevin
Kumar and Banerjee	Effects of lethal toxicity of sevin (carbaryl) on the blood parameters in <i>Clarias batrachus</i> (L).	1991	Walking catfish, <i>Clarias batrachus</i>	-	Only one exposure concentration	

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Author	Title	Date	Organism(s)	Concentration (µg/L)	Reason Unused	Expanded Reason
Kurtak et al.	Management of insecticide resistance in control of the <i>Simulium damnosum</i> complex by the onchocerciasis control programme, West Africa: Potential use of negative correlation between organophosphate resistance and pyrethroid susceptibility.	1987	Blackfly, <i>Simulium damnosum</i>	-	Not North American species; Lack of exposure details	Dilution water not characterized; Duration not given
Labenia et al.	Behavioral impairment and increased predation mortality in cutthroat trout exposed to carbaryl.	2007	Cutthroat trout, <i>Oncorhynchus clarkii clarkii</i>	6 hr decreased swimming performance at 750	Lack of exposure details	Dilution water not characterized
Lakshmi et al.	Toxicity of endosulfan and carbaryl to a brackish water oligochaete <i>Pontodrilus bermudensis</i> .	2002	Oligochaete, <i>Pontodrilus bermudensis</i>	-	Not North American species; Lack of details (procedure)	
Lange et al.	Comparison of testing acute toxicity on embryo of zebrafish, <i>Brachydanio rerio</i> and RTG-2 cytotoxicity as possible alternatives to the acute fish test.	1995	Zebrafish, <i>Danio rerio</i>	-	In vitro study	
Li and Chen	Study on the acute toxicities of commonly used pesticides to two kinds of fish.	1981	Mosquitofish, <i>Gambusia patruelis</i> Tilapia, <i>Tilapia sp.</i> Carp, <i>Cyprinus carpio</i>	48 hr TLm=955 TLm=1,958 TLm=10,000	Text in foreign language	
Lichtenstein et al.	Toxicity and fate of insecticide residues in water.	1966	Mosquito, <i>Aedes aegypti</i>	24 hr 70% mortality at 1,000	Only one exposure concentration; Lack of exposure details	Distilled water without proper salts added
Lingaraja and Venugopalan	Pesticide induced physiological and behavioural changes in an estuarine teleost <i>Therapon jarbua</i> (Forsk).	1978	Fish, <i>Therapon jarbua</i>	-	Not North American species; Lack of details (procedure)	
Lloyd	The toxicity of ammonia to rainbow trout (<i>Salmo gairdnerii</i> Richardson).	1961	Rainbow trout, <i>Oncorhynchus mykiss</i>	-	Not applicable	No carbaryl toxicity information
Lohner and Fisher	Effects of pH and temperature on the acute toxicity and uptake of carbaryl in the midge, <i>Chironomus riparius</i> .	1990	Midge, <i>Chironomus riparius</i>	24 hr EC50=96	Excessive solvent used	2 mL/L
Lowe	Effects of prolonged exposure to sevin on an estuarine fish, <i>Leiostomus xanthurus</i> Lacepede.	1967	Spot, <i>Leiostomus xanthurus</i>	-	High control mortality; Only one exposure concentration	65%
Lubick	Order matters in pesticide exposures.	2007	Amphipod, <i>Gammarus pulex</i>	-	Mixture	Carbaryl and chlorpyrifos
Ma et al.	Differential responses of eight cyanobacterial and green algal species, to carbamate insecticides.	2006	Cyanobacteria and green alga	-	Lack of detail	LC50 values are not identified to the species
Macek	Acute toxicity of pesticide mixtures to bluegills.	1975	Bluegill, <i>Lepomis macrochirus</i>	-	Mixture	Sevin and malathion

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Author	Title	Date	Organism(s)	Concentration (µg/L)	Reason Unused	Expanded Reason
MacKenzie and Shearer	Chemical control of <i>Polydora websteri</i> and other annelids inhabiting oyster shells.	1959	Worm, <i>Polydora websteri</i>	-	Only one exposure concentration	
Maly	A study of the effects of pesticide on single and mixed species cultures of algae.	1980	-	-	Only one exposure concentration; No control information	
Manna and Ghosh	Anaerobic toxicity of sublethal concentration of carbaryl pesticide sevin to guppy <i>Lebistes reticulatus</i> .	1987	Guppy, <i>Lebistes reticulatus</i>	-	Lack of details (procedure, purity not given)	
Manonmani et al.	Establishment of a standard test method for determining susceptibility of mesocyclops to different insecticides.	1989	Copepod, <i>Mesocyclops sp.</i>	-	Not North American species; Formulation	Carbaryl (50% WP)
Mansour and Hassan	Pesticides and Daphnia. 3. An analytical bioassay method, using <i>Ceriodaphnia quadrangula</i> , for measuring extremely low concentrations of insecticides in waters.	1993	Cladoceran, <i>Ceriodaphnia quadrangula</i>	-	Mixture	
Marian et al.	Acute and chronic effects of carbaryl on survival, growth, and metamorphosis in the bullfrog (<i>Rana tigrina</i>).	1983	Bullfrog, <i>Rana tigrina</i>	-	Not North American species; Formulation	50% WP
Markey et al.	Insecticides and a fungicide affect multiple coral life stages.	2007	Coral, <i>Acropora millepora</i>	18 hr EC50=1	Not North American species	
Marking and Bills	Effects of contaminants on toxicity of the lampricides TFM and Bayer 73 to three species of fish.	1985	Rainbow trout, <i>Oncorhynchus mykiss</i> White sucker, <i>Catostomus commersoni</i> Fathead minnow, <i>Pimephales promelas</i>	-	Mixture	Carbaryl and TFM or Bayer 73
Marutani and Edirveerasingam	Influence of irrigation methods and an adjuvant on the persistence of carbaryl on pakchoi.	2006	Pakchoi, <i>Brassica rapa L.</i> subsp. <i>chinensis</i>	-	Only one exposure concentration; Filed exposure; Sediment present	
Massachusetts Pesticide Board	Report of the surveillance program conducted in connection with an application of carbaryl (sevin) for the control of gypsy moth on Cape Cod, Massachusetts.	1966	-	-	Formulation; Only one exposure concentration; Aerial application	
Mathur	Toxicity of sevin to certain fishes.	1974	Barb, <i>Esomus danrica</i> Catfish, <i>Heteropneustes fossilis</i> Catfish, <i>Channa punctatus</i> Rasbora, <i>Rasbora daniconius</i>	-	Not North American species; Formulation	Sevin

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Author	Title	Date	Organism(s)	Concentration (µg/L)	Reason Unused	Expanded Reason
Matos et al.	Biochemical and histological hepatic changes of Nile tilapia <i>Oreochromis niloticus</i> exposed to carbaryl.	2007	Nile tilapia, <i>Oreochromis niloticus</i>	14 d decreased hepatic activity at 250	Only two exposure concentrations	
Matsumura	Toxicology of insecticides.	1975	-	-	Review of previous studies	
Mazzeo et al.	Interclonal variation in response to simazine stress in <i>Lemna gibba</i> (Lemnaceae).	1998	Duckweed, <i>Lemna gibba</i>	-	Not applicable	No carbaryl toxicity information
McKim et al.	Use of respiratory-cardiovascular responses of rainbow trout (<i>Salmo gairdneri</i>) in identifying acute toxicity syndromes in fish: Part 2. Malathion, carbaryl, acrolein and benzaldehyde.	1987	Rainbow trout, <i>Oncorhynchus mykiss</i>	-	Surgically altered test species	Spinally transected
Megharaj et al.	The use of unicellular soil green algae for insecticide bioassay.	1989	Green alga, <i>Chlorella vulgaris</i> <i>Scenedesmus bijugatus</i>	-	Agar exposure media	
Metcalf and Sanborn	Pesticides and environmental quality in Illinois.	1975	-	-	Review of previous studies	
Metts et al.	Interaction of an insecticide with larval density in pond-breeding salamanders (<i>Ambystoma</i>).	2005	Salamander, <i>Ambystoma maculatum</i> <i>A. opacum</i>	-	Only two exposure concentrations; Formulation	Sevin (22.5% carbaryl)
Meyer	Quarterly report of progress, April-June, 1981.	1981	Rainbow trout	-	Lack of exposure details	Dilution water not characterized; No scientific name given
Mills and Semlitsch	Competition and predation mediate the indirect effects of an insecticide on southern leopard frogs.	2004	Southern leopard frog, <i>Rana sphenoccephala</i>	-	Formulation	Sevin (21.35 carbaryl)
Mishra et al.	Toxicity of kilex carbaryl to a fresh water teleost <i>Channa punctatus</i> (Bloch).	1991	Snakehead catfish, <i>Channa punctatus</i>	96 hr TLm=14,000	Not North American species	
Mishra et al.	Responses of interregional cells of freshwater teleost, <i>Channa punctatus</i> (Bloch), exposed to sublethal concentrations of carbaryl and cartap.	2006	Snakehead catfish, <i>Channa punctatus</i>	96 hr caused kidney hyperplasia at 5,200	Not North American species; Only one exposure concentration	
Mitsubishi et al.	Effects of insecticides on cultures of insect cells.	1970	Mosquito, <i>Aedes aegypti</i>	-	Surgically altered test species	Cell culture
Mora et al.	Cholinesterase activity as potential biomarker in two bivalves.	1999	Mussel, <i>Mytilus galloprovincialis</i> Asiatic clam, <i>Corbicula fluminea</i>	-	Lack of exposure details	Dilution water not characterized
Mora et al.	Relationship between toxicokinetics of carbaryl and effect on acetylcholinesterase activity in <i>Pomacea patula</i> snail.	2000	Snail, <i>Pomacea patula</i>	-	Excessive solvent used	4 mL/L ethanol
Morgan	Monitoring pesticides by means of changes in electric potential caused by fish opercular rhythms.	1975	Largemouth bass	-	Formulation; No scientific name given	Karbaspray (50% carbaryl)

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Muirhead-Thomson	Laboratory evaluation of pesticide impact on stream invertebrates.	1973	Dragonfly naiads	-	Formulation	Sevin
Mulla et al.	Control of chironomid midges in recreational lakes.	1971	Midges	-	Formulation	20% carbaryl
Mulla et al.	Aquatic midge larvicides, their efficacy and residues in water, soil, and fish in a warm-water lake.	1973	Midges	-	Formulation	Carbaryl (20% granules)
Murray and Guthrie	Effects of carbaryl, diazinon and malathion on native aquatic populations of microorganisms.	1980	-	-	Only one exposure concentration; Lack of exposure details	Dilution water not characterized
Nalecz-Jawecki and Sawicki	Spirotox - a new tool for testing the toxicity of volatile compounds.	1999	-	-	Not applicable	No carbaryl toxicity information
Nalecz-Jawecki et al.	The sensitivity of protozoan <i>Spirostomum ambiguum</i> to selected pesticides.	2002	Protozoan, <i>Spirostomum ambiguum</i>	-	Excessive solvent used	1% acetone
Naqvi and Ferguson	Pesticide tolerances of selected freshwater invertebrates.	1968	Cyclopoid copepods	-	Prior exposure pesticides	
Naqvi and Hawkins	Toxicity of selected insecticides (thiodan, security, spartan, and sevin) to mosquitofish, <i>Gambusia affinis</i> .	1988	Mosquitofish, <i>Gambusia affinis</i>	-	Formulation	Sevin (5% carbaryl)
Nishiuchi	Toxicity of formulated pesticides to some fresh water organisms.	1976	Japanese toad, <i>Bufo bufo japonicas</i>	-	Not North American species; Lack of exposure details	Text in foreign language
Nishiuchi	Toxicity of formulated pesticides to some freshwater organisms.	1977	-	-	Not North American species; Lack of exposure details	Text in foreign language
Nishiuchi and Asano	Toxicity of formulated agrochemicals to fresh water organisms.	1978	Dragonfly, <i>Orthetrum albistylum speciosum</i>	-	Not North American species; Lack of exposure details	Text in foreign language
Nishiuchi and Yoshida	Toxicities of pesticides to some fresh water snails.	1972	Red snail, <i>Indoplanorbis exustus</i> Marsh snail, <i>Semisulcospira libertina</i> Round snail, <i>Cipangopaludina malleata</i> Saka snail, <i>Physa acuta</i>	48 hr TLm=28,000 TLm=25,000 TLm=30,000 TLm=27,000	Not North American species	
Nogrady and Keshmirian	Rotifer neuropharmacology - I. Cholinergic drug effects on oviposition of <i>Philodina acuticornis</i> (Rotifera, Aschelminthes).	1986	Rotifer, <i>Philodina acuticornis</i>	-	Formulation	Sevin

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Author	Title	Date	Organism(s)	Concentration (µg/L)	Reason Unused	Expanded Reason
Nollenberger	Toxicant-induced changes in brain, gill, liver, and kidney of brook trout exposed to carbaryl, atrazine, 2,4-dichlorophenoxyacetic acid, and parathion: a cytochemical study.	1981	Brook trout, <i>Salvelinus fontinalis</i>	-	Only one exposure concentration	
Omkar and Murti	Toxicity of some pesticides to the freshwater prawn <i>Macrobrachium dayanum</i> (Henderson) (Decapoda, Caridea).	1985	Prawn, <i>Macrobrachium dayanum</i>	-	Formulation	Sevin (50 WP)
Omkar and Shukla	Toxicity of insecticides to <i>Macrobrachium lamarrei</i> (H. Milne Edwards) (Decapoda: Palaemonidae).	1985	Prawn, <i>Macrobrachium lamarrei</i>	96 hr LC50=19	Not North American species	
Owen	Aquatic insect populations reduced by aerial spraying of insecticide sevin.	1967	-	-	Formulation	Sevin
Padhy and Mohapatra	Toxicity of two carbamate insecticides to the cyanobacterium <i>Anabaena</i> PCC 7120 and the computations of partial lethal concentrations by the probit method.	2001	Cyanobacteria, <i>Anabaena</i>	-	Formulation	Sevin (50W)
Palanichamy et al.	Effect of pesticides on protein metabolism in the freshwater catfish <i>Mystus vittatus</i> .	1989	Catfish, <i>Mystus vittatus</i>	-	Not North American species; Only one exposure concentration	
Panigrahy and Padhy	Toxicity of carbamate pesticides to cells, heterocysts and akinetes of the cyanobacterium <i>Cylindrospermum sp.</i>	2000	Cyanobacterium, <i>Cylindrospermum sp.</i>	-	Formulation	Sevin (50W)
Pant and Singh	Inducement of metabolic dysfunction by carbamate and organophosphorus compounds in a fish, <i>Puntius conchonius</i> .	1983	Barb, <i>Puntius conchonius</i>	-	Not North American species; Formulation	Sevin (50% WP)
Patil et al.	Toxicity of carbamate insecticides to freshwater crab <i>Paratelphusa jacquemontii</i> (Rathbun).	1992	Crab, <i>Paratelphusa jacquemontii</i>	-	Not North American species; Formulation	Sevimol
Patnaik and Patra	Haematopoietic alterations induced by carbaryl in <i>Clarias batrachus</i> (Linn.).	2006	Walking catfish, <i>Clarias batrachus</i>	96 hr LC50=15,300	Lack of exposure details	Dilution water not characterized
Pauli et al.	RATL: A database of reptile and amphibian toxicology literature.	2000	-	-	Review of previous studies	
Pelletier et al.	Symposium-in-print: UV effects on aquatic and coastal ecosystems. Ecotoxicological effects of combined UVB and organic contaminants in coastal waters: A review.	2006	-	-	Mixture	Carbaryl, atrazine, and acifluorfen
Perry	Pesticide and PCB residues in the Upper Snake River ecosystem, southeastern Idaho, following the collapse of the Teton Dam 1976.	1979	-	-	Mixture	

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Author	Title	Date	Organism(s)	Concentration (µg/L)	Reason Unused	Expanded Reason
Pesando et al.	Biological targets of neurotoxic pesticides analyzed by alteration of developmental events in the Mediterranean sea urchin, <i>Paracentrotus lividus</i> .	2003	Mediterranean sea urchin, <i>Paracentrotus lividus</i>	-	Not North American species; Lack of details (procedure)	
Pesch and Hoffman	Adaption of the polychaete <i>Neanthes arenaceodentata</i> to copper.	1982	Polychaete, <i>Neanthes arenaceodentata</i>	-	Not applicable	No carbaryl toxicity information
Peterson et al.	Effect of varying pesticide exposure duration and concentration on the toxicity of carbaryl to two field-collected stream invertebrates, <i>Calineuria californica</i> (Plecoptera: Perlidae) and <i>Cinygma sp.</i> (Ephemeroptera: Heptageniidae).	2001a	Stonefly, <i>Calineuria californica</i> Mayfly, <i>Cinygma sp.</i>	-	Formulation	Clean Crop® (43% carbaryl)
Peterson et al.	A test system to evaluate the susceptibility of Oregon, USA, native stream invertebrates to triclopyr and carbaryl.	2001b	Stonefly, <i>Calineuria californica</i> Mayfly, <i>Cinygma sp.</i> Caddisfly, <i>Brachycentrus americanus</i> Caddisfly, <i>Lepidostoma unicolor</i> Caddisfly, <i>Psychoglypha sp.</i> Mayfly, <i>Ameletus sp.</i>	-	Formulation	Clean Crop® (43% carbaryl)
Pozarycki	Sublethal effects of estuarine carbaryl applications on juvenile English sole (<i>Pleuronectes vetulus</i>).	1999	English sole, <i>Pleuronectes vetulus</i>	-	Mixture	
Prescott et al.	The effects of pesticides, polychlorinated biphenyls and metals on the growth and reproduction of <i>Acanthamoeba castellanii</i> .	1977	Amoeba, <i>Acanthamoeba castellanii</i>	6 d decreased growth at 10,000	Only one exposure concentration	
Pridgeon et al.	Susceptibility of <i>Aedes aegypti</i> , <i>Culex quinquefasciatus</i> Say and <i>Anopheles quadrimaculatus</i> Say to 19 pesticides with different modes of action.	2008	Mosquito, <i>Aedes aegypti</i> Mosquito, <i>Culex quinquefasciatus</i> Mosquito, <i>Anopheles quadrimaculatus</i>	-	Topically applied contaminant	
Puglis and Boone	Effects of fertilizer, an insecticide, and a pathogenic fungus on hatching and survival of bullfrog (<i>Rana catesbeiana</i>) tadpoles.	2007	Bullfrog, <i>Rana catesbeiana</i>	-	Formulation	Sevin (22.5% carbaryl)
Rajendran and Venugopalan	Effect of pesticides on phytoplankton production.	1983	-	-	Mixture	

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Ramachandran et al.	Effect of pesticides on photosynthesis and respiration of marine macrophytes.	1984	Macrophytes	-	Only one exposure concentration	
Ramakrishnan et al.	Sublethal effects of pesticides on feeding energetics in the air breathing fish <i>Channa striatus</i> .	1997a	Snakehead catfish, <i>Channa striatus</i>	-	Not North American species; Formulation	Hexavin
Ramakrishnan et al.	Sublethal effects of pesticides on physiological energetics of freshwater fish, <i>Oreochromis mossambicus</i> .	1997b	Mozambique tilapia, <i>Oreochromis mossambicus</i>	-	Not North American species; Formulation	Hexavin
Ramaswami	Adaptive trends in lipid levels of liver and muscle of <i>Sarotherodon mossambicus</i> (Peters) exposed to sevin.	1990	Mozambique tilapia, <i>Oreochromis mossambicus</i>	-	Not North American species; Formulation	Sevin (10% dust)
Ramaswamy	Effects of sevin on blood free amino acid levels of the fish <i>Sarotherodon mossambicus</i> .	1987	Mozambique tilapia, <i>Oreochromis mossambicus</i>	-	Not North American species; Formulation	Sevin
Ramaswamy and Maheswari	Comparative lactic acidosis in fishes following pesticide stress.	1993	Mozambique tilapia, <i>Oreochromis mossambicus</i>	-	Not North American species; Formulation	Sevin (10% dust)
Ramaswamy et al.	Glutamic oxaloacetic transaminase (GOT) and glutamic pyruvic transaminase (GPT) enzyme activities in different tissues of <i>Sarotherodon mossambicus</i> (Peters) exposed to a carbamate pesticide, carbaryl.	1999	Mozambique tilapia, <i>Oreochromis mossambicus</i>	-	Not North American species; Formulation	Commercial grade (100g/kg dust)
Rao	Effect of γ -hexachloran and sevin on the survival of the Black Sea mussel, <i>Mytilus galloprovincialis</i> Lam.	1981	Mussel, <i>Mytilus galloprovincialis</i>	96 hr LC50=>10,000	Not North American species	
Rao	Variations in the nitrogen products of <i>Channa punctatus</i> augmented by interaction of carbaryl and phenthoate in the media.	1987	Snakehead catfish, <i>Channa punctatus</i>	48 hr decreased level of ammonia and urea at 3,000	Not North American species; Only one exposure concentration; Lack of exposure details	Dilution water not characterized
Rao and Kannupandi	Acute toxicity of three pesticides and their effect on the behavior of the edible crab <i>Scylla serrata</i> (Forsk.)	1990	Crab, <i>Scylla serrata</i>	96 hr LC50=466	Not North American species	
Rao and Rao	Independent and combined action of carbaryl and phenthoate on snake head, <i>Channa punctatus</i> (Bloch).	1987	Snakehead catfish, <i>Channa punctatus</i>	-	Not North American species; Lack of exposure details	
Rao et al.	Relative toxicity of technical grade and formulated carbaryl and 1-naphthol to, and carbaryl-induced biochemical changes in the fish <i>Cirrhinus mrigala</i> .	1984a	Carp, <i>Cirrhinus mrigala</i>	96 hr LC50=2,500	Not North American species	
Rao et al.	Differential action of malathion, carbaryl and BHC on acetylcholinesterase activity of a teleost, <i>Tilapia mossambica</i> (Peters).	1984b	Mozambique tilapia, <i>Oreochromis mossambicus</i>	-	Formulation	Carbaryl (50% EC)

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Rao et al.	Combined action of carbaryl and phenthoate on a freshwater fish (<i>Channa punctatus</i> Bloch).	1985a	Snakehead catfish, <i>Channa punctatus</i>	48 hr LC50=8,710	Not North American species; Lack of exposure details	Dilution water not characterized
Rao et al.	Combined action of carbaryl and phenthoate on tissue lipid derivatives of murrel, <i>Channa punctatus</i> (Bloch).	1985b	Snakehead catfish, <i>Channa punctatus</i>	48 hr lethal concentration at 8,710	Not North American species; Lack of exposure details	Dilution water not characterized
Rao et al.	Inhibition and recovery of selected target enzyme activities in tissues of penaeid prawn, <i>Metapenaeus monoceros</i> (Fabricius), exposed to different insecticides.	1991	Prawn, <i>Metapenaeus monoceros</i>	48 hr LC50=137	Not North American species	
Ray and Poddar	Carbaryl induced elevation of corticosterone level and cholinergic mechanism.	1983	-	-	Not applicable	No carbaryl toxicity information
Razmi et al.	Persistence of toxicity of some insecticides against the neonate larvae of <i>Leucinodes orbonalis</i> Guen.	1991	Shoot borer, <i>Leucinodes orbonalis</i>	-	Not North American species; Aerial application	
Reddy and Rao	Tissue glycolytic potentials of penaeid prawn, <i>Metapenaeus monoceros</i> during methylparathion, carbaryl and aldrin exposure.	1991a	Prawn, <i>Metapenaeus monoceros</i>	96 hr LC50=120	Not North American species; Lack of exposure details	
Reddy and Rao	Methylparathion, carbaryl and aldrin impact on nitrogen metabolism of prawn, <i>Penaeus indicus</i> .	1991b	Prawn, <i>Penaeus indicus</i>	96 hr LC50=21	Not North American species	
Reddy and Rao	Toxicity of selected insecticides to the penaeid prawn, <i>Metapenaeus monoceros</i> (Fabricius).	1992	Prawn, <i>Metapenaeus monoceros</i>	96 hr LC50=24.3	Not North American species	
Reddy et al.	Recovery of carbaryl inhibited AChE in penaeid prawn, <i>Metapenaeus monoceros</i> .	1990	Prawn, <i>Metapenaeus monoceros</i>	96 hr LC50=24.87	Not North American species; Lack of exposure details	
Regoli et al.	Effects of copper and cadmium on the presence of renal concretions in the bivalve <i>Donacilla cornea</i> .	1992	Bivalve, <i>Donacilla cornea</i> .	-	Not applicable	No carbaryl toxicity information
Relyea	Growth and survival of five amphibian species exposed to combinations of pesticides	2004	Gray tree frog, <i>Hyla versicolor</i> Leopard frog, <i>Rana pipiens</i> Bullfrog, <i>R. catesbeiana</i> Green frog, <i>R. clamitans</i> Bullfrog, <i>Bufo americanus</i>	-	Formulation; Only two exposure concentrations	Commercial grade
Relyea	The impact of insecticides and herbicides on the biodiversity and productivity of aquatic communities.	2005	Various species	-	Formulation	Carbaryl (22.3%)

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Author	Title	Date	Organism(s)	Concentration (µg/L)	Reason Unused	Expanded Reason
Relyea	The effects of pesticides, pH, and predatory stress on amphibians under mesocosm conditions.	2006	Bullfrog, <i>Rana catesbeiana</i> Green frog, <i>R. clamitans</i>	-	Lack of exposure details; Formulation	Dilution water not characterized; Sevin (22% carbaryl)
Relyea	A cocktail of contaminants: How mixtures of pesticides at low concentrations affect aquatic communities.	2009	Gray tree frog, <i>Hyla versicolor</i> Leopard frog, <i>Rana pipiens</i>	~57 d no effect on metamorphosis, growth, and survival at 6.9 (both species)	Only one exposure concentration; High TOC	
Relyea and Mills	Predator-induced stress makes the pesticide carbaryl more deadly to gray treefrog tadpoles (<i>Hyla versicolor</i>).	2001	Gray tree frog, <i>Hyla versicolor</i>	10-16 d 3-4% of LC50 killed 10-60% of frog	Unmeasured chronic exposure	
Rettich	The susceptibility of mosquito larvae to eighteen insecticides in Czechoslovakia.	1977	Mosquitoes, <i>Aedes cantans</i> , <i>A. vexans</i> , <i>A. excrucians</i> , <i>Culex pipiens</i> , <i>A. punctor</i>	24 hr LC50=376.6 LC50=322.6 LC50=145.5 LC50=333 LC50=298.3	Lack of exposure details	Distilled water without proper salts added
Riad et al.	Aromatic sulphides, sulphoxides, and sulphones as larvicides for <i>Culex pipiens molestus</i> and <i>Aedes caspius</i> (Diptera: Culicidae).	1992	Mosquito, <i>Culex pipiens</i> <i>Aedes caspius</i>	-	Lack of exposure details; Excessive solvent used	4 mL/L acetone
Rifaat et al.	Effect of sublethal concentrations of the insecticides DDT, abate and sevin applied to 3rd stage larvae of <i>Anopheles pharoensis</i> on malaria cycle in the adult mosquito.	1974	Mosquito, <i>Anopheles pharoensis</i>	24 hr LC50=600	Lack of exposure details	Dilution water not characterized, purity not given
Rohr et al.	Lethal and sublethal effects of atrazine, carbaryl, endosulfan, and octylphenol on the streamside salamander (<i>Ambystoma barbouri</i>).	2003	Streamside salamander, <i>Ambystoma barbouri</i>	-	Prior exposure	Organisms collected in an agricultural area
Rohr et al.	Understanding the net effects of pesticides on amphibian trematode infections.	2008	Trematode, <i>Echinostoma trivolvis</i> Snail, <i>Planorbella trivolvis</i> Green frog, <i>Rana clamitans</i>	Survival 24 hr NOEC 14 d NOEC 14 d NOEC at 33.5	Only one exposure concentration	
Rossini and Ronco	Acute toxicity bioassay using <i>Daphnia obtusa</i> as a test organism.	1996	Cladoceran, <i>Daphnia obtusa</i>	48 hr EC50=11.5	Not North American species	
Ruber and Baskar	Sensitivities of selected microcrustacea to eight mosquito toxicants.	1968	Mosquitoes	-	Lack of exposure details	Dilution water not characterized
Rybakova	On the toxic effect of sevin on animals.	1966	-	-	Not applicable	No carbaryl toxicity information
Rzehak et al.	The effect of karbatox 75, a carbaryl insecticide, upon the development of tadpoles of <i>Rana temporaria</i> and <i>Xenopus laevis</i> .	1977	Common frog, <i>Rana temporaria</i> African clawed frog, <i>Xenopus laevis</i>	-	Formulation	Karbatox 75

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Sadek et al.	Effect of sublethal concentrations of the insecticides DDT, abate and sevin, applied to 3rd stage larvae of <i>Culex pipiens molestus</i> on certain biological aspects of the mosquito.	1974	Mosquito, <i>Culex pipiens molestus</i>	-	Lack of exposure details; Formulation	Dilution water not characterized
Sahai and Gupta	Residue analysis of some pesticides in the brain of a teleost fish <i>Heteropneustes fossilis</i> (Bloch).	1992	Catfish, <i>Heteropneustes fossilis</i>	-	Not North American species; Only one exposure concentration	
Sahu et al.	Reaction of blue-green algae of rice-field soils to pesticide application.	1992	Blue-green alga	-	Formulation	Carbaryl (50%)
Sakamoto et al.	Inhibition of development of anti-predator morphology in the small cladoceran <i>Bosmina</i> by an insecticide: Impact of an anthropogenic chemical on prey-predator interactions.	2006	Cladoceran, <i>Bosmina fatalis</i>	6d reduced reproduction at 2	Not North American species; Only one exposure concentration	
Sampath and Elango	Lipid metabolism in common frog (<i>Rana tigrina</i>) exposed to carbaryl.	1997	Common frog, <i>Rana tigrina</i>	-	Not North American species; Formulation	Sevin (50% WP)
Sampath et al.	Effect of carbaryl (sevin) on the carbohydrate metabolism of the common frog <i>Rana tigrina</i> .	1992	Common frog, <i>Rana tigrina</i>	-	Not North American species; Formulation	Sevin (50% WDP)
Sampath et al.	Effect of carbaryl on the levels of protein and aminoacids of common frog <i>Rana tigrina</i> .	1995	Common frog, <i>Rana tigrina</i>	-	Not North American species; Formulation; Injected toxicant	Sevin (50% WDP)
Sampath et al.	Pesticide impact on excretory physiology of the common frog, <i>Rana tigrina</i> .	2002	Common frog, <i>Rana tigrina</i>	-	Not North American species; Lack of exposure details	Dilution water not characterized, purity unknown
Sanders	Toxicity of some insecticides to four species of malacostracan crustaceans.	1972	Amphipod, <i>Gammarus fasciatus</i> Sowbug, <i>Asellus brevicaudus</i> Shrimp, <i>Palaemonetes kadinkensis</i> Crayfish, <i>Orconectes nais</i>	96 hr LC50=26 LC50=240 LC50=5.6 LC50=8.6	Excessive solvent used	1 mL/L
Sastry and Siddiqui	Chronic toxic effects of the carbamate pesticide sevin on carbohydrate metabolism in a freshwater snakehead fish, <i>Channa punctatus</i> .	1982	Snakehead, <i>Channa punctatus</i>	60 d caused blood alterations at 1,050	Not North American species; Only one exposure concentration	
Sastry and Siddiqui	Effect of the carbamate pesticide sevin on the intestinal absorption of some nutrients in the teleost fish, <i>Channa punctatus</i> .	1985	Snakehead, <i>Channa punctatus</i>	1 hr decreased rate of absorption of glucose at 10,000	Not North American species	
Sastry et al.	Acute and chronic toxic effects of the carbamate pesticide sevin on some haematological, biochemical and enzymatic parameters in the fresh water teleost fish <i>Channa punctatus</i> .	1988	Snakehead, <i>Channa punctatus</i>	-	Not North American species; Only one exposure concentration	

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Saxena and Aggarwal	Toxicity of some insecticides to the Indian catfish, <i>Heteropneustes fossilis</i> (Bloch).	1970	Catfish, <i>Heteropneustes fossilis</i>	-	Not North American species; Formulation	50% WP
Saxena and Garg	Effect of insecticidal pollution on ovarian recrudescence in the fresh water teleost <i>Channa punctatus</i> (Bloch).	1978	Snakehead catfish, <i>Channa punctatus</i>	-	Not North American species; Formulation	Carbaryl (50% WP)
Saxena et al.	Effects of some pesticides on in-vitro lipid and protein synthesis by the liver of the freshwater teleost, <i>Channa punctatus</i> (Bl.).	1989	Snakehead catfish, <i>Channa punctatus</i>	-	Not North American species; Formulation	Carbaryl (50% WP)
Sayce and Chambers	Observations on potential uptake of sevin by Pacific oysters.	1969	-	-	Lack of details	Dilution water not characterized
Scaps et al.	Biochemical and physiological responses induced by toxics in annelida: utilization as biomarkers.	2002	Polychaete, <i>Nereis diversicolor</i>	-	Only one exposure concentration	
Schacht et al.	Bioassays for risk assessment of coal conversion products.	1999	-	-	Not applicable	No carbaryl toxicity information
Scholz et al.	Dose-additive inhibition of Chinook salmon acetylcholinesterase activity by mixtures of organophosphate and carbamate insecticides.	2006	Chinook salmon, <i>Oncorhynchus tshawytscha</i>	-	Surgically altered test species	Olfactory rosettes removed
Schulz	Field studies on exposure, effects, and risk mitigation of aquatic nonpoint-source insecticide pollution: A review.	2004	-	-	Review of previous studies	
Scott and Georghiou	Malathion-specific resistance in <i>Anopheles stephensi</i> from Pakistan.	1986	Mosquito, <i>Anopheles stephensi</i>	24 hr LC50=720	Not North American species	
Scott and Sloman	The effects of environmental pollutants on complex fish behavior: integrating behavioural and physiological indicators of toxicity.	2004	-	-	Review of previous studies	
Seiffer and Schoof	Tests of 15 experimental molluscicides against <i>Australorbis glabratus</i> .	1967	Snail, <i>Australorbis glabratus</i>	6 hr 100% death at 10,000	Not North American species; Only one exposure concentration	
Selvakumar et al.	Stressor-specific induction of heat shock protein 70 in the freshwater prawn <i>Macrobrachium malcolmsonii</i> (H. Milne Edwards) exposed to the pesticides endosulfan and carbaryl.	2005	Prawn, <i>Macrobrachium malcolmsonii</i>	-	Not North American species; Only two exposure concentrations	
Semlitsch et al.	Genetic variation and a fitness tradeoff in the tolerance of gray treefrog (<i>Hyla versicolor</i>) tadpoles to the insecticide carbaryl.	2000	Green treefrog, <i>Hyla versicolor</i>	-	Only one exposure concentration	
Seuge and Bluzat	Chronic toxicity of carbaryl and lindane to the freshwater mollusc <i>Lymnaea stagnalis</i> L.	1979a	Snail, <i>Lymnaea stagnalis</i>	-	Only two exposure concentrations; Text in foreign language	

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Seuge and Bluzat	Study of the chronic toxicity of two insecticides (carbaryl and lindane) toward the F-sub-1 generation of <i>Lymnaea stagnalis</i> L. (Mollusca, Gasteropoda, Pulmonata). 2. Consequences on the reproductive potential.	1979b	Snail, <i>Lymnaea stagnalis</i>	-	Only two exposure concentrations; Text in foreign language	
Shacklock and Croft	Effect of grazers on <i>Chondrus crispus</i> in culture.	1981	Irish moss, <i>Chondrus crispus</i>	-	Formulation	Sevin (50% carbaryl)
Shaikila et al.	Adaptive trends in tissue acid and alkaline phosphatases of <i>Sarotherodon mossambicus</i> (Peters) under sevin toxicity.	1993	Mozambique tilapia, <i>Oreochromis mossambicus</i>	96 hr increased alkaline phosphatase enzyme activities of liver and muscle at 3,000	Only one exposure concentration	
Shamaan et al.	Insecticide toxicity, glutathione transferases and carboxylesterase activities in the larva of the <i>Aedes</i> mosquito.	1993	Mosquito, <i>Aedes aegypti</i>	-	Dilution water is distilled water without the proper salts added	
Shanmugam et al.	Effect of pesticides on the freshwater crab <i>Barytelphusa cunicularis</i> (West Wood).	2000	Crab, <i>Barytelphusa cunicularis</i>	-	Not North American species; Formulation	50% carbaryl
Shea	Testing of chemical and microbial insecticides for safety...some techniques.	1977	-	-	Formulation	Sevin-4-oil
Sherstneva,	Effect of some pesticides on the fresh water crustaceans.	1978	-	-	Lack of details	Text in foreign language
Shrivastava and Singh	Toxic effect of carbaryl on glucose level in the muscles of <i>Heteropneustes fossilis</i> .	2003	Catfish, <i>Heteropneustes fossilis</i>	30 d decrease in glucose content of muscles at 40	Not North American species; Only one exposure concentration	
Shrivastava and Singh	Changes in protein content in the muscle of <i>Heteropneustes fossilis</i> exposed to carbaryl.	2004	Catfish, <i>Heteropneustes fossilis</i>	30 d decrease in protein of muscles at 40	Not North American species; Only one exposure concentration	
Shrivastava et al.	Study of cholesterol content in muscle of carbaryl exposed <i>Heteropneustes fossilis</i> (Bloch.).	2005	Catfish, <i>Heteropneustes fossilis</i>	30 d decrease in cholesterol content of muscles at 40	Not North American species; Only one exposure concentration	
Shukla and Mishra	Bioassay studies on effects of carbamate insecticides on dragonfly nymphs.	1980	Dragonfly, <i>Brachythermis contaminata</i>	-	Not North American species; Formulation	Carbaryl (WDP 10%)
Shukla and Omkar	Insecticide toxicity to <i>Macrobrachium lamarrei</i> (H. Milne Edwards) (Decapoda, Palaemonidae).	1984	Prawn, <i>Macrobrachium lamarrei</i>	-	Not North American species; Formulation	Sevin (50 WP)
Shukla et al.	Acute toxicity of few pesticides to an aquatic insect, <i>Ranatra elongata</i> (Fabr.).	1982	Water scorpion, <i>Ranatra elongata</i>	-	Not North American species; Formulation	50% WP
Sikka and Rice	Interaction of selected pesticides with marine microorganisms.	1974	-	-	Lack of exposure details	
Sikka et al.	Metabolism of selected pesticides by marine microorganisms.	1973	-	-	Excessive carrier solvent	1% ethanol

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Singh and Agarwal	Toxicity of certain pesticides to two economic species of snails in northern India.	1981	Snail, <i>Lymnaea acuminata</i> <i>Pila globosa</i>	96 hr LC50=4,500 LC50=36,500	Not North American species	
Singh and Agarwal	In vivo and in vitro studies on synergism with anticholinesterase pesticides in the snail <i>Lymnaea acuminata</i> .	1983a	Snail, <i>Lymnaea acuminata</i>	96 hr LC50=4,400	Not North American species	
Singh and Agarwal	Inhibition kinetics of certain organophosphorus and carbamate pesticides on acetylcholinesterase from the snail <i>Lymnaea acuminata</i> .	1983b	Snail, <i>Lymnaea acuminata</i>	-	Not North American species; Surgically altered test species	Nervous tissue around the buccal mass
Singh and Agarwal	Carbamate and organophosphorus pesticides against snails.	1984	Snail, <i>Lymnaea acuminata</i> <i>Pila globosa</i>	96 hr LC50=4,150 LC50=35,500	Not North American species	
Singh and Agarwal	Synergistic effect of sulfoxide with carbaryl on the in vivo acetylcholinesterase activity and carbohydrate metabolism of the snail <i>Lymnaea acuminata</i> .	1986a	Snail, <i>Lymnaea acuminata</i>	-	Not North American species; Mixture	
Singh and Agarwal	Toxicity of pesticides to fecundity, hatchability and survival of young snails of <i>Lymnaea acuminata</i> .	1986b	Snail, <i>Lymnaea acuminata</i>	48 hr LOEC(fecundity) =11,000 LOEC (hatchability) =1,000	Not North American species	
Singh and Agarwal	Toxicity of piperonyl butoxide - carbaryl synergism on the snail <i>Lymnaea acuminata</i> .	1989	Snail, <i>Lymnaea acuminata</i>	-	Not North American species; Only two exposure concentrations	
Singh and Shrivastava	Histopathological changes in the liver of the fish <i>Nandus nandus</i> exposed to endosulfan and carbaryl.	1998	Fish, <i>Nandus nandus</i>	-	Not North American species; Only one exposure concentration	
Singh et al.	Evaluation of acute toxicity of carbaryl and malathion to freshwater teleosts, <i>Channa punctatus</i> (Bloch) and <i>Heteropneustes fossilis</i> (Bloch).	1984	Snakehead catfish, <i>Channa punctatus</i> Catfish, <i>Heteropneustes fossilis</i>	-	Not North American species; Formulation	50% WP
Singh et al.	Toxicity of malathion and carbaryl pesticides: effects on some biochemical profiles of the freshwater fish <i>Colisa fasciatus</i> .	2004	Gourami, <i>Colisa fasciatus</i>	96 hr LC50=8,000	Not North American species	
Sinha et al.	Thiodicarb, an effective molluscicide for grazer snails of blue green algae.	1986a	Snails	-	Formulation	Carbaryl 50 WP
Sinha et al.	An effective molluscicide for grazer snails of blue green algae.	1986b	Snails	-	Formulation	Carbaryl 50 WP
Sinha et al.	Carbaryl-induced thyroid dysfunction in the freshwater catfish <i>Clarias batrachus</i> .	1991a	Walking catfish, <i>Clarias batrachus</i>	96 hr decreased thyroxin levels at 12,000	Only two exposure concentrations	
Sinha et al.	Pesticides induced changes in circulating thyroid hormones in the freshwater catfish <i>Clarias batrachus</i> .	1991b	Walking catfish, <i>Clarias batrachus</i>	-	Formulation	50% carbaryl

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Author	Title	Date	Organism(s)	Concentration (µg/L)	Reason Unused	Expanded Reason
Sinha et al.	Effect of pesticides on extrathyroidal conversion of T ₄ to T ₃ in the freshwater catfish <i>Clarias batrachus</i> .	1993	Walking catfish, <i>Clarias batrachus</i>	-	Only one exposure concentration	
Smith and Grigoropoulos	Toxic effects of odorous trace organics.	1968	Trout Red shiner	-	Formulation; No scientific names given	Sevin (50% carbaryl)
Smulders et al.	A noncompetitive, sequential mechanism for inhibition of rat α4β2 neuronal nicotinic acetylcholine receptors by carbamate pesticides.	2004	-	-	In vitro test	
Solomon	The teratogenic effects of the insecticides DDT, carbaryl, malathion and parathion on developing medaka eggs (<i>Oryzias latipes</i>).	1978	Medaka, <i>Oryzias latipes</i>	96 hr 10,000 negatively affected embryonic development	Not North American species	
Solomon and Weis	Abnormal circulatory development in medaka caused by the insecticides carbaryl, malathion and parathion.	1979	Medaka, <i>Oryzias latipes</i>	ED50=2,500 (circulatory anomalies)	Not North American species	
Somnuek et al.	Gene expression of acetylcholinesterase in hybrid catfish (<i>Clarias gariepinus</i> x <i>Clarias macrocephalus</i>) exposed to chlorpyrifos and carbaryl.	2009	Hybrid catfish, <i>Clarias gariepinus</i> x <i>C. macrocephalus</i>	24 hr gene expression NOEC=8,652	Not North American species	
Srivastava and Pandey	Effect of carbaryl on <i>Chironomus</i> larvae (Chironomidae).	2007	Midge, <i>Chironomus</i> sp.	-	Lack of exposure details; Formulation	
Statham and Lech	Potential of the acute toxicity of several pesticides and herbicides in trout by carbaryl.	1975a	Rainbow trout, <i>Oncorhynchus mykiss</i>	-	Mixture	
Statham and Lech	Synergism of the acute toxic effects of 3,4-D butyl ester, dieldrin, rotenone, and pentachlorophenol in rainbow trout by carbaryl.	1975b	Rainbow trout, <i>Oncorhynchus mykiss</i>	-	Mixture	
Statham and Lech	Studies on the mechanism of potentiation of the acute toxicity of 2,4-D n-butyl ester and 2',5-dichloro-4'-nitrosalicylanilide in rainbow trout by carbaryl.	1976	Rainbow trout, <i>Oncorhynchus mykiss</i>	-	Mixture	
Strickman	Aquatic bioassay of 11 pesticides using larvae of the mosquito, <i>Wyeomyia smithii</i> (Diptera: Culicidae).	1985	Mosquito, <i>Wyeomyia smithii</i>	7d survival LOEC=1,000	Excessive solvent used	3.3 mL/L
Sukumar and Rao	Toxicity of γ-HCH, methyl parathion and carbaryl to two varieties of a tropical freshwater gastropod, <i>Bellamyia bengalensis</i> (Lamarck) (Gastropoda: Viviparidae).	1985	Gastropod, <i>Bellamyia bengalensis</i>	-	Not North American species; Formulation	Sevin
Sundaram and Szeto	Distribution and persistence of carbaryl in some terrestrial and aquatic components of a forest environment.	1987	Brook trout, <i>Salvelinus fontinalis</i> Slimy sculpin, <i>Cottus cognatus</i>	-	Formulation	Sevin-2-oil

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Author	Title	Date	Organism(s)	Concentration (µg/L)	Reason Unused	Expanded Reason
Suseela et al.	Toxic effects of pesticides on survival and proximate composition of <i>Tubifex tubifex</i> .	1994	Tubificid worm, <i>Tubifex tubifex</i>	-	Formulation	Sevin
Suwanchaichinda and Brattsten	Effects of exposure to pesticides on carbaryl toxicity and cytochrome P450 activities in <i>Aedes albopictus</i> larvae (Diptera: Culicidae).	2001	Mosquito, <i>Aedes albopictus</i>	24 hr LC50=830	Inappropriate dilution water	Distilled water without proper salts added
Suwanchaichinda and Brattsten	Induction of microsomal cytochrome P450s by tire-leachate compounds, habitat components of <i>Aedes albopictus</i> mosquito larvae.	2002	Mosquito, <i>Aedes albopictus</i>	-	Mixture; Inappropriate dilution water	Pure water without proper salts added
Swanson et al.	Testing for pesticide toxicity to aquatic plants: Recommendations for test species.	1991	-	-	Review of previous studies	
Takahashi and Yasutomi	Insecticidal resistance of <i>Culex tritaeniorhynchus</i> (Diptera: Culicidae) in Japan: Genetics and mechanisms of resistance to organophosphorus insecticides.	1987	Mosquito, <i>Culex tritaeniorhynchus</i>	24 hr LC50=268	Not North American species; Lack of exposure details	Dilution water not characterized
Tegelberg and Magoon	Sevin treatment of a subtidal oyster bed in Grays Harbor.	1969	-	-	Lack of exposure details; Formulation	Dilution water not characterized
Tejada et al.	Toxicity of pesticides to target and non-target fauna of the lowland rice ecosystem.	1994	Various species	-	Formulation	Sevin
Thakur and Sahai	Toxicity assessment of some commonly used pesticides to three species of fishes.	1994	Snakehead catfish, <i>Channa punctatus</i> <i>C. striatus</i> <i>Garra gotyla gotyla</i>	96 hr LC50=15,000 LC50=17,500 LC50=7,500	Not North American species	
Thakur et al.	Effect of pesticides on N-use efficiency and growth dynamic in rice.	1988	Rice	-	Formulation; Only one exposure concentration	Carbaryl 4G
Tham et al.	Assessment of <i>Clarias batrachus</i> as a source of acetylcholinesterase (AChE) for the detection of insecticides.	2009	Walking catfish, <i>Clarias batrachus</i>	-	Surgically altered test species	Brain extract
Thomas and Murthy	Acid phosphatase activity in a fresh-water air breathing fish <i>Heteropneustes fossilis</i> and the effect of certain organic pesticides on it.	1976	Catfish, <i>Heteropneustes fossilis</i>	-	Not North American species; Injected toxicant	
Tierney et al.	Linuron and carbaryl differentially impair baseline amino acid and bile salt olfactory responses in three salmonids.	2007	Coho salmon, <i>Oncorhynchus kisutch</i> Sockeye salmon, <i>O. nerka</i> Rainbow trout, <i>O. mykiss</i>	-	Surgically altered test species	Olfactory rosette removed
Tilak	Relative toxicity of carbaryl, 1-naphthol, and three formulations of carbaryl to <i>Channa punctata</i> (Bloch).	1982	Snakehead catfish, <i>Channa punctatus</i>	96 hr LC50=5,000	Not North American species	

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Author	Title	Date	Organism(s)	Concentration (µg/L)	Reason Unused	Expanded Reason
Tilak et al.	Toxicity of carbaryl and 1-naphthol to the freshwater fish <i>Labeo rohita</i> .	1980	Rohu, <i>Labeo rohita</i>	96 hr LC50=4,600	Not North American species	
Tilak et al.	Toxicity of carbaryl and 1-naphthol to four species of freshwater fish.	1981	Catfish, <i>Catla catla</i> <i>Anabas testudineus</i> <i>Mystus cavasius</i> <i>Mystus vittatus</i>	96 hr LC50=6,400 LC50=5,500 LC50=4,600 LC50=2,400	Not North American species	
Todd and Leeuwen	Effects of sevin (carbaryl insecticide) on early life stages of zebrafish (<i>Danio rerio</i>).	2002	Zebrafish, <i>Danio rerio</i>	-	Not North American species; Formulation	21.3% carbaryl
Tompkins	Report of the surveillance program conducted in connection with an application of carbaryl (sevin) for the control of gypsy moth on Cape Cod, Massachusetts.	1966	-	-	Field survey; Mixture	
Toor and Kaur	Toxicity of pesticides to the fish, <i>Cyprinus carpio communis</i> Linn.	1974	Carp, <i>Cyprinus carpio</i>	-	Formulation	Carbaryl (50% WP)
Trial	The effects of sevin-4-oil on aquatic insect communities of streams: a continuation of 1976 studies.	1978	-	-	Formulation	Sevin-4-oil
Trial	The effects of sevin-4-oil on aquatic insect communities of streams (1976-1978).	1979	-	-	Formulation	Sevin-4-oil
Trial	The effects of sevin-4-oil on aquatic insect communities of streams (1976-1979).	1980a	-	-	Formulation	Sevin-4-oil
Trial	The effectiveness of unsprayed buffers in lessening the impact of aerial applications of carbaryl on aquatic insects.	1980b	-	-	Formulation	Sevin-4-oil
Trial	The effect of carbaryl on leaf litter processing in Maine streams.	1981	-	-	Formulation; Lack of exposure details (procedure)	
Trial	The effectiveness of upstream refugia for promoting recolonization of plecoptera killed by exposure to carbaryl.	1982	-	-	Formulation	Sevin-4-oil
Trial and Gibbs	Effects of orthene®, sevin 4 oil® and dylox® on aquatic insects incidental to attempts to control spruce budworm in Maine, 1976.	1978	-	-	Formulation	Sevin-4-oil
Tripathi and Agarwal	Synergism in tertiary mixtures of pesticides.	1997	Snail, <i>Lymnaea acuminata</i>	-	Not North American species; Mixture	Sevin and Decis
Tripathi and Singh	Toxic effects of dimethoate and carbaryl pesticides on carbohydrate metabolism of freshwater snail <i>Lymnaea acuminata</i> .	2002	Snail, <i>Lymnaea acuminata</i>	96 hr decrease hepatopancreas glycogen at 3,000	Not North American species	

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Author	Title	Date	Organism(s)	Concentration (µg/L)	Reason Unused	Expanded Reason
Tripathi and Singh	Toxic effects of dimethoate and carbaryl pesticides on protein metabolism of freshwater snail <i>Lymnaea acuminata</i> .	2003a	Snail, <i>Lymnaea acuminata</i>	96 hr decrease hepatopancreas total protein at 3,000	Not North American species	
Tripathi and Singh	Toxic effects of dimethoate and carbaryl pesticides on reproduction and related enzymes of freshwater snail <i>Lymnaea acuminata</i> .	2003b	Snail, <i>Lymnaea acuminata</i>	-	Not North American species	
Tripathi and Singh	Carbaryl induced alterations in the reproduction and metabolism of freshwater snail <i>Lymnaea acuminata</i> .	2004	Snail, <i>Lymnaea acuminata</i>	96 hr decrease number of eggs at 2,000	Not North American species	
Tsuge et al.	Uptake of pesticides from aquarium tank water by aquatic organisms.	1980	Guppy, <i>Lebistes reticulatus</i> Red snail, <i>Indoplanorbis exustus</i> Mosquito, <i>Culex pipiens</i> Cladoceran, <i>Daphnia pulex</i>	-	Lack of exposure details	Text in foreign language
Upadhyay and Upadhyay	Development of marked basophilia in the liver of <i>Heteropneustes fossilis</i> by some selected chemicals.	1993	Catfish, <i>Heteropneustes fossilis</i>	-	Not North American species; Only one exposure concentration	
Vaishampayan	Mutagenic activity of alachlor, butachlor and carbaryl to a N2-fixing cyanobacterium <i>Nostoc muscorum</i> .	1985	Cyanobacterium, <i>Nostoc muscorum</i>	-	Sediment present	
Van Hoof	Evaluation of an automatic system for detection of toxic substances in surface water using trout.	1980	Rainbow trout, <i>Oncorhynchus mykiss</i>	-	Only one exposure concentration	
Van Hoof et al.	The evaluation of bacterial biosensors for screening of water pollutants.	1992	-	-	Lack of exposure details; Only two exposure concentrations	
Vasseur et al.	Interactions between copper and some carbamates used in phytosanitary treatments.	1988	Bacteria, <i>Photobacterium phosphoreum</i> Protozoa, <i>Colpidium campylum</i>	-	Mixture	Carbaryl and copper
Vasumathi et al.	Acute toxicity of endosulfan, methyl parathion and carbaryl on <i>Macropodus cupanus</i> .	2001	Paradisefish, <i>Macropodus cupanus</i>	96 hr LC50=14,540	Not North American species	
Venkateswaran and Ramaswamy	Lactic acidosis in different of <i>Sarotherdon mossambicus</i> (Peters) exposed to sevin.	1987	Mozambique tilapia, <i>Oreochromis mossambicus</i>	-	Formulation	Sevin (10% dust)
Verma and Gupta	Pesticide in relation to water pollution (accumulation of aldrin and ethyl parathion in the few tissues of <i>Colisa fasciatus</i> and <i>Notopterus notopterus</i>).	1976	Fish, <i>Colisa fasciatus</i> <i>Notopterus notopterus</i>	-	Not applicable	No carbaryl toxicity information

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Author	Title	Date	Organism(s)	Concentration (µg/L)	Reason Unused	Expanded Reason
Verma and Tonk	Biomonitoring of the contamination of water by a sublethal concentration of pesticides - a system analysis approach.	1984	Catfish, <i>Heteropneustes fossilis</i>	-	Not North American species; Formulation	Sevin (50% WP)
Verma et al.	Quantitative estimation of biocide residues in a few tissues of <i>Labeo rohita</i> and <i>Saccobranthus fossilis</i> .	1977	Rohu, <i>Labeo rohita</i> Catfish, <i>Saccobranthus fossilis</i>	-	Not North American species; Lack of exposure details	Dilution water not characterized
Verma et al.	Acute toxicity of twenty three pesticides to a fresh water teleost, <i>Saccobranthus fossilis</i> .	1979	Catfish, <i>Saccobranthus fossilis</i>	-	Not North American species; Lack of exposure details (procedure)	
Verma et al.	Studies on the accumulation and elimination of three pesticides in the gonads of <i>Notopterus notopterus</i> and <i>Colisa fasciatus</i> .	1981b	Fish, <i>Colisa fasciatus</i> <i>Notopterus notopterus</i>	-	Not North American species; Formulation	Sevin (50% WP)
Verma et al.	Bioassay trials with twenty three pesticides to a fresh water teleost, <i>Saccobranthus fossilis</i> .	1982	Catfish, <i>Saccobranthus fossilis</i>	-	Not North American species; Formulation	Sevin (50% WP)
Verma et al.	Evaluation of an application factor for determining the safe concentration of agricultural and industrial chemicals.	1984	Carp, <i>Cirrhina mrigala</i>	-	Not North American species; Formulation	Sevin (50% WP)
Virk et al.	Histopathological and biochemical changes induced by endrin and carbaryl in the stomach, intestine and liver of <i>Mystus tengara</i> .	1987	Catfish, <i>Mystus tengara</i>	-	Not North American species; Formulation	Carbaryl (50% WP)
von Windeguth et al.	The efficacy of carbaryl, propoxur, abate and methoxychlor as larvicides against field infestations of <i>Aedes aegypti</i> .	1971	Mosquito, <i>Aedes aegypti</i>	-	Formulation	0.5, 1.25 and 2.5% carbaryl
Vonesh and Buck	Pesticide alters oviposition site selection in gray tree frogs.	2007	Gray treefrog, <i>Hyla chrysoscelis</i>	-	Only one exposure concentration; Formulation	Sevin (commercial grade)
Vryzas et al.	Spatial and temporal distribution of pesticide residues in surface waters in northeastern Greece.	2009	-	-	Not applicable	No carbaryl toxicity information
Walsh	The pathology of pesticide poisoning in fish.	1974	Coho salmon, <i>Oncorhynchus kisutch</i> Lake trout, <i>Salvelinus namaycush</i>	30 d decreased relative spleen weight at 1,000	Only one exposure concentration	
Weber et al.	Toxicity of certain insecticides to protozoa.	1982	Protozoa	-	Lack of exposure details (procedure)	
Weis and Mantel	DDT as an accelerator of limb regeneration and molting in fiddler crabs.	1976	Fiddler crab, <i>Uca pugilator</i> <i>U. pugnax</i>	-	Not applicable	No carbaryl toxicity information
Weis and Weis	Retardation of fin regeneration in <i>Fundulus</i> by several insecticides.	1975	Killifish, <i>Fundulus sp.</i>	-	Surgically altered test species	

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Author	Title	Date	Organism(s)	Concentration (µg/L)	Reason Unused	Expanded Reason
Weis and Weis	Optical malformations induced by insecticides in embryos of the Atlantic silverside, <i>Menidia menidia</i> .	1976	Atlantic silverside, <i>Menidia menidia</i>	-	Lack of exposure details (procedure)	
Whitmore and Hodges	In vitro pesticide inhibition of muscle esterases of the mosquitofish, <i>Gambusia affinis</i> .	1978	Mosquito, <i>Gambusia affinis</i>	-	Surgically altered test species	Homogenized muscle tissue
Whitten and Goodnight	Toxicity of some common insecticides to tubificids.	1966	Tubificid worms, <i>Tubifex sp.</i> <i>Limnodrilus sp.</i>	-	Formulation	Sevin (20% EC)
Whyard et al.	Isolation of an esterase conferring insecticide resistance in the mosquito <i>Culex tarsalis</i> .	1994	Mosquito, <i>Culex tarsalis</i>	-	Prior exposure	
Wilder and Stanley	RNA-DNA ratio as an index to growth in salmonid fishes in the laboratory and in streams contaminated by carbaryl.	1983	-	-	Only one exposure concentration; Formulation	Sevin-4-oil
Wood et al.	Carbamate and organophosphate resistance in <i>Culex pipiens</i> L. (Diptera: Culicidae) in southern France and the significance of EST-3A.	1984	Mosquito, <i>Culex pipiens</i>	24 hr LC50=500	Inappropriate dilution water	Deionized water without proper salts added
Worthley and Schott	The comparative effects of Cs and various pollutants on fresh water phytoplankton colonies of <i>Wolffia papulifera</i> Thompson.	1972	Duckweed, <i>Wolffia papulifera</i>	-	Formulation	21.5% carbaryl
Yasutomi et al.	Insecticide-resistance of <i>Anopheles sinensis</i> and <i>Culex tritaeniorhynchus</i> in Saitama Prefecture, Japan.	1986	Mosquito, <i>Anopheles sinensis</i> Mosquito, <i>Culex tritaeniorhynchus</i>	-	Lack of exposure details	Text in foreign language
Yokoyama et al.	Sensitivity of Japanese eel, <i>Anguilla japonica</i> , to 68 kinds of agricultural chemicals.	1988	Japanese eel, <i>Anguilla japonica</i>	-	Lack of exposure details	Text in foreign language
Yoshida and Nishiuchi	Toxicity of pesticides to some water organisms.	1972	-	-	Lack of exposure details	Text in foreign language
Yoshioka et al.	The estimation for toxicity of chemicals on fish by physico-chemical properties.	1986	-	-	Review of previous studies	

Dash indicates not available